Dietary isohumulones, the bitter components of beer, raise plasma HDL-cholesterol levels and reduce liver cholesterol and triacylglycerol contents similar to PPARα activations in C57BL/6 mice

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The effects of dietary isohumulones, the main components accounting for the bitter taste of beer, on lipid metabolism were examined. Young female C57BL/6N mice were fed diets containing isomerized hop extract (IHE), which consists mainly of isohumulones. Administration of IHE with an atherogenic (high-fat and high-cholesterol) diet for 2 weeks resulted in a significant increase in plasma HDL-cholesterol (P < 0.01), along with a concomitant reduction in the atherosclerosis index, an increase in liver weight and a decrease in body weight gain in a dose-dependent manner. When animals received IHE with either a cholesterol or a basal diet for 1 week, significant decreases in the liver content of cholesterol (P < 0.01) and triacylglycerol (cholesterol diet, P < 0.01) were observed. Quantitative analyses of hepatic mRNA levels revealed that IHE administration resulted in up-regulation of mRNA for acyl-CoA oxidase, acyl-CoA synthetase, hydroxymethylglutaryl-CoA synthetase, lipoprotein lipase and fatty acid transport protein, and down-regulation of mRNA for Apo CIII and Apo AI. Administration of purified isohumulones effectively resulted in the same changes as IHE. Administration of fenofibrate, an agonist for PPARα, with a cholesterol diet caused marked hepatomegaly, an increase in plasma HDL-cholesterol, a decrease in hepatic cholesterol content, and alterations in hepatic mRNA levels similar to those observed in mice given IHE. Taken together, these results suggest that the modulation of lipid metabolism observed in mice fed diets containing isohumulones is, at least in part, mediated by activation of PPARα.

Isohumulones: Beer: PPARα: Lipid metabolism

Epidemiological studies have reported that moderate alcohol consumption is associated with decreased risk of CHD. The relationship between the amount of alcohol consumed and risk of CVD has been described as a J-shaped curve: the relative risk of mortality is lower for moderate drinkers than for non-drinkers, but then increases with higher levels of alcohol consumption (Doll et al. 1994; Fuchs et al. 1995; Gaziano et al. 2000). The protective effects of alcohol beverages on the vascular system can be attributed, in part, to the fact that ethanol can increase plasma levels of HDL and reduce thrombotic tendency by inhibiting platelet aggregation (Sub et al. 1992).

Non-alcohol components in alcohol beverages are also considered important for their cardioprotective effects. The high antioxidant activity of red wine resulting from the high content of polyphenols is one possible explanation for the ‘French paradox’ i.e. that some regions of France have lower mortality rates due to CVD than do North European countries, even though the populations are exposed to similar CVD risk factors (dietary fat intake and plasma cholesterol; Renaud & de Lorgeril, 1992; Criqui & Ringel, 1994). Hayek et al. (1997) reported that the progression of atherosclerosis in mice deficient in Apo E was reduced by the consumption of red wine or its polyphenols; this reduction was also associated with a decrease in the susceptibility of LDL to oxidation and aggregation. Furthermore, the consumption of red wine polyphenols in man has been reported to reduce the susceptibility of LDL to oxidation in vivo (Nigdliler et al. 1998).

Several studies have demonstrated that beer, as well as red wine, has beneficial effects in protecting against CHD (Hoffmeister et al. 1999; Brenner et al. 2001). It has been reported that moderate consumption of beer does not affect plasma levels of homocysteine, which promotes atherosclerosis and thrombosis, whereas the intake of red wine and spirits increases homocysteine levels (Van der Gaag et al. 2000). The high content of folate and vitamin B6 in beer is likely to be responsible for this effect. This observation is supported by epidemiological studies showing that there is a negative association between beer consumption and plasma homocysteine levels (Mayer et al. 2001; Mennen et al. 2003).

Beer is the only alcoholic beverage that contains hops, the female inflorescences of the hop plant (Humulus lupulus L.). Hops are used as a preservative and flavouring in beer. The bitter taste of beer is mainly attributed to isohumulones, which mostly comprise isohumulone, isocohumulone and isoadhumulone. Isohumulones are converted from humulones present in hops by isomerization...
during the wort boiling step of the brewing process. Humulones have been reported to have various biological activities, such as inhibiting angiogenesis, inducing differentiation of myelogenous leukaemia cells and inhibiting bone resorption (Mizobuchi & Sato, 1984, 1985; Miranda et al. 1993; Yasukawa et al. 1995; Toeb et al. 1997; Honma et al. 1998; Yamamoto et al. 2000; Shimagura et al. 2001). Although the biological activities of humulones have been studied extensively, there are few studies concerning the activities of the isohumulones found in beer.

Recently, we found from reporter gene expression analyses that isohumulones can activate PPARα and γ (Yajima et al. 2004), PPARγ activators have been shown to improve insulin sensitivity and are commonly used to treat type 2 diabetes. Dietary administration of isohumulones to diabetic mice resulted in an improvement in insulin resistance, as did pioglitazone, a well-known activator of PPARγ, which stimulate hepatic fatty acid oxidation, are used to treat hyperlipidaemia (Kersten et al. 2000). Therefore, it is reasonable to speculate that an intake of isohumulones from beer may modulate lipid metabolism.

The purpose of the present study was to evaluate the effect of isohumulones, the principal components accounting for the bitter taste of beer, on the metabolism of lipids in vivo in C57BL/6 mice.

Materials and methods

Chemicals

Two types of isomerized hop extract (IHE) containing isohumulones were used. One (Isomerized Kettle Extract; S. S. Steiner, Inc., New York, NY, USA) is a thick brown syrup and contains isohumulones (571 g/l), humulones (10 g/l), lupulones (165 g/l) and hop oil (65 g/l); the other (ISOHOP CON2; English Hop Products Co. Ltd, Tonbridge, Kent, UK) is an aqueous solution containing isohumulones (300 g/l), and is termed IHE2 in this study. Isohumulones are composed mainly of isohumulone [2-(3-methyl-2-butenyl)-3,4-dihydroxy-4-(4-methyl-3-pentenoyl)-2-cyclopentenone], isocohumulone [2-(2-methylpropanoyl)-5-(3-methyl-2-butenyl)-3,4-dihydroxy-4-(4-methyl-3-pentenoyl)-2-cyclopentenone] and isoadohumulone [2-(2-methylbutanoyl)-5-(3-methyl-2-butenyl)-3,4-dihydroxy-4-(4-methyl-3-pentenoyl)-2-cyclopentenone]. Purified isohumulones were prepared from IHE2 by silica gel chromatography as follows. The IHE2 solution was neutralized to pH 7.8 with HCl and passed over a silica gel column equilibrated with a solution of hexane–ethylacetate (2:1, v/v). Isohumulones were eluted with the same solution and the eluates were evaporated under a stream of N2 gas. The isohumulones were diluted with an appropriate amount of ethanol and stored at −80°C until use to minimize the oxidation of isohumulones. Thawed feed was provided to animals daily.

Animals

Young female C57BL/6NCrj mice, 5 weeks old, were purchased from Charles River Japan Inc. (Tokyo, Japan). To accustom them to the experimental conditions, the mice were housed in animal rooms for at least 5 d before the experiments, with free access to food and water. Three or four mice were housed per cage, and food intake was measured on a per cage basis. The animals were treated in accordance with Kirin Pharmaceutical’s ethical guidelines for animal care, handling and termination. These guidelines meet the generally accepted international criteria of humane treatment, i.e. sparing the animals needless pain and suffering and ensuring that the experiments conducted are of actual scientific benefit to mankind. At the end of the experiments, the mice were anesthetized with diethyl ether at about 10.00 hours and killed by draining blood via the postcaval vein. The livers were removed immediately, frozen in liquid N2 and stored at −80°C.

Diets and handling of samples

The atherogenic (A) diet was prepared according to Nishina et al. (1990, 1993) and contained the following ingredients per kg: 150 g butter, 524.5 g sucrose, 200 g casein, 10 g corn oil, 50 g cellulose, 35 g AIN-93G mineral mixture (Reeves et al. 1993), 10 g AIN-93 vitamin mixture (Reeves et al. 1993), 2.5 g choline chloride, 3 g cystine, 10 g cholesterol and 5 g sodium cholate. AIN76-A diet (American Institute of Nutrition, 1977), which was purchased from Dyets Inc. (Bethlehem, PA, USA), was used as the basal (B) diet. The cholesterol (C) diet was prepared by adding cholesterol (2 kg/kg) to the basal diet. In all experiments, the dietary components were mixed, rolled into pellets with distilled water and stored at −20°C until use to minimize the oxidation of isohumulones. Thawed feed was provided to animals daily.

Experimental design

In Expt 1, mice were fed the atherogenic diet for 1 week and then divided into five groups of nine animals each (except for the basal diet group, which contained six animals). Over the next 2 weeks, the groups were fed the atherogenic diet, the atherogenic diet supplemented with either two different concentrations of IHE1 (A + IHE1 (low), A + IHE1 (high)) or IHE2 (A + IHE2 (high)), or the basal diet. The A + IHE1 (low) diet contained 2 ml IHE1/kg diet (equivalent to 1.1 g isohumulones/kg diet), the A + IHE1 (high) diet contained 5 ml IHE1/kg diet (equivalent to 2.9 g isohumulones/kg diet), and the A + IHE2 diet contained 10 ml IHE2/kg diet (equivalent to 3 g isohumulones/kg diet). In Expt 2, mice were divided into four groups of five or six animals each. The groups were maintained for 1 week on the basal diet, the basal diet containing IHE2 (B + IHE2 diet; 10 ml IHE2/kg diet, equivalent to 3 g isohumulones/kg diet), the cholesterol diet, or the cholesterol diet containing IHE2 (C + IHE2 diet; 10 ml IHE2/kg diet, equivalent to 3 g isohumulones/kg diet). In Expt 3, mice were divided into three groups of eight or nine animals each. The groups were maintained for 2 weeks on the cholesterol diet without supplement, the cholesterol diet containing fenofibrate (C + F diet; 0.5 g fenofibrate/kg diet), or the cholesterol diet containing IHE2 (C + IHE2 diet; 10 ml IHE2/kg diet, equivalent to 3 g isohumulones/kg diet). In Expt 4, mice were divided into three groups of seven or eight animals each. The groups were maintained for 1 week on the cholesterol diet, the cholesterol diet containing IHE2 (C + IHE2; 10 ml IHE2/kg diet, equivalent to 3 g isohumulones/kg diet), or the cholesterol diet containing purified isohumulones (3 g isohumulones/kg diet). All blood samples were taken from non-fasted mice except for Expt 1, where blood samples were removed from animals after an overnight fast.
Biochemical analyses

Plasma lipids (total cholesterol, HDL-cholesterol and triacylglycerols) were determined using commercial kits (Cholesterol C-test, HDL-cholesterol-test and Triglyceride G-test, respectively; Wako Pure Chemical, Osaka, Japan). To analyse the hepatic lipid content, approximately 0·2 g liver tissue was homogenized in saline and the lipids were extracted with chloroform–methanol mixture (2:1, v/v) as described by Carr et al. (1993). Liver cholesterol and triacylglycerols were measured with the kits described above. Gel filtration chromatography was performed using a Superose 6 HR 10/30 column (Amersham Pharmacia Biotech, Uppsala, Sweden) on 150 μl pooled plasma (Ha & Barter, 1985). The gel was allowed to equilibrate with 10 mm-PBS containing EDTA (0·1 g/l) and sodium azide (0·1 g/l). Pooled plasma (150 μl) was applied to the column and eluted with the same buffer at 4°C at a flow rate of 0·33 ml/min. The eluate was collected in 0·5 ml fractions. Cholesterol was measured in each fraction using the commercial enzymatic kit described above.

The atherosclerosis index (AI) was calculated as follows:

\[ AI = \frac{\text{(total cholesterol} - \text{HDL-cholesterol})}{\text{HDL-cholesterol}}. \]

Real-time quantitative PCR analysis of gene expression

Light cycler analysis with SYBR green I fluorescence (Roche Diagnostics, Basel, Switzerland) was used to determine the relative mRNA levels of the genes involved in lipid metabolism. Oligonucleotide primers were designed using MacVector 6·5 (Teijin System Technology, Tokyo, Japan) for hydroxymethylglutaryl-CoA reductase (HMG-CoA reductase; HMGR; accession no. X07887 and X07889, HMG-CoA synthetase (HMSG; AK004865), LDL receptor (LDLR; NM_010700), acyl-CoA oxidase (ACO; NM_015729), acyl-CoA synthetase (ACS; U15977), fatty acid synthase (FAS; AF127033), Apo AI (NM_009693), Apo CIII (HMG)-CoA reductase (HMGR; accession no. X07887 and X07889), lipoprotein lipase (LPL; NM_008509), cholesterol acyltransferase II (ACAT2; NM_020311), lipoprotein lipase (LPL; NM_008509), cholesterol 7α-hydroxylase (CYP7A1; L23754), fatty acid transport protein (FATP; AAH009696) and acidic ribosomal phosphoprotein P0 (ARP0; X11267).

The following PCR reaction primers were used: for HMGR, 5′-TGG GCC TCC ATT GAT GTC CG-3′ (5′ primer) and 5′-GAT CTT GTT GTC GCC GGT GAA C-3′ (3′ primer); for HMGS, 5′-CCT TCA GGG TCA GTA AGC TGG AAG-3′ (5′ primer) and 5′-CAG CCA ATT GTC CGG CAG AGT G-3′ (3′ primer); for LDLR, 5′-CAT CAA GAG GTC CAA CGG-3′ (5′ primer) and 5′-CAC TTG TAG CTG CCT TCC AGG TTC-3′ (3′ primer); for ACO, 5′-ATC TAT GAC CAG GGT GTT CTG CAG GGG-3′ (5′ primer) and 5′-CCA CGC CAC TCC CCT GCT GCT CCT C-3′ (3′ primer); for ACS, 5′-GGATACT ACA GCC AAC CCC AAA G-3′ (5′ primer) and 5′-CTT GAT GTC GTC CAT AAG CAC GAG C-3′ (3′ primer); for FAS, 5′-TGC CCT ATG GTG GTG GAA GAT C-3′ (5′ primer) and 5′-TGC CCT ATG GTG GAA GAT C-3′ (3′ primer); for Apo AI, 5′-TGG ATG ATG CTG AGG TCA AAG AC-3′ (5′ primer) and 5′-TCA ATT GTA GTC CCT GCA CCT AAT CTG-3′ (3′ primer); for Apo CIII, 5′-AGG GCT ACA AGC AAC C-3′ (5′ primer) and 5′-CGA TCT CCT GCT GGT GAT C-3′ (3′ primer); for Apo A1, 5′-AGG GCT ACA AGC AAC C-3′ (5′ primer) and 5′-CGA TCT CCT GCT GGT GAT C-3′ (3′ primer); for ACO, 5′-ATC TAT GAC CAG GGT GTT CTG CAG GGG-3′ (5′ primer) and 5′-CCA CGC CAC TCC CCT GCT GCT CCT C-3′ (3′ primer); for ACS, 5′-GGATACT ACA GCC AAC CCC AAA G-3′ (5′ primer) and 5′-CTT GAT GTC GTC CAT AAG CAC GAG C-3′ (3′ primer); for FAS, 5′-TGC CCT ATG GTG GTG GAA GAT C-3′ (5′ primer) and 5′-TGC CCT ATG GTG GAA GAT C-3′ (5′ primer); for ACO, 5′-ATC TAT GAC CAG GGT GTT CTG CAG GGG-3′ (5′ primer) and 5′-CCA CGC CAC TCC CCT GCT GCT CCT C-3′ (3′ primer); for Apo AI, 5′-AGG GCT ACA AGC AAC C-3′ (5′ primer) and 5′-CGA TCT CCT GCT GGT GAT C-3′ (3′ primer); for ACO, 5′-ATC TAT GAC CAG GGT GTT CTG CAG GGG-3′ (5′ primer) and 5′-CCA CGC CAC TCC CCT GCT GCT CCT C-3′ (3′ primer).
attributed to the increase in HDL-cholesterol. Therefore, the AI values declined in a dose-dependent manner in the groups fed IHE.

Pooled plasma from mice fed the atherogenic diet plus IHE2 (A + IHE2 (high)) diet was fractionated by gel-filtration chromatography and the distribution of cholesterol was measured (Fig. 1). The profile suggested that the number of HDL particles in the plasma of IHE-fed mice increased, whereas the number of LDL particles did not change, compared with mice fed the atherogenic diet. This observation is consistent with our result indicating that there was a specific increase in the level of plasma HDL-cholesterol. Therefore, the AI, atherogenic index; TAG, triacylglycerols.

Table 1. Body weight, energy intake, organ weight and plasma lipid content in mice fed the atherogenic diet, the atherogenic diet supplemented with isomerized hop extract (IHE) and the basal diet for 2 weeks†

<table>
<thead>
<tr>
<th></th>
<th>A</th>
<th>A + IHE1 (low)</th>
<th>A + IHE1 (high)</th>
<th>A + IHE2 (high)</th>
<th>B</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean</td>
<td>SD</td>
<td>Mean</td>
<td>SD</td>
<td>Mean</td>
</tr>
<tr>
<td>Body weight gain (g)</td>
<td>1·75</td>
<td>0·75</td>
<td>0·58**</td>
<td>0·79</td>
<td>0·43**</td>
</tr>
<tr>
<td>Final body weight (g)</td>
<td>17·98</td>
<td>1·16</td>
<td>16·70**</td>
<td>0·88</td>
<td>16·44**</td>
</tr>
<tr>
<td>Food intake (kJ/2 weeks per mouse)</td>
<td>43·1</td>
<td>41·0</td>
<td>43·1</td>
<td>40·5</td>
<td></td>
</tr>
<tr>
<td>Tissue weight (g/body kg)</td>
<td>Liver: 56·47</td>
<td>4·65</td>
<td>56·34</td>
<td>4·28</td>
<td>63·42**</td>
</tr>
<tr>
<td></td>
<td>Kidney: 11·47</td>
<td>0·89</td>
<td>11·32</td>
<td>0·59</td>
<td>10·96</td>
</tr>
<tr>
<td></td>
<td>Plasma total cholesterol (mmol/l)</td>
<td>2·42</td>
<td>0·39</td>
<td>2·83*</td>
<td>0·49</td>
</tr>
<tr>
<td></td>
<td>Plasma HDL-cholesterol (mmol/l)</td>
<td>0·96</td>
<td>0·16</td>
<td>1·29*</td>
<td>0·25</td>
</tr>
<tr>
<td></td>
<td>AI</td>
<td>1·52</td>
<td>0·20</td>
<td>1·26</td>
<td>0·59</td>
</tr>
<tr>
<td></td>
<td>Plasma TAG (mmol/l)</td>
<td>0·49</td>
<td>0·07</td>
<td>0·44</td>
<td>0·12</td>
</tr>
</tbody>
</table>

A, atherogenic diet group; A + IHE1 (low), A diet supplemented with IHE1 (2 ml/kg, equivalent to 1·1 g isohumulones/kg) group; A + IHE1 (high), A diet supplemented with IHE1 (5 ml/kg, equivalent to 2·9 g isohumulones/kg) group; A + IHE2 (high), A diet supplemented with IHE2 (10 ml/kg, equivalent to 3 g isohumulones/kg) group; B, basal diet group; AI, atherogenic index; TAG, triacylglycerols.

Mean values were significantly different from those of the A diet group: *P<0·05, **P<0·01.
† For details of diets and procedures, see p. 560.

Effect of dietary isomerized hop extract on lipids in the liver (Expt 2)

We examined the effect of IHE on lipid metabolism by using two diets, the basal diet and the cholesterol diet. A significant suppression in body weight gain was observed in the group fed IHE with the cholesterol diet (C + IHE2, 76 % decrease compared with the cholesterol diet, P<0·05), although no significant differences in food intake or final body weight were observed among the four groups (Table 2). Administration of IHE caused an increase in liver weight, regardless of the presence of cholesterol, but this increase was significant only when IHE was given with the basal diet (B + IHE2, 28 % increase for the basal diet, P<0·01; Table 2). Dietary IHE tended to increase the level of plasma HDL-cholesterol and thus decrease the AI, although the differences were not so marked as those detected in the groups fed the atherogenic diets. Liver contents of cholesterol and triacylglycerols in mice fed the cholesterol diet plus IHE (C + IHE2) were markedly reduced compared with those found in mice fed the cholesterol diet (38 % and 37 % decrease, respectively, P<0·01; Table 2). A reduction in liver cholesterol and triacylglycerol contents was also observed in mice fed IHE with the basal diet (B + IHE2, 33 % (P<0·01) and 12 % decrease, respectively; Table 2).

Effect of dietary isomerized hop extract on hepatic mRNA levels (Expt 2)

We performed quantitative, real-time, RT-PCR analyses to measure mRNA levels in the livers of mice from four dietary groups (Fig. 2). mRNA levels, which were first normalized to levels of 36B4 mRNA in the same samples as an internal control, were determined relative to the mRNA levels in the group fed the cholesterol diet. Administration of IHE caused an approximately twofold increase in the levels of mRNA for enzymes involved
Table 2. Body weight, energy intake, liver weight, plasma lipid content and hepatic lipid content in mice fed the cholesterol diet, the basal diet, and the cholesterol and basal diets supplemented with isomerized hop extract (IHE) for 1 week†

(Mean values with standard deviations for five or six mice per group)

<table>
<thead>
<tr>
<th></th>
<th>C</th>
<th>C + IHE²</th>
<th>B</th>
<th>B + IHE²</th>
<th>ANOVA</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean</td>
<td>SD</td>
<td>Mean</td>
<td>SD</td>
<td>Mean</td>
</tr>
<tr>
<td></td>
<td>Cholesterol</td>
<td>Isohumulone</td>
<td>Interaction</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Body weight gain (g)</td>
<td>1·07</td>
<td>0·41</td>
<td>0·25*</td>
<td>0·42</td>
<td>0·68</td>
</tr>
<tr>
<td>Final body weight (g)</td>
<td>17·09</td>
<td>0·99</td>
<td>16·33</td>
<td>0·40</td>
<td>16·05</td>
</tr>
<tr>
<td>Food intake (kJ/week per mouse)</td>
<td>54·0</td>
<td>51·1</td>
<td>54·4</td>
<td>61·5</td>
<td></td>
</tr>
<tr>
<td>Liver weight (g/body kg)</td>
<td>61·92</td>
<td>4·87</td>
<td>67·77</td>
<td>5·47</td>
<td>52·2</td>
</tr>
<tr>
<td>Plasma total cholesterol (mmol/l)</td>
<td>1·90</td>
<td>0·13</td>
<td>2·08</td>
<td>0·14</td>
<td>2·22</td>
</tr>
<tr>
<td>Plasma HDL-cholesterol (mmol/l)</td>
<td>0·98</td>
<td>0·15</td>
<td>1·12</td>
<td>0·05</td>
<td>1·28</td>
</tr>
<tr>
<td>AI</td>
<td>0·98</td>
<td>0·26</td>
<td>0·86</td>
<td>0·15</td>
<td>0·73</td>
</tr>
<tr>
<td>Liver weight (µmol/g liver)</td>
<td>30·07</td>
<td>5·40</td>
<td>18·50**</td>
<td>3·60</td>
<td>8·48</td>
</tr>
<tr>
<td>Cholesterol</td>
<td>39·66</td>
<td>8·14</td>
<td>24·97**</td>
<td>3·16</td>
<td>24·97</td>
</tr>
</tbody>
</table>

C, cholesterol diet group; C + IHE², diet supplemented with IHE² (10 ml/kg, equivalent to 3 g isohumulones/kg) group; B, basal diet group; B + IHE², diet supplemented with IHE² (10 ml/kg, equivalent to 3 g isohumulones/kg) group; AI, atherogenic index; TAG, triacylglycerols.

Mean values were significantly different from those of the C group: *P < 0·05, **P < 0·01.

Mean values were significantly different from those of the B group: ††P < 0·01.

† For details of diets and procedures, see p. 560.

Effects of dietary isomerized hop extract and fenofibrate on liver lipid and gene expression (Expt 3)

We compared the effects of IHE on liver lipid content and hepatic mRNA levels with those of fenofibrate, a well-known PPARα agonist that is used for the medical treatment of hyperlipidaemia. Administration of fenofibrate with the cholesterol diet for 2 weeks exaggerated the changes that had been observed in IHE-fed mice, i.e. marked hepatomegaly (64 % increase in the weight), a significant decrease in hepatic cholesterol content (53 %), a significant increase in plasma HDL-cholesterol (69 %) and a significant decrease in plasma triacylglycerols (56 %) were observed in the group given fenofibrate (all P < 0·01, Table 3). IHE increased liver weight by 19 % and decreased hepatic cholesterol by 32 % (P < 0·01). Administration of IHE also caused a slight increase in HDL-cholesterol and a decrease in plasma triacylglycerols, although the changes were not significant. Notably, IHE caused a greater reduction in hepatic triacylglycerol content than did fenofibrate (33 % and 7 % reductions, respectively).

The effects of fenofibrate and IHE were assessed by determining the levels of mRNA that are potentially regulated by PPARα agonist to a greater extent than did fenofibrate.

Comparison of the effects of dietary isomerized hop extract and isohumulones (Expt 4)

Isohumulones were purified from IHE² and given to the mice for 1 week. The administration of purified isohumulones (3 g/kg cholesterol diet) had almost the same effect as administration of IHE² (10 ml/kg cholesterol diet, equivalent to 3 g isohumulones/
are major ingredients, led to a significant increase in plasma of the atherogenic diet containing IHE, in which isohumulones CoA oxidase; LPL, lipoprotein lipase.

<table>
<thead>
<tr>
<th></th>
<th>C</th>
<th>C + IHE2</th>
<th>C + F</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean ± SD</td>
<td>Mean ± SD</td>
<td>Mean ± SD</td>
<td>Mean ± SD</td>
</tr>
<tr>
<td>Body weight gain (g)</td>
<td>1.47 ± 0.22</td>
<td>1.12 ± 1.00</td>
<td>2.52 ± 1.05</td>
</tr>
<tr>
<td>Final body weight (g)</td>
<td>17.09 ± 0.61</td>
<td>16.52 ± 0.91</td>
<td>17.48 ± 1.23</td>
</tr>
<tr>
<td>Food intake (kJ/2 weeks per mouse)</td>
<td>44.4 ± 41.0</td>
<td>66.27 ± 8.47</td>
<td>91.08 ± 9.26</td>
</tr>
<tr>
<td>Liver weight (g/body kg)</td>
<td>55.48 ± 3.12</td>
<td>2.22 ± 0.24</td>
<td>3.43 ± 0.58</td>
</tr>
<tr>
<td>Plasma total cholesterol (mmol/l)</td>
<td>1.37 ± 1.44</td>
<td>0.22 ± 0.27</td>
<td>2.31 ± 0.34</td>
</tr>
<tr>
<td>Plasma HDL-cholesterol (mmol/l)</td>
<td>0.63 ± 0.06</td>
<td>0.09 ± 0.13</td>
<td>0.90 ± 0.12</td>
</tr>
<tr>
<td>AI</td>
<td>1.07 ± 0.37</td>
<td>1.09 ± 0.31</td>
<td>1.47 ± 0.05</td>
</tr>
<tr>
<td>Plasma TAG (mmol/l)</td>
<td>41.12 ± 3.93</td>
<td>28.01 ± 3.86</td>
<td>19.28 ± 1.54</td>
</tr>
<tr>
<td>Liver lipid (μmol/g liver)</td>
<td>57.18 ± 8.93</td>
<td>38.31 ± 11.41</td>
<td>53.56 ± 12.66</td>
</tr>
</tbody>
</table>

C, cholesterol diet group; C + IHE2, C diet supplemented with IHE2 (10 ml/kg, equivalent to 3 g isohumulones/kg) group; C + F, C diet supplemented with fenofibrate (5 g/kg) group; AI, atherogenic index; TAG, triacylglycerols.

Mean values were significantly different from those of the C diet group: *P < 0.05, **P < 0.01. ACO, acyl-CoA oxidase; LPL, lipoprotein lipase.

Table 3. Body weight, energy intake, liver weight, plasma lipid content and hepatic lipid content in mice fed the cholesterol diet, and the cholesterol diets supplemented with isomerized hop extract (IHE) and fenofibrate for 2 weeks†

†For details of diets and procedures, see p. 560.

Fig. 3. Quantitative analysis of mRNA in the liver of mice fed the cholesterol diet (C, □), the cholesterol diet supplemented with isomerized hop extract (C + IHE2, III) and the cholesterol diet supplemented with fenofibrate (C + F, □□) for 2 weeks. For each gene, the mRNA level is shown relative to its level in the C group (set as 1). Values are means with standard deviations represented by vertical bars for five mice per group. Mean values were significantly different from those of the C group: *P < 0.05, **P < 0.01. ACO, acyl-CoA oxidase; LPL, lipoprotein lipase.

kg cholesterol diet) on liver lipid content and mRNA levels (Table 4 and Fig. 4). No notable difference was observed in the extent of the increase in ACO and LPL mRNA levels or in the extent of the reduction in hepatic cholesterol and triacylglycerol content between the two groups (Table 4 and Fig. 4). From these results, it seems reasonable to conclude that isohumulones are responsible for the hypolipidaemic effects of IHE.

Discussion

This study was conducted to examine whether isohumulones, the main components accounting for the bitter taste of beer, affect lipid metabolism in vivo by using an animal model. Administration of the atherogenic diet containing IHE, in which isohumulones aref major ingredients, led to a significant increase in plasma HDL-cholesterol, coupled with a concomitant reduction in the AI, in a dose-dependent manner in mice (Table 1). A specific increment in the HDL-cholesterol fraction in plasma was confirmed by gel filtration chromatography (Fig. 1). It should be noted that the significant increase in plasma HDL-cholesterol by IHE was observed on the atherogenic diet (Table 1), while the increases in plasma HDL-cholesterol by IHE on the basal and cholesterol diets were not significant (Tables 2 and 3). A significant decrease in the hepatic triacylglycerol and cholesterol content in mice given cholesterol and basal diets supplemented with IHE was also observed (Tables 2 and 3).

Quantitative analyses of the mRNA in liver revealed that administration of IHE with either the cholesterol or the basal diet caused a significant increase in, or tended to increase, the levels of ACO, ACS, FATP, LPL and HMGS mRNA, and also caused a significant decrease in, or tended to decrease, the levels of Apo AI and Apo CIII mRNA (Fig. 2). The enzymes encoded by ACS, FATP and ACO mRNA are involved in cellular fatty acids uptake, first step prior to transport of fatty acids into the mitochondria, and the peroxisomal β-oxidation of fatty acids, respectively (Kersten et al. 2000). An up-regulation of these genes by IHE would therefore increase β-oxidation activity in the liver and would result in a reduction in hepatic triacylglycerol content. Apo CIII and LPL play crucial roles in triacylglycerol metabolism, with early studies having demonstrated that Apo CIII inhibits the hydrolysis of VLDL triacylglycerols by LPL (Schaeck & Eisenberg, 1991; Aalto-setälä et al. 1992). The decrease in Apo CIII mRNA and the increase in LPL mRNA in the liver of IHE-fed mice may also be responsible for the reduction in hepatic triacylglycerol content and plasma triacylglycerol levels. Administration of IHE was observed to cause a significant reduction in plasma triacylglycerol levels only in Expt 1 (Table 1, A + IHE2 (high) diet). This may be due to the timing of blood collection (blood was taken from non-fasted mice in Expts 2, 3 and 4). Although a significant decrease in hepatic cholesterol content was observed in mice given IHE (Tables 2, 3 and 4), there was no alteration in the hepatic mRNA levels of HMGR and CYP7A1, which are responsible for the synthesis and excretion of cholesterol, respectively.
Isohumulones improve lipid metabolism

Notably, no difference was detected in the content of cholesterol and bile acid in faeces between the control and IHE-fed mice (data not shown).

The genes encoding ACO, ACS, FATP and LPL, whose mRNA were up-regulated by the administration of IHE, are known to be regulated by PPARα, and our experiments using a transient co-transfection assay in mammalian cells have indicated that isohumulones activate PPARαs at a physiological concentration of 10 μM (Yajima et al. 2004). We therefore compared the effects of IHE on lipid metabolism with those of fenofibrate, a well-known PPARα agonist. It has been shown that agonists of PPARα induce significant hepatomegaly in mice and accelerate the β-oxidation of fatty acids in mitochondria, peroxisomes and microsomes in liver (Reddy et al. 1986; Schoonjans et al. 1996a,b; Kersten et al. 2000; Fruchart, 2001). In the present study, we confirmed that the administration of fenofibrate induced hepatomegaly, reduced hepatic cholesterol content, and increased plasma HDL-cholesterol with a concomitant increase in total cholesterol and a decrease in the AI (Table 3). These changes were similar to those observed in mice fed diets containing IHE, although the significant decrease in hepatic triacylglycerol content was observed only in the IHE-fed mice and the significant increase in plasma HDL-cholesterol was observed only in fenofibrate-fed mice.

Although fenofibrate has been consistently reported to increase HDL-cholesterol levels in humans (Fruchart, 2001), the results from animal studies are controversial. One study has reported that fenofibrate decreased plasma HDL-cholesterol concentration in a rodent model (Berthou et al. 1996). By contrast, fenofibrate has been found to increase HD-cholesterol levels in other rodent models (Oliver et al. 1988; Wang et al. 2001). It has also suggested that dietary fat content may affect lipoprotein metabolism in animals treated with fibrate-like compounds (Guo et al. 2001, Wang et al. 2001). In our experimental conditions, administration of the cholesterol diets containing fenofibrate resulted in the significant increase in plasma HDL-cholesterol (Table 3). Notable increase in HDL-cholesterol by IHE could not be detected in the same condition, although the administration of IHE and the atherogenic diet caused a significant increase in plasma HDL-cholesterol (Table 1). This may be due to the difference in lipid and cholesterol contents between the atherogenic and cholesterol diets.

The decrease in Apo AI mRNA levels observed in mice given IHE was inconsistent with the tendency towards an increase in HDL-cholesterol (Fig. 2 and Table 2). The reason for this discrepancy is unclear. Notably, the level of Apo AI mRNA in the liver of fenofibrate-fed mice was also decreased (data not shown). ACO and LPL mRNA in the liver of fenofibrate-fed mice were up-regulated, as were those in mice fed IHE or isohumulones (Figs. 3 and 4). Taken together, these results suggest that the changes in lipid metabolism observed in the mice given IHE are due, at least in part, to the activation of hepatic PPARαs by isohumulones. Fenofibrate has been reported to reduce the hepatic cholesterol content in rodents (Petit et al. 1988). The decrease in hepatic cholesterol content observed in IHE-fed mice may be caused by a similar mechanism. Recently, fenofibrate was shown to inhibit a high-fat diet-induced body weight gain in C57BL/6 mice (Guerre-Millo et al. 2000), which is compatible with the reduction in body weight observed in mice fed the atherogenic diet with IHE (Table 1).

The present study indicates that IHE i.e. isohumulones and fenofibrate have similar modulatory effects on both plasma and hepatic lipid levels and on the expression of genes involved in lipid metabolism. Our results suggest that activation of the

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**Table 4.** Body weight, liver weight and hepatic lipid content in mice fed the cholesterol diet, and the cholesterol diets supplemented with isomerized hop extract (IHE) and purified isohumulones for 1 week†

(Mean values with standard deviations represented by vertical bars for six mice per group)

<table>
<thead>
<tr>
<th></th>
<th>C</th>
<th>C + IHE2</th>
<th>C + purified isohumulones</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean</td>
<td>SD</td>
<td>Mean</td>
</tr>
<tr>
<td>Body weight gain (g)</td>
<td>0.80</td>
<td>0.75</td>
<td>-0.40**</td>
</tr>
<tr>
<td>Final body weight (g)</td>
<td>17.45</td>
<td>1.19</td>
<td>16.22*</td>
</tr>
<tr>
<td>Liver weight (g/body kg)</td>
<td>64.39</td>
<td>6.38</td>
<td>69.71*</td>
</tr>
<tr>
<td>Liver lipid (μmol/g liver)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cholesterol</td>
<td>25.52</td>
<td>2.26</td>
<td>19.94**</td>
</tr>
<tr>
<td>TAG</td>
<td>41.29</td>
<td>11.38</td>
<td>33.70</td>
</tr>
</tbody>
</table>

C, cholesterol diet group; C + IHE2, C diet supplemented with IHE2 (10 ml/kg, equivalent to 3 g isohumulones/kg group); C + purified isohumulones, C diet supplemented with purified isohumulones (3g/kg) group; AI, atherogenic index; TAG, triacylglycerols.

Mean values were significantly different from those of the C diet group: *P<0.05, **P<0.01.

† For details of diets and procedures, see p. 560.
nuclear receptor PPARs in liver by isohumulones is at least partly responsible for the effects observed in the mice given IHE. However, the increases in mRNA levels of PPARs-regulated genes were smaller in the mice given IHE than in those given fenofibrate, although a significant effect on the hepatic lipid content was observed in both groups (Fig. 3). It should be noted that the administration of a low amount of IHE with the cholesterol diet (equivalent to 0.9 g isohumulones/kg cholesterol diet) significantly decreased hepatic cholesterol and triacylglycerol content in mice, without causing a notable increase in ACO and LPL mRNA (data not shown). This result may suggest that the modulatory effects of isohumulones on lipid metabolism are only partially mediated by PPARs. The amounts of isohumulones taken by the animals fed the diet containing isohumulones at 3 g/kg diet, the most effective concentration, was approximately 500 mg/kg body weight (10 mg per animal). Therefore, the amount of beer could be roughly calculated to be 5–50 l/kg body weight per d, since the concentration of isohumulones in beer varies between 10 and 100 mg/l. It is not realistic to take sufficient amounts of beer to expect the modulatory effect on lipid metabolism. On the basis of our current findings, it is likely that the compounds responsible for beer bitterness, isohumulones, function to normalize lipid metabolism. Further studies are necessary to verify how isohumulones affect lipid metabolism and to assess their potential application as therapeutic agents.

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References


