The efficacy of ferrous bisglycinate and electrolytic iron as fortificants in bread in iron-deficient school children

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Fe deficiency is the most common nutritional deficiency in both the industrialised and developing worlds, affecting mostly infants, children, and women of childbearing age. The fortification of staple foods is an important recognised long-term strategy for addressing micronutrient deficiencies, including that of Fe (Cook & Reusser, 1983; Hurrell, 1997). Finding the ideal Fe fortification compound, however, remains a challenge. In the present study the effect of ferrous bisglycinate as fortificant in brown bread was compared with that of electrolytic Fe among Fe-deficient school children in a randomised controlled trial. Children (n 160), aged 6–11 years, with serum ferritin <20 μg/l, were randomly assigned to one of three treatment categories: (i) standard unfortified bread; (ii) bread with electrolytic Fe as fortificant; and (iii) bread with ferrous bisglycinate as fortificant. Each child received four slices of bread (120 g) on school days, which supplied an average of 3·66 mg elemental Fe per intervention day for 137 d (2·52 mg/d for 75 d and 5·04 mg/d for 62 d) over a period of 7·5 months. Hb, serum ferritin, serum Fe and transferrin saturation were measured at baseline and at the end of the intervention. Significant treatment effects were observed for Hb (P=0·013), serum Fe (P=0·041) and transferrin saturation (P=0·042) in the ferrous bisglycinate group, but not in the electrolytic Fe group. There were no significant intervention effects for serum ferritin in either treatment group. Overall, ferrous bisglycinate as Fe fortificant in brown bread performed better than electrolytic Fe in a group of Fe-deficient school children over a period of 7·5 months.

Iron fortification: Ferrous bisglycinate: Electrolytic iron: Fortified bread: School children

Fe has also been selected as Fe fortificant in the South African national food fortification programme for the fortification of maize meal and wheat flour (Department of Health, 2003). Cereal flours are often used as vehicles for micronutrient fortification, because they are widely consumed and are usually centrally processed. Unfortunately, the large amounts of phytic acid present in cereal flours can have a potent inhibitory effect on the absorption of Fe (Hallberg et al. 1989). One way of countering this inhibitory effect is to add an Fe absorption enhancer, such as ascorbic acid, to the fortified product (Siegenberg et al. 2003). Ascorbic acid, however, is easily oxidised, and expensive sophisticated packaging may be required to prevent its degradation during storage. It is also destroyed to a great extent during food preparation, and its use as enhancer in cereal flours is therefore not always feasible (Hurrell, 2002).

An alternative solution is to use an Fe fortificant in which the Fe is protected from the effects of absorption inhibitors, e.g. chelated Fe compounds, and thus eliminating the need for ascorbic acid. Ferrous bisglycinate is an amino acid chelate which is formed by the binding of two molecules of glycine to one Fe2+ atom (Allen, 2002), and has a bioavailability two to four times that of ferrous sulfate (Olivares et al. 1997; Bovell-Benjamin et al. 2000; Layrisse et al. 2000). Because of its relatively low interaction with food (Layrisse et al. 2000), it is also less likely to cause sensory changes in the food vehicle. Ferrous bisglycinate has been acknowledged by the US Food and Drug Administration as being Generally Regarded As Safe for use as Fe fortificant in foods (Jeppsen, 2001).

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To date, there are very few controlled trials establishing the efficacy of either electrolytic Fe or ferrous bisglycinate in human populations, and certainly no studies comparing the effect of electrolytic Fe with that of ferrous bisglycinate. The aim of the present study was to compare the effect of ferrous bisglycinate, used as a fortificant in brown bread, with the effect of electrolytic Fe on the Fe and Hb status of Fe-deficient primary-school children in a randomised controlled trial. Because response to an Fe intervention may be affected by the vitamin A status of a population (Van Stuijvenberg et al. 1997), serum retinol was also measured.

Methods
Study population and design
The study was conducted in the Northern Cape, South Africa, in a primary school that is 400 km from Cape Town and serves a community that is of low socio-economic status. Of the 482 grade 1–3 children who were screened, 161 children with serum ferritin concentrations <20 μg/l were selected to take part in the study. One child with Hb <90 g/l was, for ethical reasons, not randomised and referred to the local clinic for treatment (Fig. 1). Of the 160 children participating in the study, 71.9 % were Fe-deficient (serum ferritin <15 μg/l), 7.5 % anaemic (Hb <115 g/l) and less than 1 % were vitamin A-deficient (serum retinol <20 μg/dl). Based on the serum ferritin results of a previous Fe fortification study in primary-school children (Van Stuijvenberg et al. 1999), it was calculated that if only children with low Fe stores were included in the study, and a common standard deviation of 15 μg/l for serum ferritin were assumed, a minimum sample size of forty-seven per group would be required to detect a difference in response in serum ferritin between the active and control groups, at a 5 % significance level, with 80 % power. The study was approved by the Ethics Committee of the South African Medical Research Council, and permission was obtained from the Northern Cape Department of Education and the headmaster of the school. Written informed consent was obtained from the parents or guardians of all participants.

Subjects were stratified by school grade and then randomly assigned to one of three groups, using random number tables. These groups were then randomly assigned to three different treatment categories: (i) a group receiving standard unfortified brown bread (n 53); (ii) a group receiving fortified brown bread using electrolytic Fe as the Fe fortificant (n 53); and (iii) a group receiving fortified brown bread using ferrous bisglycinate as Fe fortificant (n 54). The flour used in the bread had an extraction rate of 88 %. The three types of bread were similar in macro-nutrient composition, taste and appearance. To optimise compliance and to avoid cross-contamination between groups, the bread was delivered at the school in colour-coded (i.e. red, blue and yellow) containers; each loaf (pre-sliced by machine) was individually packed in a plastic sleeve and marked with the appropriate colour-coded sticker; sandwiches (spread with peanut butter and jam) were prepared at colour-coded tables and then distributed to the classrooms in smaller containers that were also colour-coded. Each child received four slices of bread (120 g) per school day over two meal periods. The bread was eaten under supervision of the class teacher and compliance was recorded daily, using colour-coded record sheets for the respective treatment groups. The bread provided as part of the study replaced the two slices of bread the children normally received via the school feeding system. Blood was drawn at baseline and again after 7·5 months of intervention. Children were dewormed 3 weeks prior to the baseline measurements being taken (500 mg mebendazole) and again 6 months later. All measurements were done blind and only the project leader was aware of group allocation (single blind study).

Fortification
Fortification of the wheat flour was done by an independent pharmaceutical manufacturing company (Zedchem (Pty) Ltd, Cape Town, South Africa). The flour was fortified with Fe, Zn, vitamin A, thiamine, riboflavin, niacin, pyridoxine and folic acid at levels that were in line with those prescribed by the national food fortification programme of the South African Department of Health (Table 1). At the time of the study, mandatory fortification of wheat flour had not yet come into effect. The four slices of fortified bread supplied 2.52 mg elemental Fe (35 mg Fe/kg flour), in addition to Fe naturally present in bread. Due to time constraints, the study was limited to 7·5 months instead of the envisaged 12 months. In order to compensate for the shorter trial period, the level of fortificants in both Fe groups was doubled after the first 4·5 months of the trial (Table 1), and for the remaining 3 months the four slices of bread supplied 5·04 mg of elemental Fe, over and above the Fe naturally present in bread. No intervention took place during school holidays, on weekends or public holidays; the bread was provided for a total of 137 school days over the 7·5-month intervention period, supplying a total of 501·5 mg elemental Fe: 2·52 mg/d for 7·5 d and 5·04 mg/d for 62 d, i.e. an average of 3·66 mg Fe per intervention day. The electrolytic Fe (particle size <45 μm; 325 mesh), as well as the other micronutrients, was supplied by Roche Vitamins South Africa (Pty) Ltd, Johannesburg, South Africa and the ferrous bisglycinate (Ferrochel®) by Albion Laboratories, Inc., Clearfield, UT, USA. The control wheat flour was not fortified with Fe or any of the other micronutrients. The three types of flour were packaged in colour-coded flour bags and sent to a local baker, who was...
contracted to bake and deliver the bread to the school on a daily basis during the school week.

Quality control
To enhance quality control, the procedures followed by the baker, especially in terms of keeping the three types of flour and bread apart throughout the baking process, were observed and recorded in detail by a member of the research team, who visited the bakery unannounced twice during the intervention period. In addition, bread samples were sent to the Council for Scientific and Industrial Research ( Pretoria, South Africa) once a month for analysis of Fe content (Table 2).

Measurements
Biochemical indices. Blood (5 ml) was obtained by antecubital venepuncture. All blood samples were collected between 08.30 and 13.00 hours, and the three treatment groups were distributed evenly over this period. Care was taken to protect the blood from direct sunlight. Hb was measured in the field by means of the direct cyanmethaemoglobin method, using Drabkin’s solution and a standard photometer. The rest of the blood was centrifuged and the serum stored at −80°C until analysed. Serum ferritin was determined by an immunoradiometric assay (Ferritin MAb Solid Phase Component System; ICN Pharmaceuticals, Orangeburg, NY, USA), using an Auto Gamma 500C counting system (United Technologies Packard, IL, USA). Serum Fe and transferrin were determined spectrophotometrically with a Technicon RA-1000 automated system, using colorimetric and turbidimetric methods from Bayer Corporation (Tarrytown, NY, USA), respectively; these values were then used to calculate transferrin saturation. C-reactive protein was determined with a turbidimetric method from Bayer Corporation and measured spectrophotometrically with a Technicon RA-1000 automated system. Serum retinol was determined (under dimmed light) by reversed-phase HPLC, based on the method described by Catignani and Bieri (1983).

Anthropometry. Weight was measured (in light clothing) to the nearest 0.05 kg, and height (without shoes) to the nearest 0.1 cm. Height-for-age, weight-for-age and weight-for-height were expressed as Z scores, using the National Center for Health Statistics median as reference (Hamill et al. 1979). The birth date of each child was obtained from the school register.

Statistical analyses
Data were analysed using the SPSS for Windows program (SPSS Inc., Chicago, IL, USA, version 11.0). Hb data were analysed on an intention-to-treat basis. For serum ferritin, serum Fe and transferrin saturation, however, five children with infection (C-reactive protein >10 mg/l) at baseline or follow-up were excluded from the analyses. The paired t test was used to compare pre- and post-intervention values within each treatment group. To estimate treatment effects an ANOVA was done on the measurement after the intervention period, as well as an analysis of covariance using baseline measurement and gender as covariates. Values of P<0.05 were considered statistically significant. Because only children with low serum ferritin concentrations (<20 μg/l) were included in the study, serum ferritin values were normally distributed and log transformation before analysis was therefore not necessary.

Results
Of the 160 children who were randomised for treatment, 153 completed the trial. Reasons for dropping out were leaving the area (n=4), failure to obtain blood from the child at the follow-up assessment (n=1), withdrawal of parental consent (n=1)
and a follow-up Hb value <90 g/l (n 1). The trial profile is given in Fig. 1. Mean compliance (defined as the actual number of bread slices consumed, expressed as a percentage of the total number of slices provided over the trial period) was 96·5 %, 95·7 % and 95·3 % in the control, electrolytic Fe and ferrous bisglycinate groups, respectively. Absence from school was the main reason for non-compliance.

Baseline characteristics of the control and intervention groups are given in Table 3. The three groups were similar with regard to age, height, weight, Fe, Hb and vitamin A status. There were slightly more boys in the control group than in the other two groups. Almost 30 % of the children were stunted or underweight, yet very few were anaemic (7·5 %) and almost none were vitamin A-deficient (<1 %). The mean serum retinol of the study population was 35·5 (sd 7·6) µg/dl.

**Intervention effects**

The Hb concentrations in the control and two intervention groups at baseline and after 7·5 months of intervention are shown in Table 4. There was a significant increase from baseline to follow-up in the ferrous bisglycinate group only, with the intervention effect being significant when gender and the baseline Hb measurement were adjusted for. The increase in Hb in the ferrous bisglycinate group was also significant compared with the electrolytic Fe group, and the intervention effect four times that in the electrolytic Fe group. Table 5 shows the serum Fe and transferrin saturation values before and after the intervention. Significant treatment effects for both serum Fe and transferrin saturation were observed in the ferrous bisglycinate group, while the electrolytic Fe group showed no significant response. Serum ferritin concentrations at the beginning and end of the intervention period are shown in Table 6. There was no significant intervention effect for serum ferritin in either the ferrous bisglycinate or electrolytic Fe group, and neither when gender and serum ferritin at baseline were adjusted for.

**Discussion**

The choice of Fe compound in a fortification programme will determine whether or not the Fe-deficient sector of a population will benefit from that programme. Trials validating the efficacy of different Fe fortification compounds are therefore important, and should be a step prior to the implementation of such programmes. Results of the present randomised controlled trial showed that ferrous bisglycinate as fortificant in brown bread performed better than electrolytic Fe, in terms of improved Hb and circulating Fe levels, in a group of Fe-deficient school children over a period of 7·5 months. Significant intervention effects were observed for Hb, serum Fe and transferrin saturation in the ferrous bisglycinate group, while the electrolytic Fe group showed no significant intervention effects for any of the variables measured.

Finding the ideal Fe fortification compound is not easy and usually represents a compromise between factors such as bioavailability, potential for causing organoleptic problems and cost (Dury, 2002). While ferrous bisglycinate has a bioavailability two to four times that of ferrous sulfate (Olivares et al. 1997; Bovell-Benjamin et al. 2000; Layrisse et al. 2000), it is expensive. A recent cost analysis carried out by a SUSTAIN Task Group showed the cost of ferrous bisglycinate, taking bioavailability into account, to be between 7·2 and 18·5 times that of ferrous sulfate (Moore et al. 2004). Wider usage and increased competition in future may, however, reduce cost.

Furthermore, concerns had been raised regarding the tendency of ferrous bisglycinate to cause lipid peroxidation and rancidity in maize meal (Bovell-Benjamin et al. 1999a). These changes can, however, be prevented by the addition of an antioxidant (Bovell-Benjamin et al. 1999a), and it has been shown that ferrous bisglycinate-fortified maize meal porridge was acceptable to both toddlers and their mothers, with or without the addition of the antioxidant (Bovell-Benjamin et al. 1999b). In the present study, no obvious organoleptic changes were noticed in the bread fortified with ferrous bisglycinate; the bread looked and tasted the same as the control and electrolytic Fe-fortified breads. However, sensory tests were not carried out and the fortified flour was not stored for longer than 1 month at a time. Further investigation into the stability of wheat flour fortified with ferrous bisglycinate is therefore needed.

Electrolytic Fe, while being relatively inexpensive (costing half as much as ferrous sulfate for the same amount of Fe; Hurrell, 1997) and causing few organoleptic problems, has an estimated bioavailability of approximately only half that of ferrous sulfate. This estimate is based on Hb repletion studies in rats (Hurrell et al. 1999).
and there are very few studies evaluating its effectiveness as fortificant in human populations. In the present study, no significant treatment effects were observed in the group consuming the bread fortified with electrolytic Fe for 137 d over a period of 7-5 months. The average amount of Fe ingested per intervention day was equivalent to the amount that would have been taken in if five to six slices of bread, fortified according to the South African food fortification regulations, had been consumed. The South African government based their decision to use electrolytic Fe in its fortification programme on the guidelines issued by SUSTAIN (2001a), which were interim recommendations and based on the information available at the time. SUSTAIN has since launched a comprehensive evaluation of the different elemental Fe powders in use today, and human efficacy trials based on the information available at the time. SUSTAIN has since launched a comprehensive evaluation of the different elemental Fe powders in use today, and human efficacy trials based on the information available at the time. SUSTAIN has since launched a comprehensive evaluation of the different elemental Fe powders in use today, and human efficacy trials based on the information available at the time.

Table 4. Hb concentrations before and after 7-5 months of intervention
(Mean values and standard deviations except for intervention effects)

<table>
<thead>
<tr>
<th></th>
<th>Control (n 51)</th>
<th>Electrolytic Fe (n 53)</th>
<th>Ferrous bisglycinate (n 49)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean</td>
<td>SD</td>
<td>Mean</td>
</tr>
<tr>
<td></td>
<td>Hb (g/l)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baseline</td>
<td>126·8</td>
<td>7·6</td>
<td>125·9</td>
</tr>
<tr>
<td></td>
<td>7-5 months</td>
<td>127·5</td>
<td>7·4</td>
</tr>
<tr>
<td>Change‡</td>
<td>0·7</td>
<td>5·3</td>
<td>0·2</td>
</tr>
<tr>
<td>Intervention effect¶</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Crude</td>
<td>–0·4 (–3·3, 2·5)</td>
<td></td>
<td>1·6 (–1·2, 4·7)</td>
</tr>
<tr>
<td>Adjusted for gender and baseline measurement</td>
<td>0·6 (–1·5, 2·6)</td>
<td></td>
<td>2·6 (0·6, 4·6)†</td>
</tr>
</tbody>
</table>

* Mean change from baseline to follow-up.
† Intervention effect — difference (95 % CI) in estimated marginal means at follow-up between intervention and control group.
‡ Mean value was significantly different compared with baseline (paired t test): $P_{\text{adj}}=0·001$.
¶ Significant effect compared with control (analysis of covariance): $P_{\text{adj}}=0·013$.
† Significant effect compared with electrolytic Fe (analysis of covariance): $P_{\text{adj}}=0·043$.

Table 5. Serum iron and transferrin saturation before and after 7-5 months of intervention
(Mean values and standard deviations except for intervention effects)

<table>
<thead>
<tr>
<th></th>
<th>Control (n 50)</th>
<th>Electrolytic Fe (n 53)</th>
<th>Ferrous bisglycinate (n 45)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean</td>
<td>SD</td>
<td>Mean</td>
</tr>
<tr>
<td>Serum Fe (µmol/l)*</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baseline</td>
<td>9·91</td>
<td>3·42</td>
<td>9·76</td>
</tr>
<tr>
<td>7-5 months</td>
<td>9·43</td>
<td>2·91</td>
<td>9·87</td>
</tr>
<tr>
<td>Change†</td>
<td>–0·48</td>
<td>4·10</td>
<td>0·11</td>
</tr>
<tr>
<td>Intervention effect¶</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Crude</td>
<td>0·44 (–0·93, 1·82)</td>
<td></td>
<td>1·49 (0·05, 2·92)§</td>
</tr>
<tr>
<td>Adjusted for baseline measurement</td>
<td>0·46 (–0·93, 1·84)</td>
<td></td>
<td>1·50 (0·06, 2·94)‖</td>
</tr>
<tr>
<td>Transferrin saturation (%)*</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baseline</td>
<td>15·43</td>
<td>5·79</td>
<td>15·84</td>
</tr>
<tr>
<td>7-5 months</td>
<td>14·70</td>
<td>5·04</td>
<td>16·19</td>
</tr>
<tr>
<td>Change</td>
<td>–0·73</td>
<td>6·71</td>
<td>0·35</td>
</tr>
<tr>
<td>Intervention effect¶</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Crude</td>
<td>1·49 (–0·84, 3·81)</td>
<td></td>
<td>2·59 (0·17, 5·02)†</td>
</tr>
<tr>
<td>Adjusted for baseline measurement</td>
<td>1·40 (–0·90, 3·69)</td>
<td></td>
<td>2·48 (0·09, 4·87)‖</td>
</tr>
</tbody>
</table>

* Children with C-reactive protein ≥10 mg/l at baseline or follow-up excluded.
† Mean change from baseline to follow-up.
‡ Intervention effect — difference (95 % CI) in estimated marginal means at follow-up between intervention and control group.
§ Mean value was significantly different compared with baseline (paired t test): $P_{\text{adj}}=0·043$.
|| Significant effect compared with control (analysis of covariance): $P_{\text{adj}}=0·041$.
† Significant effect compared with electrolytic Fe (analysis of covariance): $P_{\text{adj}}=0·036$.
** Significant effect compared with control (analysis of covariance): $P_{\text{adj}}=0·042$.
Ferrous bisglycinate v. electrolytic iron

The bioavailability of ferrous bisglycinate has never been compared directly with that of electrolytic Fe. Extrapolating from studies comparing the bioavailability of either ferrous bisglycinate or electrolytic Fe with that of ferrous sulfate, however, it can be deduced that ferrous bisglycinate is absorbed four to eight times better than electrolytic Fe (Olivares et al. 1997; Bovell-Benjamin et al. 2000; Layrisse et al. 2000; Hurrell et al. 2002). In our study the intervention effect in terms of Hb for ferrous bisglycinate was four times that of electrolytic Fe.

Ferrous bisglycinate, because of its high bioavailability and relative low reactivity, especially in milk products (Allen, 2002), appears to be an ideal Fe fortificant. Ferrous bisglycinate also has the advantage in that it is considered a ‘natural’ product (Hurrell, 2002). In addition, amino acid chelates have been reported to come down and the issues around organoleptic problems are resolved. Another Fe chelate that can be considered an attractive option for fortification programmes is NaFeEDTA. Its potential as fortificant has been confirmed in foods, especially in milk products, making this compound an attractive option for fortification. Its current high cost at this stage may, however, limit its usefulness at programme level, and alternative options that are as bioavailable but less expensive, e.g. NaFeEDTA, should also be explored.

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Table 6. Serum ferritin before and after 7.5 months of intervention

<table>
<thead>
<tr>
<th></th>
<th>Control (n 50)</th>
<th>Electolytic Fe (n 53)</th>
<th>Ferrous bisglycinate (n 45)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serum ferritin (µg/l)*</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baseline</td>
<td>11.29</td>
<td>11.66</td>
<td>10.90</td>
</tr>
<tr>
<td>7.5 months</td>
<td>11.72</td>
<td>13.22</td>
<td>12.18</td>
</tr>
<tr>
<td>Change†</td>
<td>0.43</td>
<td>1.56</td>
<td>1.28</td>
</tr>
<tr>
<td>Intervention effect‡</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Crude</td>
<td>1.50 (–0.57, 3.56)</td>
<td>0.46 (–1.69, 2.61)</td>
<td></td>
</tr>
<tr>
<td>Adjusted for gender and baseline measurement</td>
<td>1.45 (–0.62, 3.53)</td>
<td>0.63 (–1.53, 2.78)</td>
<td></td>
</tr>
</tbody>
</table>

* Children with C-reactive protein ≥ 10 mg/l at baseline or follow-up excluded.
† Mean change from baseline to follow-up.
‡ Intervention effect = difference (95% CI) in estimated marginal means at follow-up between intervention and control group.

References


