

Secondary Invasion and Reinvasion after Russian-Olive Removal and Revegetation

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Russian-olive is a nitrogen-fixing tree invading riparian corridors in western North America. The premise of revegetation after weed removal is that revegetation is required to return native species to a removal site and that revegetation improves site resistance to invasion or reinvasion via competitive exclusion. Therefore, we expected that revegetation would reduce invasive species cover and increase native species cover compared with non-revegetated controls. Native understory species diversity increased with time since removal. We recorded 18.2 native species in 2012, and 28.2 native species in 2016. Out of 22 planted species, 2 did not establish. Diversity in revegetated plots did not differ from unplanted controls, likely because species spread quickly across plot boundaries. Native perennial grass, seeded species, and annual bromes increased over time, while nonnative forbs and native forbs decreased over time. Only invasive perennial grass cover responded to the revegetation treatment with cover much higher in controls compared with revegetated plots (25.7% vs. 7.7%); this was likely a response to a preplanting herbicide treatment. All categories of species diversity except invasive species diversity increased over time. Only 4% of Russian-olive stumps resprouted in the first year of removal, less than 1% resprouted 2 yr after removal. There was no Russian-olive emergence from seed in the removal year, and seed emergence varied exponentially among following years. Seeded native species did not have trouble establishing once adequate spring moisture occurred in the second growing season after Russian-olive removal, indicating that removal did not present substantial obstacles to successful revegetation. Follow-up control of Russian-olive is critical after initial treatment.

Nomenclature: Russian-olive, *Elaeagnus angustifolia* L. **Key words:** Population maintenance, restoration, riparian.

Russian-olive (*Elaeagnus angustifolia* L.) was common as an ornamental plant in the southwestern United States early in the 20th century and was introduced in the 1930s in the U.S. Great Plains for soil conservation (Stannard et al. 2002). Russian-olive produces thousands of oblong fruits per tree late in the fall (Katz and Shafroth 2003). Fruits may be vertebrate or water dispersed, and seeds are long-lived

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(Scianna et al. 2012). Trees begin to produce fruit as early as 5 yr of age and do so reliably around 10 yr of age (Lesica and Miles 1999). Population maintenance includes stump and root sprouting after fire, mechanical injury, or other disturbance (Stannard et al. 2002). Russian-olive is invasive in riparian areas throughout the western United States (Nagler et al. 2011) and has the potential for cascading effects, including changing beaver population dynamics, restructuring watershed food webs, and altering nitrogen cycling in both local terrestrial and aquatic habitats (Lesica and Miles 1999; Mineau et al. 2011; Pearce and Smith 2001).

Riparian corridors contribute significantly to regional native biodiversity (Naiman et al. 1993) and provide many other ecosystem services such as groundwater recharge, regulation of streamflows, irrigation for agricultural uses, and streambank stabilization (Richardson et al. 2007). The diversity and abundance of alien plants have increased in riparian zones throughout the world (Richardson et al. 2007). Riparian populations of Russian-olive reduce

Management Implications

Russian-olive (*Elaeagnus angustifolia* L.) is invasive in riparian areas throughout the western United States. Trees can grow close together, creating a dense canopy that will shade out other trees and understory plants and eventually replace the native components of riparian ecosystems. We tested the effectiveness of a revegetation planting to increase native plant diversity and cover and reduce secondary invasion.

Cutting the Russian-olive trees using a tree shear to ground level, immediately treating the stumps with herbicide, and removing the trees and stacking into burn piles was very effective (96% kill rate). The limited number of stumps that resprouted did so for 2 yr postremoval, so yearly spraying of resprouts was necessary for complete kill. Russian-olive reinvasion via seedling emergence was substantial; therefore, reducing seedbanks may be critical for long-term control. Russian-olive trees do not produce seeds until they are 5 to 10 yr of age, so young Russian-olives do not need to be removed every year, but we recommend inspecting removal sites every other year to control reinvasion. Active revegetation successfully increased native plant diversity, but plantings may need to be in place longer than 5 yr before competitive exclusion of invasive species is apparent.

recreational and agricultural use in addition to threatening native populations of cottonwood (*Populus* spp.) and willow (*Salix* spp.) trees (Lesica and Miles 2001).

Russian-olive tree removal can create a high level of disturbance, reducing existing plant biomass, removing canopy cover, and providing opportunities for other invasive species to colonize (i.e., secondary invasion) as well as creating a new set point for succession to begin (Blanchard and Holmes 2008; Corenblit et al. 2007; Gabler and Siemann 2012; Srivastava and Vellend 2005). After removal of invasive species, secondary invasions often follow (Blanchard and Holmes 2008; Denslow and D'Antonio 2005; Galatowitsch and Richardson 2005; Rinella et al. 2009), and natural successional patterns may change (Blanchard and Holmes 2008; DeMeester and Richter 2010). Legacies from invasive plants may inhibit native plant germination and growth (e.g., Corbin and D'Antonio 2004; Perkins et al. 2016; Zavaleta et al. 2001), which may make postremoval tactics such as revegetation critical for increasing the likelihood of native plant colonization and establishment but may also make revegetation more likely to fail.

Active restoration such as revegetation can increase species diversity (Srivastava and Vellend 2005), preempt niche space, prevent establishment of invasive plant species (Blanchard and Holmes 2008), and assist in recolonization by native plants (Ehrenfeld 2000). However, ruderal species, nonnative species, and/or invasive species are often a problem in revegetation projects (Mulhouse and Galatowitsch 2003; Zedler and Callaway 1999; but see Rinella et al. 2012). Other research on tree invasions has shown that understory species that survive invasions are a drastically

reduced subset of understory species in uninvaded areas (de Abreu and Durigan 2011; Galatowitsch and Richardson 2005; Tererai et al. 2015; Zavaleta et al. 2001); therefore, revegetation may be required to return the full complement of native understory species to a site after tree removal. Revegetating a site with active restoration can be costly (Kimball et al. 2015) with variable success rates. Allowing a site to revegetate passively via colonization from neighboring plant populations or implementing non-revegetation restoration practices (such as grazing removal) can be less costly but may not be as effective (Gornish et al. 2017; Ruwanza et al. 2013). Because the establishment of a native plant community passively brings both aboveground and belowground fauna to the site (Hobbs and Cramer 2008), revegetation may speed the return of ecosystem functions other than plant species diversity (Palmer et al. 1997).

In previous research (Espeland et al. 2014a), we found that after Russian-olive removal, revegetation increased the diversity of desirable native species compared with unplanted controls after 2 yr. Because of the disturbance involved in revegetation (as in Robichaud et al. 2000), the slow-growing, late seral species often planted in revegetation projects (Suding 2011) are unlikely to exclude ruderal weeds (e.g., Espeland and Perkins 2017) and fast-growing invasive species (Galatowitsch and Richardson 2005). Very early in the restoration process, plantings may have low vegetative cover. At this stage, reinvasion of Russian-olive and secondary invasions of other species may be limited by propapressure, not competitive exclusion. establishment, the growth of native species may reach a stage at which native species are able to competitively exclude secondary invasions and reinvasion of Russian-olive (e.g., Levine et al. 2004). The prior publication on this research (Espeland et al. 2014a) focused on the establishment stage of revegetation; 3 yr hence we expect competitive exclusion to begin to drive plant community succession (Richardson et al. 2007) toward native species dominance.

Preventing Russian-olive reinvasion after removal means minimizing stump resprouting (Stannard et al. 2002) and establishment from seed. We expect stump resprout rates to be primarily determined by the effectiveness of the control technique, not by revegetation success, because resprouting is not likely to be limited by competition. Reinvasion of Russian-olive removal sites via seed germination would depend on propagule pressure (i.e., seedbanks and dispersal, as in Blanchard and Holmes 2008) until competitive exclusion comes into play.

In this publication, we report on Russian-olive stand regeneration 5 yr postremoval, and we hypothesize that active revegetation after removal is necessary to reduce secondary invasion and to increase native plant diversity and cover. Our hypotheses were: (1) revegetation increasingly excludes invasive species as the planting matures, compared with non-revegetated controls; (2) revegetation increases

native understory cover and diversity compared with controls; (3) transplanted tree and shrub species vary in survivorship, with some species unable to establish; and (4) resprouting dominates Russian-olive stand regeneration after removal, and the contribution of seeds to reinvasion decreases as seedbanks become depleted. Hypotheses 3 and 4 are evaluated descriptively, whereas experimental controls are available to assess hypotheses 1 and 2 statistically.

Materials and Methods

Study Location. This study was conducted within the mixed-grass prairie of the Northern Great Plains at the 22,500-ha USDA-Agricultural Research Service (USDA-ARS) Fort Keogh Livestock and Range Research Laboratory located near Miles City, MT 46°24.431 N, 105°52.618 W. Native vegetation includes grama-needlegrass-wheatgrass (Bouteloua-Stipa-Agropyron) mix (Barker and Whitman 1988) with shrubs including silver sagebrush (Artemisia cana Pursh ssp. cana), big sagebrush (Artemisia tridentata Nutt.), winterfat [Krascheninnikovia lanata (Pursh) A. Meeuse & Smit, formerly Ceratoides lanata (Pursh) J. T. Howell], and small trees such as common juniper (Juniperus communis L.). Average annual precipitation is 316 mm, of which 85% is received April to October.

Experimental Design. Four replicate 0.5-ha blocks were established adjacent to the Yellowstone River on a Glendive fine sandy loam soil. Blocks were located in closed-canopy Russian-olive forest, equidistant from the edge of the river. Russian-olive trees were completely removed from these blocks in April and May of 2011 using a John Deere 326D skid steer with tree shear and herbicide-spray attachments (Grace Manufacturing, 11200 Turkey Ridge Drive, Plato, MO 65552). Triclopyr (Element® 4, Dow Agrosciences, 9330 Zionsville Road, Indianapolis, IN 46268) was applied at 479 g ai L⁻¹ with an oil carrier (Blue Basal Bark oil, Dow Agrosciences) immediately following a cut-stump treatment. This was the equivalent of a 25% v/v treatment on a product basis. The cost per hectare of removing these 2,500 trees (excluding equipment costs) was 17.7 person-hours, 39.5 L of fuel, and US\$427 in herbicide costs (7.9 L of triclopyr ha⁻¹). Resprouts and new Russian-olive seedlings were spot sprayed with a backpack using a mixture consisting of 2.59 g ai L⁻¹ Element[®] 4, 1.29 g ai L⁻¹ Milestone[®] (aminopyralid, Dow Agrosciences, Canada, Suite 2100, 450 1st Street SW, Calgary, AB T2P 1M4, Canada), and 2.6 g surfactant mixed in water in fall of 2011 and 2012. Spottreatment spray application was also conducted in summer of 2013 to 2016 and included saltcedar (Tamarix ramosissima Ledeb.). Some Russian-olive seedlings were hand pulled in 2014 and 2015; in other years, herbicide use ranged from 1.2 to 8.4 L of the spot-spray treatment, mixed

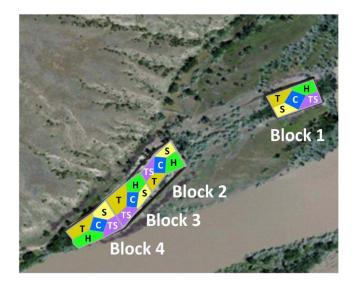


Figure 1. Aerial view of removal plots along the Yellowstone River near Miles City, MT, including block and treatment placement. Treatments were: C, control plots that did not receive any active restoration; H, herbaceous seeds were broadcast seeded in plots; S, herbaceous seeding with shrub transplants; T, herbaceous seeding with shrub and tree transplants; and TS, herbaceous seeding with shrub and tree transplants.

as mentioned above, per hectare. We recorded the number of treated saplings per block.

Each block was divided equally into five revegetation treatments. For logistic reasons, non-revegetated controls were always in the center and other treatments were randomized within blocks (Figure 1). We seeded all revegetation treatments with a herbaceous mix, and trees and shrubs were transplanted in a factorial design. Revegetation occurred in April of 2012, because a 50-yr flood in May 2011 placed most plots underwater until the beginning of July. Local sources supplied 1-yr-old conservation-grade woody stock for the tree and shrub plantings. All plots except controls were sprayed with glyphosate (Roundup PowerMax®, Monsanto, 800 N Lindbergh Boulevard, St Louis, MO 63167) in the fall of 2011 and before revegetation in spring of 2012. Glyphosate use was 0.54 kg ai ha⁻¹ and spraying took 1.6 personhours ha⁻¹. Herbaceous seed was applied to all plots (described below). One treatment ("H") received only this seeding; "T" plots received approximately 38 trees plot⁻¹; "S" plots received approximately 150 shrubs; and "TS" plots received 75 shrubs and 20 trees, with equal numbers of each species planted in each experimental unit. Each tree and shrub transplant received approximately 3.75 L of water upon planting and none thereafter. Weed barrier fabric (0.91 m by 0.91 m) surrounded half the transplanted individuals. Transplanted woody species were narrowleaf cottonwood (Populus angustifolia James), plains cottonwood

[Populus deltoides W. Bartram ex Marshall ssp. monilifera (Aiton) Eckenwalder], boxelder (Acer negundo L.), green ash (Fraxinus pennsylvanica Marshall), golden currant (Ribes aureum Pursh), common chokecherry (Prunus virginiana L.), silver buffaloberry [Shepherdia argentea (Pursh) Nutt.], and Woods' rose (Rosa woodsii Lindl.). Excluding weed fabric, transplanting costs were 10 person-hours and US\$111 in materials per hectare. The tree and shrub treatments were established to examine the potential for facilitation among plant life history groups. Since trees and shrubs were still too small to interact with each other in 2016, we use T, S, and TS plots only to report on survival of transplanted species in this paper. We did not use these plots for the analysis of understory vegetation because of the potential for hidden treatments (as in Huston 1997).

Herbaceous species seeded were: slender wheatgrass [Elymus trachycaulus (Link) Gould ex Shinners], western wheatgrass [Pascopyrum smithii (Rydb.) Á. Löve], Prairie cordgrass (Spartina pectinata Link), switchgrass (Panicum virgatum L.), common yarrow (Achillea millefolium L.), upright prairie coneflower [Ratibida columnifera (Nutt.) Wood & Standl.], American vetch (Vicia americana Muhl. ex Willd.), Canada milkvetch (Astragalus canadensis L.), white prairie clover (Dalea candida Michx. ex Willd.), purple prairie clover (Dalea purpurea Vent.), Maximilian sunflower (Helianthus maximiliani Schrad.), blue flax (Linum perenne L.), Rocky Mountain beeplant (Cleome serrulata Pursh.), and Rocky Mountain penstemon (Penstemon strictus Benth.). Herbaceous seed was broadcast, and harrows and hand rakes ensured seed-soil contact. The total application of all seed was 1.2 kg live seed ha⁻¹ (see Supplementary Material for seed mix fractions) at a cost of 0.7 person-hours and US\$105 in materials per hectare. All plots were fenced to USDA-Natural Resource Conservation Service (USDA-NRCS) wildlife fence (NRCS 2006, 2008) specifications to protect the woody plants from wildlife and cattle browsing. Riparian tree plantings are unlikely to survive without protection from herbivory (Sweeney and Czapka 2004).

Data Collection. In all treatment plots and non-revegetated controls, understory plant cover by functional group (native perennial grass, invasive perennial grass, nonnative forb, forb, invasive annual grass, seeded species) was collected using the line-point intercept method (Jonasson 1988) along two 25-m transects per experimental unit, with 50 evenly spaced data-collection points on each transect. We also recorded the herbaceous species present throughout each growing season in the entire plot. We tracked secondary invasion by recording dynamics of invasive species: black henbane (*Hyoscyamus niger* L.), smooth brome (*Bromus inermis* Leyss.), clasping pepperweed (*Lepidium perfoliatum* L.), halogeton [*Halogeton glomeratus* (Stephen ex Bieb.) C. A. Mey.], houndstongue (*Cynoglossum officinale* L.), nonnative thistles [*Cirsium arvense* (L.) Scop.;

Cirsium vulgare (Savi) Ten.], spotted knapweed (Centaurea stoebe L.), absinth wormwood (Artemisia absinthium L.), Kentucky bluegrass (Poa pratensis L.), crested wheatgrass [Agropyron cristatum (L.) Gaertn.], annual brome grasses (Bromus arvensis L.; Bromus tectorum L.), Siberian elm (Ulmus pumila L.), and saltcedar. We also documented native species that naturally colonized the plots and were not seeded species. We did not have any invasive native species at our site.

All transplanted trees and shrubs were mapped at planting in spring 2012. Each living tree and shrub transplant was individually tagged in spring of 2013. Maps were compared with locations of live plants to determine survivorship over the first winter. Tagged plants were recensused in 2016, and their survival was recorded.

Each summer of 2011 to 2016, Russian-olive saplings were counted in each block and sprayed with herbicide or hand pulled. In 2011 and 2012, resprouts were counted separately from seedlings. No reprouts were observed in 2014 to 2016 western wheatgrass [*Pascopyrum smithii* (Rydb.) A. Love].

Data Analysis. We tested the influence of year, treatment, year by treatment interaction, and block (random factor) on three invasive cover variables (invasive perennial grass, nonnative forbs, and annual bromes) and three native cover variables (native perennial grass, native forbs, and seeded species) in a multivariate analysis of variance (MANOVA; JMP v. 12.1.0, SAS Institute, Cary, NC). When the MANOVA was significant [year F(4,70) =14.486, P < 0.001; treatment F(1,70) = 5.153; P < 0.027, year*treatment F(4,70) = 3.405, P < 0.014], we used protected generalized linear mixed models to analyze the influence of year, treatment, year by treatment interaction, and block (random factor) on each cover group separately. We used the same model statement in a MANOVA for four diversity measures: total, native, invasive, and seeded species diversity. When the MANOVA was significant [year F(4, 30) = 7.3151, P = 0.0003; treatment and treatment*year interaction P > 0.3], we used protected generalized linear mixed models to analyze the influence of year on each diversity group separately. All cover and diversity data were normally distributed (Shapiro-Wilk test W > 0.84), and cover data were square-root transformed for analysis to improve the distribution of residuals. All averages are presented ± 1 SD.

We calculated conditional survival probabilities for individuals from each transplanted tree and shrub species. The probability that an individual of age x survives to age x + 1, P(x), is calculated as:

$$P(x) = \frac{S(x+1) / S(0)}{S(x) / S(0)}$$
 [1]

where S(x) is number of individuals survived to time x, and S(0) is the original number of individuals planted. Probabilities were generated from data pooled across all plots combined.

Table 1. Number of Russian-olive saplings sprayed or hand pulled after removal in 2011, separated by establishment type (resprout or seedling), and average number of saplings treated per block (±1 SD) and percent stand regeneration (out of the 2,500 trees removed).

Year	Number of resprouts	Number of seedlings	Average per block	Regeneration rate
2011	98	0	24.5 (9.04)	3.9%
2012	71	515	128 (204)	24%
2013	No data	238ª	62.5 (95.7)	10%
2014	0	938 ^b	246 (464)	39%
2015		5383°	<u> </u>	215%
2016	_	618 ^d	154 (172)	25%

^a Approximately 200 saltcedar saplings were sprayed, and counts of resprouts and seedlings were combined in 2013.

Results and Discussion

Our removal technique of cutting Russian-olive trees with a tree shear to ground level and immediately spraying the stump with herbicide resulted in a 4% resprout rate the following year (Table 1). No resprouts were detected in the third year after removal and all following years (Table 1). We observed no seedlings emerging in early summer in 2011, the year we conducted tree removal.

Block 1 consistently had the highest Russian-olive sapling numbers establishing from seed, averaging 1,474 trees year⁻¹ (433, 206, 941, 5,383, and 408 for years 2012 to 2016, respectively); Block 3 had the lowest numbers, with an average of 18.5 saplings year⁻¹ (13, 17, 7, 36, and 37 for years 2012 to 2016, respectively). The stand-regeneration rate was lowest in 2011 (the year of removal), which was the only year when resprouts contributed more to stand

regeneration than seedling establishment (Table 1). Stand-regeneration rates varied considerably among years, with estimates ranging from 4% to 215%.

Half or more of cottonwood and boxelder transplants were lost to transplant shock and/or winterkill in the first year (Figure 2). In following years, cottonwood survival remained poor, and compared with other species, cottonwood essentially failed to establish from our plantings. Other species survival remained relatively high and trended upward between 2014 and 2016. Woods' rose was especially susceptible to herbicide-spray drift from follow-up Russian-olive control: 40% of the mortality in 2014 was due to herbicide damage. However, remaining Woods' rose transplants maintained relatively high survival into 2016.

All planted understory species established in the plots, with the exception of American vetch, which was seeded at a very low rate (see Supplementary Material). While the

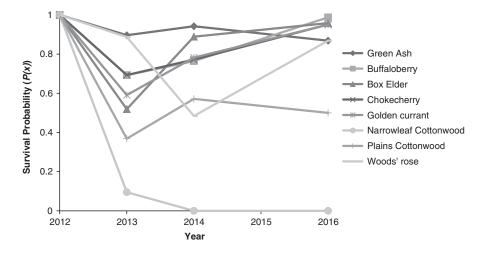


Figure 2. Conditional survival probability to time x + 1, given survival to time x from initial planting (2012) to 2016. Several species had substantial mortality the first year after planting. However, survival has largely increased over time for the remaining transplants, with the exception of narrowleaf cottonwood.

^b Approximately 100 saltcedar saplings were also sprayed in 2014.

^c Only Block 1 was treated in 2015.

^d The 2016 data include saplings emerged in 2015 and 2016 for Blocks 2–4.

interaction term for revegetation over time (treatment*year) was significant in the MANOVA, it was not significant for any single cover group, which indicates that the community as a whole had a nuanced response to revegetation. All plant cover groups responded to the main effect of year. Native perennial grass [F(1,67) = 9.8885, P < 0.0001], seeded species [F(1,67) = 8.6743, P < 0.0001], and annual bromes [F(1,67) = 9.4877, P < 0.0001] increased over time (Figure 3A), while nonnative forbs (despite an initial increase in 2013) [F(1,67) = 13.7476, P < 0.0001] and native forbs [F(1,67) = 4.0611, P < 0.006] decreased over time (Figure 3B). Invasive perennial grass cover decreased initially in 2013 and continued to increase over time [Figure 3C, F(1,67) = 3.884, P < 0.007]. Only invasive perennial grass cover responded to the revegetation

treatment [F(1, 67) = 12.230, P < 0.001] with cover much higher in controls compared with revegetated plots $(25.7 \pm 21.2\% \text{ vs. } 7.7 \pm 13.4\%)$. All categories of species diversity except invasive species diversity (P > 0.06) increased over time (P < 0.004, Figure 3D). Contrary to our expectations, revegetation did not appear to competitively exclude secondary invasions, even after five growing seasons. The establishment phase of secondary invasion appeared to be unaffected by revegetation: the number of invasive species did not differ between planted plots and unplanted controls. However, in terms of growth (Figure 3C), the cover of invasive grass species was lower in revegetated plots compared with non-revegetated controls. Because native cover was not significantly higher in revegetation provided

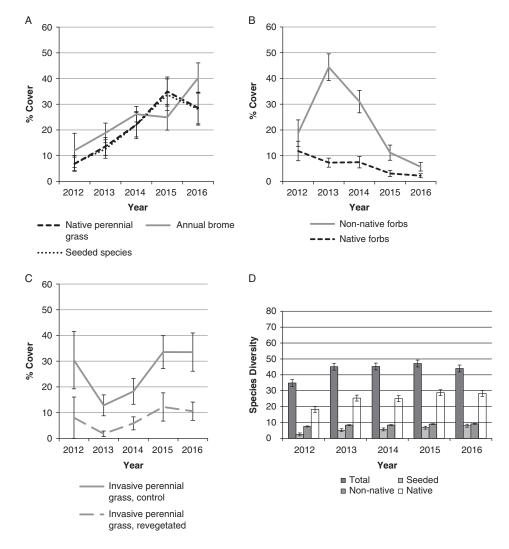


Figure 3. Cover and diversity response to time: (A) native perennial grass, annual bromes, and seeded species increased over time (P < 0.05); (B) nonnative forbs and native forbs decreased over time; (C) invasive perennial grass cover was greater in non-revegetated controls, and increased over time in both revegetated and control plants (P < 0.05); (D) total, seeded, and native species diversity increased over time (P < 0.05), while nonnative species diversity did not change (P < 0.06). Note that D is on a logarithmic scale.

competitive exclusion of invasive grasses. The herbicide treatment as part of revegetation may be responsible for the reduction in invasive grass cover, although the strength of the treatment effect could be a combination of herbicide application, seeded species establishment (i.e., revegetation), and subsequent competitive pressure. Few restoration experiments extend monitoring to the stage at which competitive interactions can be observed (Blanchard and Holmes 2008; Rinella et al. 2012), and we have not observed competitive exclusion of secondary invasive species in this study. Revisiting revegetation plantings many years after their installation shows that successful revegetation establishment does not always confer the function of competitive exclusion of weeds (Rinella et al. 2012). Because invasive grasses in the Northern Great Plains reduce overall native plant species diversity (Toledo et al. 2014), reduce wildlife habitat (Ellis-Felege et al. 2013), and have the potential to prohibit shrub establishment (Rinella et al. 2015), the secondary invasion we observed may represent as significant a threat to native riparian ecosystems as Russian-olive.

Russian-olive removal is a long-term commitment: stands are capable of complete regeneration from seed, and resprouts must be controlled for at least two consecutive years postremoval with the method we used. Removal with a skid steer resulted in soil disturbance that was combined with a 50-yr flood just after removal; these events may have altered both Russian-olive resprout rates and the composition of the understory vegetation compared with less disturbance (Richardson et al. 2007). The first year after removal, a 4% resprout rate included resprouts from tree roots that were exposed by floodwater action. This percentage decreases to 0.4% if we report only stump resprouts, which likely would have been the case had the flood not occurred. Smaller trees are more likely to be killed by herbicide, and spraying larger trees has a greater likelihood of damaging adjacent, desirable vegetation due to spray drift. Nonfoliar treatment techniques such as basal bark herbicide application may avoid these problems but were not tested. Biological control of this tree (potential agents listed in Bean et al. 2008) will likely lead to different limitations to revegetation and reinvasion, just as would be the case if control was achieved from methods other than those used in this experiment. No matter the technique, effective control of reinvasion can have costs to desirable vegetation if appropriate care is not exercised (Blanchard and Holmes 2008).

Our plot design had important effects on our results. Because it is surrounded by untreated land, Block 1 has more opportunities for gravity-based seed inputs from surrounding Russian-olive trees compared with the other experimental blocks. Also, water-dispersed fruits were more likely to settle at this block due to geomorphological factors: this block experienced a river backflow due to an ice jam in March of 2014. Stand regeneration was consistently highest in Block 1. The average 18.4% per year regeneration rate we

observed in Block 3 is more likely to reflect reinvasion dynamics when Russian-olive control is successfully undertaken at the stand level and when water and wildlife dispersal are low. While Russian-olive seed is very long-lived in laboratory storage (Scianna et al. 2012), seed buried deeper than 5 cm rarely emerges and can experience 100% mortality (Hybner and Espeland 2014), which is promising for Russian-olive control in depositional environments such as on the banks of the undammed Yellowstone River (e.g., Graumlich et al. 2002). Because Russian-olive trees do not consistently set fruit every year until they reach about 10 yr of age (Lesica and Miles 1999), it is possible to control reinvasions by conducting reentry kills less frequently than once per year. High stand-regeneration rates throughout the study period illustrate the importance of repeated site inspection and periodic control of this invasive species.

Transplanted tree and shrub survivorship was high, even though southeastern Montana experienced in 2012 one of the four driest years on record since 1878 (National Climatic Data Center, Asheville, NC; Table 2). Low cottonwood survival from our transplants does not indicate the inability of cottonwood to grow in soils with a Russianolive legacy: natural establishment after the flood led to considerable numbers of plains cottonwood seedlings in the experiment area, particularly in the control plots that did not receive herbicide or the additional disturbance of seeding and transplanting in 2012. Cottonwood typically has low seedling survivorship alongside unregulated rivers (Andersen 2005; Bradley and Smith 1986; Horton and Clark 2001; Li Kui and Stella 2016), but due to prolific germination and emergence, this species is able to establish new populations even with very low survival. Because establishment of native cottonwood and willow trees is limited by shade, Russian-olive prevents their recruitment (Lesica and Miles 2001); therefore, these native trees are unlikely to maintain their populations without intervention in the face of continuing Russian-olive invasion.

Placing non-revegetated plots at the center of each block maximized the potential for seeded species to colonize this

Table 2. Percent understory plant cover and growing-season rainfall amounts per year of the study.

Year	Understory cover	SD	April–July rainfall ^a	August–October rainfall
	%		mm	mm
2012	23.0	19.8	78	34
2013	86.6	14.9	302	108
2014	96.4	6.6	208	75
2015	88.0	8.4	149	50
2016	96.0	4.0	224	115

^a April–July rainfall is shown to indicate rainfall that occurred before the cover measurement taken that year.

neighboring unseeded area, which may be partly responsible for the lack of effect our revegetation treatment had on native species' cover and diversity. The disturbance of tree removal that created substantial bare ground, the flood, and subsequent drought were likely the largest drivers of changes in understory plant cover in the short term (as in Richardson et al. 2007). Additionally, at the time of study completion, seeded species had moved beyond the boundaries of the plots. Data analyses conducted in 2013 showed a main effect of revegetation on native species diversity (Espeland et al. 2014a), but the longer time frame of the study we report on here has allowed plot boundaries to blur. Thus, our study may unintentionally support the use of "seed islands"—seeding small areas to provide a source for native species to recolonize degraded lands—in restoration to improve landscapes (as in Reever Morghan et al. 2005).

Removing Russian-olive in dense, closed-canopy populations with a skid steer resulted in considerable bare ground. This disturbance may be responsible for the "year" main effect we observed in plant cover. Both native and introduced forbs appeared to capitalize on the disturbance with high cover early in the experiment; perennial grasses increased their cover over time following disturbance. While many studies show that restoration can introduce high levels of problem species (summarized in Robichaud et al. 2000), all our experimental plots experienced very high levels of disturbance, equalizing that particular invasion vector among controls and treatments. Conducting Russian-olive removal using a technique resulting in less soil disturbance (e.g., with a chainsaw instead of a skid steer) would probably result in very different restoration and reinvasion outcomes.

Ecological research in habitats with high levels of historical and contemporary disturbance means natural variation can occlude the effects of experimental treatments (Richardson et al. 2007; Sweeney and Czapka 2004). We were fortunate that none of our plots were destroyed via natural fluvial processes, and even though large blocks of ice were found inside the wildlife fence in 1 yr, we did not lose any plots to scouring (as in Espeland et al. 2014b). Because stand-regeneration (or reinvasion) numbers varied exponentially among blocks, future ecological research on Russian-olive removal should include not only controlling for tree density and distance from the river (as we did in this study) but also seedbank (Blanchard and Holmes 2008) and dispersal factors that include fluvial geomorphology (Nagler et al. 2011) and wildlife corridors (Stannard et al. 2002). Because seeds are so plentiful under closed-canopy populations, a seedbank-reduction technique such as scraping (e.g., Morgan 1999) or seed-eating biological control (Bean et al. 2008) may be beneficial, or other techniques could be developed to exploit the low survival of buried seed (Hybner and Espeland 2014). This research shows the potential to regain native plant diversity and structure (i.e., trees and shrubs) in riparian areas after the removal of invasive trees.

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Supplementary material

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Literature Cited

Andersen DC (2005) Characterizing flow regimes for floodplain forest conservation: an assessment of factors affecting sapling growth and survivorship on three cold desert rivers. Can J Forest Res 35:2886–2899 Barker WT, Whitman WC (1988) Vegetation of the northern Great Plains. Rangelands 10:266–272

Bean D, Norton A, Jashenko R, Schaffner U (2008) Status of Russian olive biological control in North America. Ecol Restor 26:105–107

Blanchard R, Holmes PM (2008) Riparian vegetation recovery after invasive alien tree clearance in the Fynbos Biome. S Afr J Bot 74:421–431

Bradley CE, Smith DG (1986) Plains cottonwood recruitment and survival on a prairie meandering river floodplain, Milk River, southern Alberta and northern Montana. Can J Bot 64:1433–1442

Corbin JD, D'Antonio CM (2004) Effects of exotic species on soil nitrogen cycling: implications for restoration 1. Weed Technol 18:1464–1467

Corenblit D, Tabacchi E, Steiger J, Gurnell AM (2007) Reciprocal interactions and adjustments between fluvial landforms and vegetation dynamics in river corridors: a review of complementary approaches. Earth-Sci Rev 84:56–86

de Abreu RC, Durigan G (2011) Changes in the plant community of a Brazilian grassland savannah after 22 years of invasion by *Pinus elliottii* Engelm. Plant Ecol Divers 4:269–278

DeMeester JD, Richter D deB (2010) Restoring restoration: removal of the invasive plant *Microstegium vimineum* from a North Carolina wetland. Biol Invasions 12:781–793

Denslow JS, D'Antonio CM (2005) After biocontrol: assessing indirect effects of insect releases. Biol Control 35:307–318

Ellis-Felege SN, Dixon CS, Wilson SD (2013) Impacts and management of invasive cool-season grasses in the Northern Great Plains: challenges and opportunities for wildlife. Wildlife Soc B 37:510–516

- Ehrenfeld JG (2000) Defining the limits of restoration: setting realistic goals. Restor Ecol 8:2–9
- Espeland EK, Perkins L (2017) Weed establishment and persistence after water pipeline restoration and reclamation in the mixed grass prairie of western North Dakota. Ecol Restor 35:303–310
- Espeland EK, Petersen M, Muscha J, Scianna J, Kilian R (2014a) Revegetation after Russian olive (*Eleaeagnus angustifolia* L.) removal along the Yellowstone River: a cost and 2-year success assessment. Pages 13–22 *in* Proceedings of the 3rd Northern Rockies Invasive Plants Council Conference. Airway Heights, WA: USDA Forest Service
- Espeland EK, Rand TA, Delaney KJ (2014b) Russian olive fruit production in shelterbelt and riparian populations in Montana. Ecol Restor 32:354–357
- Gabler CA, Siemann E (2012) Environmental variability and ontogenetic niche shifts in exotic plants may govern reinvasion pressure in restorations of invaded ecosystems. Restor Ecol 20:545–550
- Galatowitsch S, Richardson DM (2005) Riparian scrub recovery after clearing of invasive alien trees in headwater streams of the Western Cape, South Africa. Biol Conserv 122:509–521
- Gornish ES, Lennox MS, Lewis D, Tate KW, Jackson RD (2017) Comparing herbaceous plant communities in active and passive riparian restoration. PLoS ONE 12:e0176338
- Graumlich LJ, Pisaric MFJ, Waggoner LA, Littell JS, King JC (2002). Upper Yellowstone River Flow Reconstruction, International Tree-Ring Data Bank. IGBP PAGES/World Data Center for Paleoclimatology Data Contribution Series #2002-074. https://gcmd.nasa.gov/records/GCMD_ NOAA_NCDC_PALEO_2002-074.html. Accessed: July 17, 2017
- Hobbs RJ, Cramer VA (2008) Restoration ecology: interventionist approaches in restoration and maintaining ecosystem function in the fact of rapid environmental change. Annu Rev Env Resour 33:39–61
- Horton JL, Clark JL (2001) Water table decline alters growth and survival of Salix gooddingii and Tamarix chinensis seedlings. Forest Ecol Manag 140:239–247
- Huston MA (1997) Hidden treatments in ecological experiments: re-evaluating the ecosystem function of biodiversity. Oecologia 110:449–460
- Hybner R, Espeland EK (2014). Russian Olive *Elaeagnus angustifolia* L. Effect of Seed Burial Depth on Seedling Emergence and Seed Viability. USDA Natural Resources Conservation Service Plant Materials Technical Note No. MT-107. 6 p
- Jonasson S (1988) Evaluation of the point intercept method for the estimation of plant biomass. Oikos 52:101–106
- Katz GL, Shafroth PB (2003) Biology, ecology and management of Elaeagnus angustifolia L. (Russian olive) in western North America. Wetlands 23:763–777
- Kimball S, Kulow M, Sorenson Q, Balazs K, Fang Y, Davis S, O'Connell M, Huxman T (2015) Cost-effective ecological restoration. Restor Ecol 23: 800–810
- Lesica P, Miles S (1999) Russian olive invasion into cottonwood forests along a regulated river in north-central Montana. Can J Bot 77: 1077–1083
- Lesica P, Miles S (2001) Natural history and invasion of Russian olive along eastern Montana rivers. West N Am Nat 61:1–10
- Levine JM, Adler PB, Yelenik SG (2004) A meta-analysis of biotic resistance to exotic plant invasions. Ecol Lett 7:975–989
- Li Kui L, Stella JC (2016) Fluvial sediment burial increases mortality of young riparian trees but induces compensatory growth response in survivors. Forest Ecol Manag 366:32–40

- Mineau MM, Baxter CV, Marcarelli AM (2011) A non-native riparian tree (*Elaeagnus angustifolia*) changes nutrient dynamics in streams. Ecosystems 14:353–365
- Morgan JW (1999) Have tubestock plantings successfully established populations of rare grassland species into reintroduction sites in western Victoria? Biol Conserv 89:235–243
- Mulhouse JM, Galatowitsch SM (2003) Revegetation of prairie pothole wetlands in the mid-continental US: twelve years post-reflooding. Plant Ecol 169:143–159
- [NRCS] Natural Resources Conservation Service. (2006). Establishment of Bareroot and Container Stock in Riparian Areas. http://www.mt. nrcs.usda.gov/technical/ecs/forestry/technotes/forestryMT26. Accessed: May 10, 2011
- [NRCS] Natural Resources Conservation Service. (2008). Riparian Forest Buffer Specifications 391. http://efotg.sc.egov.usda.gov/references/ public/MT/391_spec_Nov_2008.pdf. Accessed: May 10, 2011
- Nagler PL, Glenn EP, Jarnevich CS, Shafroth PB (2011) Distribution and abundance of saltcedar and Russian olive in the western United States. Crit Rev Plant Sci 30:508–523
- Naiman RJ, Decamps H, Pollock M (1993) The role of riparian corridors in maintaining regional biodiversity. Ecol Appl 3:209–212
- Palmer MA, Ambrose RF, Poff NL (1997) Ecological theory and community restoration ecology. Restor Ecol 5:291–300
- Pearce CM, Smith DG (2001) Plains cottonwood's last stand: can it survive invasion of Russian olive onto the Milk River, Montana floodplain? Environ Manag 28:623–637
- Perkins LB, Hatfield G, Espeland EK (2016) Invasive grasses consistently create similar plant-soil feedback types in soils collected from geographically distant locations. J Plant Ecol 9:180–186
- Reever Morghan KJ, Sheley RL, Denny MK, Pokorny ML (2005) Seed islands may promote establishment and expansion of native species in reclaimed mine sites (Montana). Ecol Restor 23:214–215
- Richardson DM, Holmes PM, Esler KJ, Galatowitsch SM, Stromberg JC, Kirkman SP, Pysek P, Hobbs RJ (2007) Riparian vegetation: degradation, alien plant invasions, and restoration prospects. Divers Distrib 13:126–139
- Rinella MJ, Hammond DH, Bryant AEM, Kozar BJ (2015) High precipitation and seeded species competition reduce seeded shrub establishment during dryland restoration. Ecol Appl 25:1044–1053
- Rinella MJ, Mangold JM, Espeland EK, Sheley RL, Jacobs JS (2012) Long-term effects of introducing desired plants into invaded grass-lands. Ecol Appl 22:1320–1329
- Rinella M J, Maxwell BD, Fay PK, Weaver T, Sheley RL (2009) Control effort exacerbates invasive-species problem. Ecol Appl 19:155–162
- Robichaud PR, Beyers JL, Neary DG (2000). Evaluating the effectiveness of postfire rehabilitation treatments. General Technical Report RMRS-GTR-63. Fort Collins, CO: Rocky Mountain Research Station. 85p
- Ruwanza S, Gaertner M, Richardson DM (2013) The effectiveness of active and passive restoration on recovery of indigenous vegetation in riparian zones in the Western Cape, South Africa: a preliminary assessment. S Afr J Bot 88:132–141
- Scianna J, Kilian R, Muscha J (2012). Russian Olive *Elaeagnus angustifolia* L. Seed Longevity. USDA Natural Resources Conservation Service Plant Materials Technical Note No. MT-86. 3 p
- Stannard M, Ogle D, Holzworth L, Scianna J, Sunleaf E (2002) History, Biology, Ecology, Suppression and Revegetation of Russian-Olive Sites. Boise, ID: USDA Natural Resources Conservation Service Plant Materials. Technical Note No 47:14 p

- Srivastava DS, Vellend M (2005) Biodiversity-ecosystem function research: is it relevant to conservation? Annu Rev Ecol Evol Syst 36:267–294
- Suding KN (2011) Toward an era of restoration in ecology: successes, failures, and opportunities ahead. Annu Rev Ecol Evol Syst 42:465–487
- Sweeney BW, Czapka SJ (2004) Riparian forest restoration: why each site needs an ecological prescription. Forest Ecol Manag 192:361–373
- Tererai F, Gaertner M, Jacobs S, Richardson DM (2015) Resilience of invaded riparian landscapes: the potential role of soil-stored seed banks. Environ Manag 55:86–99
- Toledo D, Sanderson M, Spaeth K, Hendrickson J, Printz J (2014) Extent of Kentucky bluegrass and its effect on native plant species

- diversity and ecosystem services in the Northern Great Plains of the United States. Invasive Plant Sci Manag 7:543–552
- Zavaleta ES, Hobbs RJ, Mooney HA (2001) Viewing invasive species removal in a whole-ecosystem context. Trends Ecol Evol 16:454–459
- Zedler JB, Callaway JC (1999) Tracking wetland restoration: do mitigation sites follow desired trajectories? Restor Ecol 7:69–73

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