Antioxidant vitamin supplements do not reduce reactive oxygen species activity in Helicobacter pylori gastritis in the short term

Simon M. Everett*, Ian M. Drake2, Kay L. M. White1, Nic P. Mapstone1, Douglas M. Chalmers1, Chris J. Schorah1 and Anthony T. R. Axon1

1The Centre for Digestive Diseases, The General Infirmary at Leeds, Great George Street, Leeds LS1 3EX, UK
2Chorley and South Ribble District General Hospital, Preston Road, Chorley, Lancashire, PR7 1PP, UK

(Received 20 February 2001 – Revised 17 July 2001 – Accepted 28 August 2001)

Gastric cancer is the second commonest cause of death from malignant disease worldwide (Parkin et al. 1993). Strategies to prevent this condition are therefore of great importance. It is well established that diets containing a high intake of the antioxidant vitamin C are associated with reduced risk of developing gastric cancer (Block, 1991). Vitamin E, which also has antioxidant properties, may play an additional protective role (Knekt et al. 1991). In recent years it has also been recognized that chronic infection of the gastric mucosa by Helicobacter pylori plays a pivotal role in gastric carcinogenesis (International Agency for Research on Cancer, 1994).

Infection of the gastric mucosa with Helicobacter pylori usually causes both an acute and chronic inflammatory cell infiltrate, leading to an increase in reactive oxygen species (ROS) (Davies et al. 1994). These are highly reactive compounds capable of combining with DNA in a number of potentially genotoxic ways (Jackson et al. 1989; Guyton & Kensler, 1993). ROS can react with the lipid bilayer releasing peroxidation products, such as malondialdehyde. These compounds, which are also able to react with DNA, have been shown to accumulate in H. pylori gastritis (Marnett, 1994; Farinati et al. 1996; Drake et al. 1998). These processes could lead to alterations in the structure of DNA facilitating mutations and carcinogenesis.

In the normal stomach, ascorbic acid and total vitamin C are highly concentrated from plasma into the mucosa and (to a lesser extent) gastric juice (Sobala et al. 1989; Banerjee et al. 1994). Vitamin C exists as ascorbic acid or dehydroascorbic acid. Ascorbic acid is the reduced form of the vitamin and can act as a potent antioxidant, able to scavenge ROS in gastric mucosa (Frei, 1991; Drake et al. 1996). This has been proposed as one means by which it exerts an anticarcinogenic effect (Correa, 1995). Ascorbic acid may also prevent formation of N-nitroso compounds in gastric juice by scavenging nitrite (Mirvish et al. 1972). Ascorbic acid is found principally in the aqueous phase.
whereas vitamin E (α-tocopherol) exists mainly in the lipid phase. Vitamin E regenerates ascorbic acid from its oxidised form dehydroascorbic acid and acts synergistically with ascorbic acid to prevent lipid peroxidation (Cadenas et al. 1996).

One way in which it may be possible to prevent carcinogenesis would be to reduce ROS damage to cellular constituents, especially DNA. It has been demonstrated that eradication of H. pylori leads to a reduction in ROS activity in the gastric mucosa (Drake et al. 1998). An alternative or additional approach might be to supplement antioxidant vitamins in the diet. The aim of the present study was, therefore, to determine whether dietary supplements of vitamins C and E are able to reduce levels of ROS and lipid peroxidation in the gastric mucosa, either alone or in combination with H. pylori eradication. We have randomized H. pylori positive patients in a 2 x 2 study design to receive eradication therapy or placebo for 2 weeks followed by vitamins C and E or placebo for 4 weeks (making four separate randomized groups) and assessed ROS and lipid peroxidation before and at the end of treatment.

Methods
Study design
This was a randomized, double blind, controlled trial. Patients with dyspepsia aged between 18 and 80 years attending for routine out-patient endoscopy were invited to participate. Patients were excluded if any of the following were present: pregnancy or lactation, significant co-morbidity, antibiotic use in the previous 6 weeks, previous attempts at H. pylori eradication, previous gastric surgery or a previous history of alcohol or drug abuse.

All patients attended for endoscopy at the same time (14.00 hours) after an overnight fast. Prior to endoscopy, a 10 ml sample of venous blood was withdrawn into a lithium heparin tube. Antral biopsies were taken for H. pylori culture and urease test, along with two biopsies from antrum and corpus for histological assessment. If at least two out of these three tests were positive then patients were deemed H. pylori positive, and H. pylori negative if all three were negative. Control patients were also attending for investigation of dyspepsia but had all tests negative for H. pylori and histologically normal gastric mucosa. Further biopsies were taken from all patients as follows: two from the antrum for luminol-enhanced chemiluminescence, one from the antrum for malondialdehyde assessment, and one from both antrum and corpus for ascorbic acid and vitamin C determination.

H. pylori-positive patients were randomized on a 1:1:1:1 basis to receive one of the following treatments: (1) colloidal bismuth subcitrate (120 mg) plus tetracycline (500 mg), both four times per day, plus metronidazole (400 mg) three times per day (triple therapy), followed by ascorbic acid (200 mg) plus vitamin E, α-tocopherol (50 mg), both twice per day; (2) triple therapy followed by vitamin placebo; (3) triple therapy placebo followed by vitamins C and E; or (4) triple therapy placebo followed by vitamin placebo. In all cases, antibiotic (triple) therapy or its placebo was for 2 weeks followed by vitamin supplementation or its placebo for 4 weeks. Thus, patients received triple therapy and antioxidant vitamins (triple–vitamins), triple therapy alone (triple–placebo), vitamins alone (placebo–vitamins), or neither (placebo–placebo). Consecutive patients were allocated treatment randomly according to a list kept in pharmacy. Placebo tablets (lactose) were identical in appearance to the study drugs and all drugs were dispensed by the pharmacy to maintain investigator blinding. At the end of the 6-week treatment period, patients returned for repeat endoscopy and sampling. Patients were instructed not to take tablets on the day of endoscopy. The study was approved by the Local Research Ethics Committee and all patients gave written informed consent.

Histology
Biopsies were fixed in 10% buffered formalin. Sequential 3 μm thick sections were cut and stained with haematoxylin and eosin and modified Giemsa’s stain. Gastritis was scored from 0–3 for acute inflammation, chronic inflammation, atrophy, intestinal metaplasia and H. pylori infection density according to the updated Sydney classification by a single blinded histopathologist (Dixon et al. 1996).

Helicobacter pylori culture
A single biopsy was immediately transferred to brain–heart infusion broth and incubated microaerophilically at 37°C on a VPAT plate (vancomycin, polymyxine B, amphotericin, trimethoprim) for 8 d. Plates were inspected for H. pylori at 4, 6 and 8 d.

Luminol-enhanced chemiluminescence
Chemiluminescence probes such as luminol and lucigenin react with ROS to form 3-aminophthalate and N-methyl-acridone. The excited electrons in these compounds relax with the emission of energy as light which can be detected by a scintillation counter. Luminol has been found to be a more sensitive chemiluminescent probe than lucigenin in colonic and gastric biopsies (Simmonds et al. 1992; Davies et al. 1994). The reaction of H₂O₂ and hypochlorite with luminol is catalysed by neutrophil-derived myeloperoxidase; inhibition of this enzyme by sodium azide leads to maximal reduction in chemiluminescence suggesting that these are the most important metabolites detected by this technique. Consistent with this, chemiluminescence results correlate with acute inflammatory scores in mucosal biopsies. Chemiluminescence is also inhibited by catalase and dimethyl superoxide suggesting detection of H₂O₂ and hydroxyl radicals respectively. Thus, luminol was selected for the present study as it is a suitable probe to detect neutrophil-derived ROS in gastric biopsies (Drake et al. 1998).

Two antral biopsies from each patient were transferred immediately into pre-oxygenated PBS (pH 7.4) with added glucose (5 mM), M and C, and assayed within 3 h. Prior to weighing, each biopsy was added to a pre-counted scintillation vial containing 1 ml 75 μM luminol solution and counted for 5 min in a liquid scintillation counter set in
Malondialdehyde

For each patient, one antral biopsy was snap-frozen in liquid N₂ at the time of endoscopy, stored at −70°C, and assayed within 2 weeks by a modification of the procedure of Yagi (1976). Following thawing, blotting and weighing, each biopsy was immersed in 4 ml water. Thiobarbituric acid solution (1 ml), made by dissolving 0.167 g thiobarbituric acid solution in 50 ml water—glacial acetic acid (1:1, v/v), was then added. A set of malondialdehyde standards was also freshly prepared and added to 1 ml thiobarbituric acid solution prepared in the same way. All mixtures were heated at 100°C for 60 min. After cooling on ice, 5 ml butan-1-ol was added to extract the malondialdehyde equivalents. The tubes were centrifuged at 1100 g for 10 min to separate the aqueous and butan-1-ol phases. Fluorescence of the butan-1-ol phase at 555 nm was determined using an excitation wavelength of 515 nm. Values from the tissue specimens were compared with the standard solutions. The within-subject CV of this technique is 16.8% (based on paired biopsies from 208 patients).

It should be noted that there can be difficulties in the interpretation of the results of this assay. First, aldehydes other than malondialdehyde produced in lipid peroxidation can be detected. Second, much of the malondialdehyde measured is generated by decomposition of lipid hydroperoxides during the acid-heating stage of the assay. In addition, other compounds such as bilirubin, sugars and amino acids can also be reactive towards thiobarbituric acid. Finally, malondialdehyde detected in this assay may also be generated as a byproduct of cyclooxygenase activity in platelets (Emma et al. 2000). However, the first two of these processes reflect other pathways in lipid peroxidation and suggest that, whilst measuring thiobarbituric acid reactive substances in this assay is not specific for malondialdehyde levels in vivo, it is reflective of lipid peroxidation. This is supported by previous studies that have shown significant correlations between malondialdehyde concentrations measured by this assay and both chemiluminescence and its own DNA adduct, malondialdehyde deoxyguanosine (Drake et al. 1998; Everett et al. 2001). Thus, although non-specific, results from this assay complement the chemiluminescence results and together these assays provide estimations of neutrophil-derived ROS and consequent lipid peroxidation.

Ascorbic acid and total vitamin C measurements

Venous blood samples were centrifuged. One 0.5 ml aliquot of plasma was added to 1 ml metaphosphoric acid (20 ml/l). A second 0.5 ml aliquot was added to 9 mg dithiothreitol prior to addition of 1·0 ml of metaphosphoric acid (20 ml/l) for reduction of dehydroascorbic acid to ascorbic acid and subsequent analysis of total vitamin C. All samples were snap-frozen in liquid N₂ and stored at −70°C. Prior to analysis, samples were thawed and centrifuged at 1000 g. The supernatant fraction was analysed by HPLC using reversed-phase ion pair chromatography on a C₁₈ column (Sobala et al. 1991). Ascorbic acid was selectively measured using an electrochemical detector set at 350 mV. Total vitamin C (the sum of ascorbic acid and dehydro-ascorbic acid) was determined from the solutions initially prepared with dithiothreitol after incubation at 45°C for 120 min.

Biopsy samples were frozen in liquid N₂ immediately after endoscopy, stored at −70°C and assayed within 2 weeks. After thawing, biopsies were homogenized in 0.5–1.0 ml metaphosphoric acid and divided into two parts. For total vitamin C determination, dithiothreitol was added to one part to a final concentration of 6 mg/ml and incubated at 45°C for 120 min prior to analysis. HPLC was then performed for both parts as described for plasma. The within-subject CV for both ascorbic acid and total vitamin C was 5%.

Statistical analysis

Data for most variables are skewed, and in some instances normality could not be achieved by log transformation. For consistency, therefore, data are presented as median values with interquartile ranges and non-parametric statistics are used (with the exception of linear regression). For patients receiving triple therapy, results presented are for those in whom H. pylori was successfully eradicated, and for patients receiving placebo triple therapy, results presented are for those who remained H. pylori positive at follow-up. The Wilcoxon signed-ranks test was used to compare paired pre- and post-treatment data for the randomized groups. Mann-Whitney U tests were used for comparison of unpaired data and Kruskall-Wallis test was used for comparison of more than one group. Spearman’s ρ was calculated for correlations. To calculate the effect of multiple variables on plasma ascorbic acid, multiple linear regression was used with log conversion of the plasma ascorbic acid to maintain normality and constant variance; data are presented as regression slope (B) with 95% CI. Two-tailed P values were calculated in all cases; a value <0.05 is taken to imply statistical significance. Calculations were performed using SPSS for Windows 8.0 (SPSS UK Ltd, Woking, Surrey).

Results

Patients

H. pylori-positive patients (n 117) were randomized to treatment. Of these, ninety-eight patients returned for the second visit. Control patients with normal gastric histology and negative H. pylori tests (n 63) were also studied. Sex, age, smoking, alcohol intake, use of non-steroidal anti-inflammatory drugs and acid suppressive therapy were recorded prior to the first endoscopy. Patients with normal
Helicobacter pylori gastritis, and the four randomized groups
(Mean values and ranges)

<table>
<thead>
<tr>
<th></th>
<th>Controls</th>
<th>All</th>
<th>Triple+placebo</th>
<th>Placebo+vitamin†</th>
<th>Placebo+placebo</th>
<th>Triple*+placebo</th>
<th>Placebo+vitamins†</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean</td>
<td>63</td>
<td>58</td>
<td>30</td>
<td>29</td>
<td>29</td>
<td>30</td>
<td>29</td>
</tr>
<tr>
<td>Range</td>
<td>117</td>
<td>29–66</td>
<td>49</td>
<td>29–65</td>
<td>49</td>
<td>53</td>
<td>29–65</td>
</tr>
<tr>
<td>Age (years)</td>
<td>38</td>
<td>17–75</td>
<td>49***</td>
<td>17–75</td>
<td>52</td>
<td>17–75</td>
<td>48</td>
</tr>
<tr>
<td>Sex (% males)</td>
<td>60</td>
<td>35 7</td>
<td>15 5</td>
<td>8 13</td>
<td>15</td>
<td>5</td>
<td>8</td>
</tr>
<tr>
<td>Smokers (%)‡</td>
<td>27</td>
<td>15 7</td>
<td>15 7</td>
<td>15 7</td>
<td>24</td>
<td>2</td>
<td>24</td>
</tr>
<tr>
<td>NSAID (%)§</td>
<td>9</td>
<td>46 47</td>
<td>47 13</td>
<td>47 47</td>
<td>47</td>
<td>47</td>
<td>47</td>
</tr>
<tr>
<td>Acid suppression (%)</td>
<td>26</td>
<td>18 17</td>
<td>20 6</td>
<td>20 6</td>
<td>20</td>
<td>20</td>
<td>20</td>
</tr>
</tbody>
</table>

**NSAID, non-steroidal anti-inflammatory drugs.**

* Triple: eradication therapy comprising colloidal bismuth subcitrate, tetracycline and metronidazole for 2 weeks.
† Vitamin E for 4 weeks.
‡ Daily smoker of cigarettes or cigars.
§ Intake of NSAID, within the previous month.

There were no significant differences in baseline measurements of malondialdehyde, chemiluminescence and plasma, antral and corpus ascorbic acid and total vitamin C in the four randomized groups (Table 1).

Antral chemiluminescence and malondialdehyde were higher in *H. pylori*-negative patients than uninfected patients (P<0.001 for both), reflecting increased ROS activity. Malondialdehyde levels correlated significantly with chemiluminescence (r 0.2, P=0.009).

Plasma ascorbic acid was significantly lower in *H. pylori*-positive patients than in *H. pylori*-negative patients (P=0.02) and the percentage plasma vitamin C in the reduced ascorbic acid form was also significantly lower in *H. pylori*-positive than *H. pylori*-negative patients (89.9 v. 95.3 %, P=0.006). There were no differences in antral and corpus ascorbic acid levels between *H. pylori*-negative and -positive patients. However, ascorbic acid and total vitamin C were both significantly higher in the antrum than the corpus. This was true both for *H. pylori*-positive (584.8 v. 356.0 μmol/kg, P<0.001 for ascorbic acid and 444.6 μmol/kg, P<0.001 for total vitamin C) and *H. pylori*-negative patients (451.4 v. 340.1 μmol/kg, P<0.001 for ascorbic acid and 572.9 v. 392.9 μmol/kg, P<0.001 for vitamin C). There was a significant positive correlation between chemiluminescence and antral ascorbic acid (r 0.19, P=0.02) and total vitamin C (r 0.17, P=0.03). No such correlation existed between malondialdehyde and either ascorbic acid or total vitamin C.

Chemiluminescence and malondialdehyde were not affected by patient sex, smoking history (smokers v. non-smokers), intake of non-steroidal anti-inflammatory drugs or use of acid-suppressive therapy. Chemiluminescence correlated significantly with age (r 0.27, P<0.001). This correlation was not evident, however, when analysis was limited to *H. pylori*-positive patients (r 0.02, P=0.8), suggesting that this is a result of confounding by the older age of *H. pylori*-positive patients.

Plasma ascorbic acid (16.5 v. 47.1 μmol/l, P=0.02), total plasma vitamin C (18.2 v. 50.5 μmol/l, P<0.001), antral ascorbic acid (403.7 v. 633.1 μmol/kg, P=0.007) and total antral vitamin C (538.3 v. 687.0 μmol/kg, P=0.01) were all lower in smokers than non-smokers, and plasma ascorbic acid (30.7 v. 44.9 μmol/l, P=0.03) and total plasma vitamin C (35.8 v. 47.7 μmol/l, P=0.05) were both lower in males than females. Furthermore, plasma ascorbic acid and total plasma vitamin C correlated negatively with age (r −0.21, P=0.008 and r −0.21, P=0.007 respectively). There were no other significant correlations between age and measures of ascorbic acid or total vitamin C. In order to determine which of age, sex, smoking and *H. pylori* infection were most important in determining plasma ascorbic acid concentrations a multiple linear regression model was constructed, with log-converted plasma ascorbic acid as the dependent factor. Age (B = −0.07, 95 % CI −0.01,
Table 2. Pretreatment chemiluminescence, malondialdehyde, plasma, antral and corpus ascorbic acid in patients with normal gastric histology, Helicobacter pylori gastritis, and the four ran-

Helicobacter pylori-positive groups.* (Median values and interquartile ranges)

<table>
<thead>
<tr>
<th></th>
<th>Controls (n=63)</th>
<th>Placebo—vitamins (n=29)</th>
<th>Triple—vitamins (n=30)</th>
<th>Placebo—placebo (n=11)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chemiluminescence</td>
<td>296–2834</td>
<td>460–748</td>
<td>389–494</td>
<td>436–619</td>
</tr>
<tr>
<td>Malondialdehyde</td>
<td>78–156</td>
<td>85–142</td>
<td>86–145</td>
<td>86–145</td>
</tr>
<tr>
<td>Plasma ascorbic acid</td>
<td>114–242</td>
<td>114–242</td>
<td>114–242</td>
<td>114–242</td>
</tr>
<tr>
<td>Antral ascorbic acid</td>
<td>43–239</td>
<td>32–292</td>
<td>114–242</td>
<td>114–242</td>
</tr>
</tbody>
</table>

*For details of subjects and procedures, see Table 1 and p. 6.

Six-week follow-up

The results of the follow-up at 6 weeks are summarized in Table 3. Of the initial 117 H. pylori-positive patients, ninety-eight (84%) returned and could be evaluated. Of the fifty-nine patients who received eradication therapy, forty-nine returned and thirty-six were H. pylori-negative after 6 weeks (eradication rate 73.5%). Four patients were indeterminate for H. pylori at 6 weeks and were excluded from analysis (all were H. pylori-positive on histology but negative on urease test and culture). H. pylori was eradicated in one patient in the placebo–vitamins group. There was no evidence that this patient had taken any antibiotics but high-dose vitamin C has been shown to result in eradication of H. pylori (Jarosz et al. 1998). This patient was also excluded from analysis. For all of the ensuing data, no significant changes were seen in the placebo–placebo group.

Plasma ascorbic acid and vitamin C

Plasma ascorbic acid and total vitamin C approximately doubled in both groups of patients that received vitamin supplements (Table 4). There were no changes in plasma ascorbic acid or total vitamin C in either group not receiving vitamin supplements and H. pylori eradication had no effect on plasma vitamin levels. There was, however, a significant increase in the percentage plasma vitamin C as ascorbic acid in the triple–vitamins group (86.6 to 93.3%, P=0.03), whereas no such change was seen in the placebo–vitamins group.

Tissue vitamin C and ascorbic acid

There was a significant increase in total antral vitamin C and smaller (non-significant) increases in antral ascorbic acid, corpus ascorbic acid and total corpus vitamin C in the triple–vitamins group (Table 4). In the placebo–vitamins group there were significant increases in corpus ascorbic acid and total corpus vitamin C along with smaller (non-significant) increases in antral ascorbic acid and total antral vitamin C. When all patients receiving vitamins are...
combined, regardless of eradication therapy, there were significant increases after treatment in antral ascorbic acid (586.5 to 695.6 μmol/kg, *P*=0.03), total antral vitamin C (654.7 to 824.4 μmol/kg, *P*=0.01), corpus ascorbic acid (315.7 to 529.2 μmol/kg, *P*=0.001), and total corpus vitamin C (405.4 to 617.2 μmol/kg, *P*=0.003).

Chemiluminescence and malondialdehyde
Chemiluminescence fell in 18/19 patients in the triple–vitamins group and in 15/17 in the triple–placebo group (Table 4). The difference between pre- and post-treatment chemiluminescence was significant for both groups (*P*<0.001). In the placebo–vitamins group, chemiluminescence fell in ten but increased in thirteen patients. Although there was a small reduction in median chemiluminescence after treatment (from 23,993 to 9699 cpm/mg) this reduction was much smaller in magnitude than that seen for the *H. pylori* eradication groups and was not significant (*P*=0.6). In the placebo–placebo group chemiluminescence fell in eleven and increased in ten patients (*P*=1.0).

Malondialdehyde fell in 13/18 patients in the triple–placebo group. The difference was significant (*P*=0.03). In the triple–vitamins group malondialdehyde levels fell in 13/16 but the difference between pre- and post-treatment values was not significant (*P*=0.1). In the placebo–vitamins group, malondialdehyde fell in fifteen but increased in eight (*P*=0.4) and in the placebo–placebo group it fell in ten and increased in twelve patients (*P*=0.7). Thus, vitamin supplements alone did not significantly reduce either chemiluminescence or malondialdehyde, and had no additional effect to *H. pylori* eradication.

**Discussion**

There has been considerable interest in the potential for antioxidant vitamins to reduce gastric cancer risk in *H. pylori* infection by attenuating oxidative stress and oxidative DNA damage. However, the precise role of ascorbic acid and other antioxidants in *vivo* remains uncertain. Thus, vitamin C supplements have been shown to reduce gastric mucosal adduct levels in one study (Dyke *et al.* 1994) whereas in another, patients given supplements of α-tocopherol and β-carotene were more likely to develop bronchial carcinoma than unsupplemented patients (The Alpha-Tocopherol, Beta-Carotene Cancer Prevention Study Group, 1994; Omenn *et al.* 1996). Furthermore, vitamin C and E supplements have been shown to reduce oxidative stress in peripheral lymphocytes and abrogate Fe-induced oxidative stress in the rat intestine (Brennan *et al.* 2000; Srirgridhar & Nair, 2000). There is a need, therefore, to clarify the antioxidant effects of vitamin supplements in the gastric mucosa before advocating this approach.

This is the first study to have randomized *H. pylori*-infected patients to vitamin C and E supplements and eradication therapy, alone and in combination, and examine the effect of these treatments on ROS activity in gastric biopsies. There was no specific determination of compliance but plasma ascorbic acid more than doubled in the vitamin-supplemented patients to concentrations that are considered to be the plateau for ascorbic acid. This is good evidence that the tablets were taken largely as prescribed, and that the dose of ascorbic acid, at seven times the recommended daily intake, was sufficient for the purposes of the study (Levine *et al.* 1996).

Our baseline data closely reflect previous studies, some from our own department (Sobala *et al.* 1989; Banerjee *et al.* 1994; Davies *et al.* 1994; Farinati *et al.* 1996; Drake *et al.* 1998). ROS and malondialdehyde levels were elevated in *H. pylori* infection and malondialdehyde concentrations correlated significantly with chemiluminescence. Ascorbic acid and total vitamin C were highly concentrated in the mucosa, and levels were not affected by *H. pylori* infection. Plasma ascorbic acid, total vitamin C, and the percentage plasma vitamin C as ascorbic acid were all significantly diminished in *H. pylori*-positive patients compared with controls. This latter finding is in contrast with previous work and probably reflects the larger number of patients in our present study (Phull *et al.* 1998). It is unlikely, however, that the reduction in plasma ascorbic acid is a direct effect of *H. pylori* infection since levels did not alter after eradication of the organism. A lack of independent effect was confirmed by multiple linear regression. It is more likely that the difference reflects different dietary intake or other factors such as socio-economic status. This is reflected by the lower proportion of smokers in the control group, and is consistent with studies of nutritional factors and *H. pylori* infection rates in Colombian children (Goodman *et al.* 1997).

Eradication of *H. pylori* resulted in expected reductions in mucosal chemiluminescence and malondialdehyde concentrations, in line with previous studies (Farinati *et al.* 1996; Waring *et al.* 1996; Drake *et al.* 1998). Vitamin supplements, either alone or in concert with *H. pylori* eradication, led to increases in tissue and plasma ascorbic acid and total vitamin C concentrations. However, supplements had a negligible effect on tissue chemiluminescence and malondialdehyde. It is possible that a small effect of vitamin supplements on ROS

**Table 3. Follow-up of randomized patients**

<table>
<thead>
<tr>
<th></th>
<th>Triple†+vitamins‡</th>
<th>Placebo+vitamins‡</th>
<th>Triple†+placebo</th>
<th>Placebo+placebo</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Randomized (<em>n</em>)</td>
<td>29</td>
<td>29</td>
<td>30</td>
<td>29</td>
<td>117</td>
</tr>
<tr>
<td>Returned at 6 weeks (<em>n</em>)</td>
<td>24</td>
<td>26</td>
<td>25</td>
<td>23</td>
<td>98</td>
</tr>
<tr>
<td><em>H. pylori</em> negative at 6 weeks (<em>n</em>)</td>
<td>19</td>
<td>1</td>
<td>17</td>
<td>0</td>
<td>37</td>
</tr>
<tr>
<td>Indeterminate for <em>H. pylori</em> at 6 weeks (<em>n</em>)§</td>
<td>1</td>
<td>0</td>
<td>2</td>
<td>0</td>
<td>4</td>
</tr>
</tbody>
</table>

* For details of subjects and procedures, see Table 1 and p. 4.
† Triple: *H. pylori* eradication therapy comprising colloidal bismuth subcitrate, tetracycline and metronidazole for 2 weeks.
‡ Vitamins: ascorbic acid and α-tocopherol for 4 weeks.
§ All were *H. pylori* positive on histology but negative on urease test and culture and were excluded from analysis.
Table 4. Paired plasma, antral and corpus ascorbic acid and total vitamin C, chemiluminescence and malondialdehyde for patients with *Helicobacter pylori* gastritis, before and 6 weeks after treatment* 

(Median values and interquartile ranges)

<table>
<thead>
<tr>
<th>Time (weeks)</th>
<th>Triple†+vitamins‡ (n 19)§</th>
<th>Placebo+vitamins‡ (n 24)‖</th>
<th>Triple†+placebo (n 17)§</th>
<th>Placebo+placebo (n 23)‖</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Median (Interquartile range)</td>
<td>Median (Interquartile range)</td>
<td>Median (Interquartile range)</td>
<td>Median (Interquartile range)</td>
</tr>
<tr>
<td>Plasma ascorbic acid (μmol/l)</td>
<td>29.8 (10.8–44.3)</td>
<td>65.0 (53.4–90.8)</td>
<td>36.3 (11.4–61.9)</td>
<td>74.9 (58.5–81.8)</td>
</tr>
<tr>
<td>Plasma total vitamin C (μmol/l)</td>
<td>39.7 (13.6–48.3)</td>
<td>73.2 (56.2–94.8)</td>
<td>40.3 (14.2–68.1)</td>
<td>78.9 (69.8–85.7)</td>
</tr>
<tr>
<td>Antral ascorbic acid (μmol/kg)</td>
<td>546.2 (266.3–739.8)</td>
<td>691.6 (418.5–973.2)</td>
<td>647.9 (343.5–855.7)</td>
<td>699.5 (496.3–1071.4)</td>
</tr>
<tr>
<td>Antral total Vitamin C (μmol/kg)</td>
<td>568.9 (371.9–853.4)</td>
<td>760.9 (530.3–1151.5)</td>
<td>719.4 (445.7–995.4)</td>
<td>854.0 (607.5–1188.4)</td>
</tr>
<tr>
<td>Corpus ascorbic acid (μmol/kg)</td>
<td>405.4 (168.6–534.3)</td>
<td>473.5 (316.3–670.0)</td>
<td>304.3 (189.1–472.4)</td>
<td>541.1 (241.9–794.4)</td>
</tr>
<tr>
<td>Corpus total vitamin C (μmol/kg)</td>
<td>480.4 (290.7–725.1)</td>
<td>562.7 (440.6–813.7)</td>
<td>403.1 (270.8–652.4)</td>
<td>663.2 (353.2–988.5)</td>
</tr>
<tr>
<td>Chemiluminescence (c.p.m./mg)</td>
<td>17.316 (11.270–106.386)</td>
<td>684.3 (374–1264)</td>
<td>23.993 (6698.5–5452)</td>
<td>9699 (1899–31820)</td>
</tr>
<tr>
<td>Malondialdehyde (μmol/kg)</td>
<td>97.7 (77.3–140.5)</td>
<td>89.8 (68.7–131.3)</td>
<td>124.5 (97.9–141.9)</td>
<td>115.7 (98.6–113.1)</td>
</tr>
</tbody>
</table>

* For details of subjects and procedures, see Table 1 and p. 4.
† Triple: *H. pylori* eradication therapy comprising colloidal bismuth subcitrate, tetracycline and metronidazole for 2 weeks.
‡ Vitamins: ascorbic acid and α-tocopherol for 4 weeks.
§ For patients receiving triple therapy, data are for those in whom *H. pylori* was successfully eradicated.
‖ For patients receiving placebo triple therapy, data are for those who remained *H. pylori* positive.

Median values were significantly different from paired pre-treatment values (wilcoxon signed ranks test): ††P=0.04, †‡P=0.03, §§P=0.02, **P=0.01, †††P=0.004, ***P=0.001. All other paired comparisons were non-significant.
activity has been missed due to the relatively small numbers in each group and the large CV for chemiluminescence. It is clear from the results, however, that any effect of vitamins is far less than eradication of H. pylori. The reduction in chemiluminescence after treatment of the infection was several orders of magnitude greater than that of vitamin supplements and it would be difficult to demonstrate an effect over and above this. Likewise, the results relating to malondialdehyde indicate that the most effective treatment is H. pylori eradication and vitamin supplements do not add to this effect.

The failure of antioxidant vitamin supplements to significantly effect levels of ROS is not surprising, because, even in the inflamed stomach, ascorbic acid is highly concentrated into the mucosa. Concentrations of the ascorbyl radical are increased in the mucosa of H. pylori gastritis, but its concentration remains considerably lower than ascorbic acid (Drake et al. 1996). This lends support to an antioxidant role for ascorbic acid but suggests that, even in the presence of continued inflammation, the scavenging capability of ascorbic acid is replete. Supplements are only likely to be of use when ascorbic acid stores are significantly diminished.

A second consideration is that ascorbic acid can act not only as antioxidant but also, in the presence of transition metals, as pro-oxidant (Halliwell & Gutteridge, 1998). In keeping with this, and consistent with earlier work, we found that antral mucosal ascorbic acid had a weak positive correlation with chemiluminescence (Drake et al. 1998; Everett et al. 2001). Alternative explanations for this relationship include an influx of ascorbic acid into the mucosa in response to oxidative stress or as a consequence of the rich content of ascorbic acid found in leukocytes. However, the possibility of ascorbic acid acting as pro-oxidant in vivo suggests that, depending on the circumstances, vitamin supplements might have either a beneficial or a detrimental effect (Herbert, 1994).

It is worthy of comment that epidemiological data mainly support a protective effect of high fruit and vegetable intake rather than antioxidants per se. Our results do not conflict with this observation, which may result from the presence of other compounds such as folic acid or the balance between different micronutrients in fruits and vegetables (La Vecchia et al. 1994). Furthermore, the results of the present study do not rule out the possibility that antioxidant vitamin supplements may prevent the development of gastritis. Indeed it has already been observed that diets high in vitamin C may prevent infection with H. pylori (Goodman et al. 1997; Jarosz et al. 1998). Vitamin supplements could also have a beneficial effect on the formation of N-nitroso compounds in gastric juice. However, although it is tempting to speculate that longer and larger doses of antioxidants may yet be shown to be beneficial, the potential for harm is also present. Not only is ascorbic acid potentially pro-oxidant but recent in vivo work has also shown that vitamin C may mediate formation of genotoxins from lipid hydroperoxides in the absence of transition metal ions (Lee et al. 2001). These issues may only be settled by large population studies or using direct measurements of DNA damage.

In conclusion, we have found that dietary supplementation of antioxidant vitamin supplements do not reduce ROS activity in the gastric mucosa. Oxidative stress, and hence oxidative DNA damage, in the gastric mucosa are only likely to be reduced if H. pylori is eradicated, regardless of dietary manipulation.

Acknowledgements

This study was supported by a grant from the Ivy Hobson Trust for Cancer Research.

References


