Respective contributions of intestinal Niemann-Pick C1-like 1 and scavenger receptor class B type I to cholesterol and tocopherol uptake: in vivo v. in vitro studies

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Abstract

The intestinal absorption of cholesterol and lipid micronutrients such as vitamin E has been shown to share some common pathways. The present study aims to further compare the uptake of cholesterol ([3H]cholesterol v. 22-N-(7-nitrobenz-2-oxa-1,3-diazol-4-yl)amino)-23,24-bisnor-5-cholen-3-ol (NBD-cholesterol)) and tocopherol in Caco-2 TC-7 cells and in mouse intestine, with special focus on the respective roles of scavenger receptor class B type I (SR-BI) and Niemann-Pick C1-like 1 (NPC1L1). Conversely to NBD-cholesterol, the uptakes of [3H]cholesterol and tocopherol by Caco-2 cells were impaired by both block lipid transport-1 and ezetimibe, which inhibit SR-BI and NPC1L1, respectively. These inhibitions occurred only when cholesterol or tocopherol was delivered to cells included in micelles that contained biliary acid and at least oleic acid as a lipid. In vivo, after 2 h of digestion in mice, the uptake of the two cholesterol analogues and of tocopherol all showed distinct patterns along the duodenum–jejunum axis. [3H]Cholesterol uptake, which correlated closely to NPC1L1 mRNA expression in wild-type (wt) mice, was strongly inhibited by ezetimibe. Intestinal SR-BI overexpression did not change NPC1L1 expression and led to a significant increase in [3H]cholesterol uptake in the distal jejunum. Conversely, neither ezetimibe treatment nor SR-BI overexpression had an effect on NBD-cholesterol uptake. However, in contrast with SR-BI mRNA expression, tocopherol absorption increased strongly up to the distal jejunum in wt mice where it was specifically inhibited by ezetimibe, and was increased in the proximal intestine of intestinal SR-BI-overexpressing mice. Thus, cholesterol and tocopherol uptakes share common pathways in cell culture models, but display different in vivo absorption patterns associated with distinct contributions of SR-BI and NPC1L1.

Key words: Vitamin E; Cholesterol; Intestinal absorption; Membrane transporters

Over the last few years, there have been significant advances in our understanding of the mechanisms governing the intestinal absorption of dietary lipids and lipid micronutrients. Intestinal cholesterol absorption was initially thought to occur by passive diffusion. It is now well acknowledged that micellar cholesterol uptake by the enterocyte depends mainly on Niemann-Pick C1-like 1 (NPC1L1) activity(1,2), although the molecular mechanisms by which micellar cholesterol crosses the brush-border membrane remain unknown. Other membrane transporters, such as scavenger receptor class B type I (SR-BI), have also been identified, to a lesser extent, as playing roles in cholesterol uptake(3,4) and lipid sensing(5) in the enterocyte. The roles of NPC1L1 and SR-BI in the intestine are not limited to cholesterol transport. It has recently been shown

Abbreviations: BLT1, block lipid transport 1; DMEM, Dulbecco’s minimum essential medium; iSR-BI tg, intestinal scavenger receptor class B type I-overexpressing transgenic; NBD, 22-N-(7-nitrobenz-2-oxa-1,3-diazol-4-yl)amino)-23,24-bisnor-5-cholen-3-ol; NPC1L1, Niemann-Pick C1-like 1; SR-BI, scavenger receptor class B type I; wt, wild type.

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that both proteins were involved in the uptake of micellar vitamin E\(^{6,7}\), but their relative involvement in tocopherol or cholesterol uptake is not known.

However, note that some of the above observations vary strongly depending on model and experimental conditions used. In particular, \textit{in vitro} and \textit{in vivo} data are sometimes conflicting. For example, although it appears clear that SR-BI is involved in cholesterol uptake in brush-border membrane vesicles\(^8\), there are still doubts over its \textit{in vivo} involvement, as the distribution of the cholesterol absorption along the small intestine in mice fed with a basal diet is not different in SR-BI\(^{-/-}\) or wild-type (wt) animals\(^9\). Conversely, although cholesterol uptake in brush-border membrane vesicles prepared from small intestines of wt and NPC1L1\(^{-/-}\) mice has been reported as equivalent\(^{10}\), NPC1L1 visibly has a clear \textit{in vivo} contribution\(^{10}\). Indeed, both wt mice treated with ezetimibe and NPC1L1-deficient mice displayed an inhibition from 50 to 70 % of cholesterol absorption. Residual pathways independent of NPC1L1 and ezetimibe are thus responsible for absorption of the remaining 30 % of cholesterol\(^1\) and the totality of fluorescent 22-ylamino)-23,24-bisnor-5-cholen-3-ol (NBD)-cholesterol\(^{(11)}\).

NBD-cholesterol and tocopherol, both used to characterise and compare the uptake of [3H]cholesterol, cholesterol and vitamin E uptake, the aim of the present study was to investigate the effects of inhibition of SR-BI or NPC1L1 in Caco-2 cells in \textit{in vitro} and \textit{in vivo}. The simplest M1 micelles contained 5 mM of taurocholate\(^{(15)}\) in DMEM associated with a mixture of 5 mM-cholesterol, 7.4 kBq/ml \(^{[3]H}\)cholesterol (1.48 MBq/\(\mu\)mol) and 10 mM-NBD-cholesterol, or 10 mM-\(\alpha\)-tocopherol. The M2 micelles contained oleic acid (0.5 mM). The M3 micelles also contained oleic acid (0.5 mM) and other lipids usually found during digestion process. Appropriate volumes of each lipid stock solution were dried under a stream of \(N_2\). The lipids were first resuspended in 208 mM of DMEM containing 24 mM-taurocholate, mixed for 2 min with a vortex to form the micelles and then diluted to 1 ml in a solution of DMEM and 5 % lipoprotein-deficient serum (prepared as previously described\(^{13}\) with minor changes) at the final concentrations presented in Table 1. The simplest M1 micelles contained 5 mM of taurocholate in DMEM associated with a mixture of 5 mM-cholesterol, 7.4 kBq/ml \(^{[3]H}\)cholesterol (1.48 MBq/\(\mu\)mol) and 10 mM-NBD-cholesterol, or 10 mM-\(\alpha\)-tocopherol. The M2 micelles contained oleic acid (0.5 mM). The M3 micelles also contained oleic acid (0.5 mM) and other lipids usually found during digestion process. Appropriate volumes of each lipid stock solution were dried under a stream of \(N_2\). The lipids were first resuspended in 208 mM of DMEM containing 24 mM-taurocholate, mixed for 2 min with a vortex to form the micelles and then diluted to 1 ml in a solution of DMEM and 5 % lipoprotein-deficient serum to reach 5 mM final of taurocholate\(^{(15)}\).

Cell culture

Preparation of micelles. Cholesterol or tocopherol was delivered to the cells in three different types of micelles in Dulbecco’s minimum essential medium (DMEM) containing 5 % lipoprotein-deficient serum (prepared as previously described\(^{13}\) with minor changes) at the final concentrations presented in Table 1. The simplest M1 micelles contained 5 mM of taurocholate in DMEM associated with a mixture of 5 mM-cholesterol, 7.4 kBq/ml \(^{[3]H}\)cholesterol (1.48 MBq/\(\mu\)mol) and 10 mM-NBD-cholesterol, or 10 mM-\(\alpha\)-tocopherol. The M2 micelles contained oleic acid (0.5 mM). The M3 micelles also contained oleic acid (0.5 mM) and other lipids usually found during digestion process. Appropriate volumes of each lipid stock solution were dried under a stream of \(N_2\). The lipids were first resuspended in 208 mM of DMEM containing 24 mM-taurocholate, mixed for 2 min with a vortex to form the micelles and then diluted to 1 ml in a solution of DMEM and 5 % lipoprotein-deficient serum to reach 5 mM final of taurocholate\(^{(15)}\).

Culture conditions

Caco-2 cell line, clone TC-\(7^\text{16}\), was a gift from Dr M. Roussset (UMRS 872, Villejuif, France). Cells were cultured in T-75 flasks in DMEM supplemented with 10 % heat-inactivated fetal bovine serum, 1 % non-essential amino acid and 1 % antibiotics (complete medium), as previously described\(^{17}\).

Uptake experiments

Cells were seeded in twenty-four-well plates at a density of 20 000 cells per well, and then differentiated for 20–21 d.

\begin{table}[h]
\centering
\begin{tabular}{|c|c|c|c|}
\hline
Lipids & Micelles no. 1 (M1, \(\mu\)l) & Micelles no. 2 (M2, \(\mu\)l) & Micelles no. 3 (M3, \(\mu\)l) \\
\hline
Taurocholate & 5 & 5 & 5 \\
Oleic acid & – & 0.5 & 0.5 \\
Phosphatidylcholine & – & – & 0.04 \\
Lysophosphatidylcholine & – & – & 0.16 \\
Monoolein & – & – & 0.3 \\
\hline
\end{tabular}
\caption{Lipid composition of the three types of micelles used in cell experiments\(^*\)}
\end{table}

\(^*\) Micelles were prepared as described in Experimental methods. Each type of micelle contained a mixture of 5 mM-cholesterol, 10 mM-NBD-cholesterol and 744 kBq \(^{[3]H}\)-cholesterol (1.48 MBq/\(\mu\)mol of total cholesterol) or 10 mM-\(\alpha\)-tocopherol.
Cells were pre-incubated for 16 h in serum-free medium, and then for 1 h in medium containing 5% lipoprotein-deficient serum supplemented with dimethyl sulphoxide (control), 10 µM-BLT1 or 100 µM-ezetimibe. For lipid absorption measurement, cells were finally incubated for 1 h with different micelles supplemented or not with the inhibitors. After incubation, the culture medium was harvested, the cells were washed twice in 500 µl PBS containing 5 mM-taurocholate, then lysed in 500 µl PBS containing 0·25% SDS. Protein concentrations were estimated on aliquots of cell suspensions using a bicinchoninic acid kit (Pierce, Montluçon, France).

**Cholesterol and 2R,4′R,8′R-γ-tocopherol uptake in mice**

All animal studies were performed in conformity with the Public Health Service Policy on Human Care and Use of Laboratory Animals and in accordance with the local ethics committee of the Toulouse animal facility platform (Anexplo).

**Preparation of emulsions**

For cholesterol and tocopherol delivery to mice, emulsions were prepared as described previously with minor changes. Briefly, for each mouse, 100 µl of 9% NaCl was added to 100 µl of maize oil containing a mixture of 30 µg of cholesterol, 10 kBq of [3H]cholesterol (0·118 MBq/µg) and 5 µg of NBD-cholesterol, or 5 mg of α-tocopherol. Emulsions were formed by bath sonication (Branson 3510; Branson, Danbury, CT, USA) for 15 min at 0°C, and were freshly used for force feeding. γ-Tocopherol was chosen instead of α-tocopherol as (i) it shows no discrimination in absorption and (ii) because γ-tocopherol, in contrast to α-tocopherol, is not detectable in the intestine by HPLC before oral administration.

**Animals**

Mice overexpressing SR-BI in the intestine were generated as previously described. C57BL/6 J (wt) mice were purchased from Janvier (Le-Genest-St-Isle, France). They were housed in a temperature-, humidity- and light-controlled room. They were given a standard chow diet with water ad libitum and a guard column. The mobile phase was 100% methanol. Flow rate was 1·5 ml/min, and the column was kept at a constant temperature (30°C). The HPLC system comprised a Dionex separation module (P680 HPLC Pump and ASI100 Automated Sample Injector; Dionex, Aix-en-Provence, France) and a Jasco fluorometric detector (Jasco, Nantes, France). Tocopherols were detected at 325 nm after light emission at 292 nm, and identified by retention time compared with pure (≥95%) standards. Quantification was performed using Chromeleon software (version 6·50 SP4 Build 1000; Dionex, Aix-en-Provence, France) comparing peak area with standard reference curves. All solvents used were of HPLC grade from SDS (Pepsin, France).

**Tocopherol extraction and HPLC analysis**

Tocopherol was extracted from 500 µl aqueous samples using tocol as an internal standard, as previously described. Tocopherol and tocol were then separated using a 250 × 4·6-mm RP C18, 5 µm Zorbax column (Interchim, Montluçon, France) and a guard column. The mobile phase was 100% methanol. Flow rate was 1·5 ml/min, and the column was kept at a constant temperature (30°C). The HPLC system comprised a Dionex separation module (P680 HPLC Pump and ASI100 Automated Sample Injector; Dionex, Aix-en-Provence, France) and a Jasco fluorometric detector (Jasco, Nantes, France). Tocopherols were detected at 325 nm after light emission at 292 nm, and identified by retention time compared with pure (≥95%) standards. Quantification was performed using Chromeleon software (version 6·50 SP4 Build 1000; Dionex, Aix-en-Provence, France) comparing peak area with standard reference curves. All solvents used were of HPLC grade from SDS (Pepsin, France).

**RNA extraction and quantitative real-time RT-PCR**

Total RNA was isolated from the four different fragments of intestine using the TRIzol method (Invitrogen, Cergy-Pontoise, France). Complementary DNA was reverse-transcribed from 2 µg of total RNA and diluted to 20 ng/µl using sterilised water. Quantitative real-time RT-PCR was performed on 12·5 ng of complementary DNA in a final volume of 12·5 µl, using Power SYBR Green PCR Master Mix and an ABI-Prism 7000 thermal cycler (Applied Biosystems, Villebon-sur-Yvette, France) under standard PCR conditions. Primer sequences are reported in Table 2. All data were normalised to cyclophilin expression levels and were analysed using LinRegPCR.

**Table 2. Oligonucleotide sequences for quantitative real-time PCR**

<table>
<thead>
<tr>
<th>Gene</th>
<th>NCBI Refseq</th>
<th>Forward primer (5′–3′)</th>
<th>Reverse primer (5′–3′)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cyclophilin</td>
<td>NM_011149</td>
<td>tggagagcaccacaagaagacacaagaca</td>
<td>fgccggtctgagcataagat</td>
</tr>
<tr>
<td>SR-BI</td>
<td>NM_016741</td>
<td>tctcaatcataatcgtcaagggcat</td>
<td>acggctgcagatctccgctgt</td>
</tr>
<tr>
<td>NPC1L1</td>
<td>NM_207242</td>
<td>gacagacaccaacattgctggagt</td>
<td>aacgctcaggattgctggaga</td>
</tr>
</tbody>
</table>

SR-BI, scavenger receptor class B type 1; NPC1L1, Niemann-Pick C1-like 1.
Membrane preparation and protein analysis

The mice were killed and the intestine was removed and cut into four fragments, as described above. Each section was opened and the mucosa was scraped, immediately frozen in liquid N2 and stored at −80°C. Tissues were then homogenised at 4°C in PBS, as previously described(4). Proteins were determined using a bicinchoninic acid kit, and 50 μg were used for western blot analysis. Proteins were separated by SDS-PAGE using 12% gels under reducing conditions according to Laemmli(20) and were electrophoretically transferred to polyvinylidene fluoride membranes according to Burnette(21). The blotting membrane was incubated with rabbit polyclonal anti-SR-BI IgG (NB-400-104; Novus Biologicals, Littleton, CO, USA) or with a 1:500 dilution of rabbit serum against NPC1L1 generously donated by Dr Helen Hobbs. For visualisation, monoclonal anti-rabbit IgG (Sigma) were used as secondary antibodies at 1:5000 dilution.

Statistical analysis

Differences between more than two groups of unpaired data were tested by the non-parametric Kruskal–Wallis test. The non-parametric Mann–Whitney U test was used as a post hoc test when the Kruskal–Wallis test showed significant differences between groups. Differences between only two groups of unpaired data were tested by the Mann–Whitney U test. Values of P<0·05 were considered to be significant. All statistical analyses were performed using Statview software version 5.0 (SAS Institute, Cary, NC, USA).

Results

Effect of micellar composition on [3H]- and NBD-cholesterol or tocopherol uptake via scavenger receptor class B type I and Niemann-Pick C1-like 1 in Caco-2 cells

We first investigated the role of SR-BI and NPC1L1 in the uptake of cholesterol associated with three different types of micelles (Table 1) by Caco-2 cells. When compared to M1 micelles, adding oleic acid to taurocholate (M2 micelles) significantly increased the uptake of both [3H]- and NBD-cholesterol (Fig. 1(a) and (b)). More complex micelles (M3) induced a further increase in the uptake of both cholesterol analogs. Inhibition of SR-BI (BLT1, 10 μM) or NPC1L1 (ezetimibe, 100 μM) had no effect on M1 micelle-mediated uptake or NBD-cholesterol uptake in M2 or M3 micelles, but impaired [3H]cholesterol uptake in M2 (∼30% inhibition) and M3 (50% inhibition) micelles. This indicated that the uptake of [3H]cholesterol and not NBD-cholesterol, which was associated with micelles containing at least oleic acid, was partially dependent on SR-BI or NPC1L1. The effects of the inhibitors were not additive, suggesting similar routing pathways for both proteins.

When the experiment was performed with tocopherol (Fig. 1(c)), we again observed a significantly increased (although less pronounced) uptake among M1, M2 and M3 micelles. The inhibition of SR-BI or NPC1L1 did not affect M1 micelle-mediated uptake, but strongly decreased tocopherol uptake in M2 and very strongly decreased tocopherol uptake in M3 micelles (M2, −27 and −46% and M3, −55 to −77% for ezetimibe and BLT1, respectively). BLT1 and ezetimibe has no additive effect.

In vivo uptake of [3H]- and NBD-cholesterol or tocopherol in wild-type mice and transgenic mice overexpressing scavenger receptor class B type I in the intestine

We investigated in vivo the intestinal absorption of cholesterol and tocopherol in wt C57BL/6j mice treated or not with ezetimibe and in a mouse model of intestinal overexpression of SR-BI that we have previously developed(4). We first quantified the expression of SR-BI and NPC1L1 mRNA in four different fragments of mouse intestine (duodenum and proximal, medium and distal jejunum) from wt or iSR-BI tg mice (Fig. 2). SR-BI mRNA and protein expression decreased along the duodenum–jejunum axis in wt mice, as already described(4), but was strongly increased in all intestinal fragments of iSR-BI tg mice (Fig. 2(a)). This mRNA expression was similar to our previous observations on total intestine (25-fold overexpression). In contrast, NPC1L1 mRNA and protein expression slightly increased from duodenum to medium jejunum in wt mice, but were not modified by intestinal SR-BI overexpression (Fig. 2(b)).

To focus on the intestinal absorption process, we then analysed the uptake of [3H]-NBD-cholesterol or tocopherol at 2 h after feeding in wt mice, which is a time long enough to obtain significant uptake values without significant secretion. Indeed, as the half-time of gastric emptying after a liquid meal is about 15 min in mice(22), 2 h after oral administration were sufficient to allow mixed micelles to form, make contact with both the duodenal and jejunal mucosa and be taken up by enterocytes. [3H]-Cholesterol displayed an increased uptake from duodenum to medium jejunum, which was well correlated to NPC1L1 mRNA expression (Fig. 3(a)). Conversely, the uptake of NBD-cholesterol was similar in the first three duodenal and jejunal fragments and decreased by 30% in the distal fragment (Fig. 3(d)). In contrast, the uptake profile of tocopherol strongly increased along the duodenum–distal-jejunum axis (16-fold increase between the first and the last fragment (Fig. 3(g)), partly in agreement with the increased NPC1L1 mRNA expression, but inversely correlated to SR-BI mRNA expression (Fig. 2).

Cross-comparing uptake values between wt and iSR-BI tg mice revealed a significant increase in the incorporation of [3H]cholesterol in the distal jejunum of iSR-BI tg mice (Fig. 3(b)), in agreement with the stronger overexpression of SR-BI mRNA in the distal jejunum (150-fold increase) than in the duodenum (16-fold increase). NBD-cholesterol uptake remained unchanged in the iSR-BI tg mice (Fig. 3(c)), whereas tocopherol uptake was significantly increased in the duodenum (3.2-fold) and proximal jejunum (2.4-fold) in iSR-BI tg mice compared to wt mice (Fig. 3(h)), indicating a major difference in the absorption of these lipid molecules related to the intestinal overexpression of SR-BI.
Finally, ezetimibe treatment of wt mice strongly decreased the absorption of \([3H]\)cholesterol (by over 50%) in all four intestinal fragments (Fig. 3(c)). This suggested an active participation of NPC1L1 in \([3H]\)cholesterol absorption in the duodenum and jejunum. Ezetimibe also inhibited tocopherol absorption, but only in the distal jejunum, where it is mainly absorbed in control wt mice (Fig. 3(i)). In contrast, NBD-cholesterol uptake was not inhibited by ezetimibe, but slightly increased by up to 20% in duodenum (Fig. 3(f)).

**Discussion**

Understanding the molecular mechanisms of lipid absorption by the enterocyte is a critical challenge, as both cholesterol (one-third of total body cholesterol comes from diet) and fat-soluble vitamins may be involved. The objectives of the present study were to compare the absorption of two lipid nutrients, i.e. cholesterol (\([3H]\)- or NBD-analogues) and tocopherol, by the enterocyte *in vitro* and *in vivo*, and to consider the relative involvement of two major membrane lipid transporters, i.e. NPC1L1 and SR-BI, in this process. The Caco-2 cell line is a widely used human intestinal *in vitro* model giving reproducible results that usually correlate closely with human *in vivo* figures\(^\text{[23]}\). This cell model has been extensively used to study the absorption of both cholesterol\(^\text{[24–26]}\) and tocopherol\(^\text{[7,27,28]}\). The clone TC-7 was chosen specifically because it is more homogenous than the parent Caco-2 cell line\(^\text{[29]}\) and because it shows good viability after apical incubation with mixed micelles\(^\text{[16]}\). A key step appears to be related to the composition of the lipid donors interacting with uptake efficiency\(^\text{[12,13,15,25,30,51]}\). Here, we used three types of micelles as donor vehicles. The simplest M1 micelles contained only the bile salt taurocholate. More conventional M2 micelles contained, in addition, 10% oleic acid. Therefore, the M3 micelles mimicking the mixed micelles found in dietary conditions\(^\text{[17]}\) were M2 micelles.
Cholesterol and vitamin E intestinal uptake

Fig. 2. Scavenger receptor class B type I (SR-BI) and Niemann-Pick C1-like 1 (NPC1L1) expression along the duodenal–jejunal axis in wild-type (wt) and transgenic (tg) mice overexpressing SR-BI in the intestine. Relative expression profiles of (a) SR-BI and (b) NPC1L1 mRNA were analysed by real-time quantitative RT-PCR in wt and intestinal SR-BI-overexpressing tg mice (iSR-BI tg) in the duodenum (4 cm) and in proximal (P, 4 cm), medium (M, 6 cm) and distal (D, 6 cm) jejunum fragments. Values were normalised to cyclophilin expression and are expressed as mean values with their standard errors represented by vertical bars, n = 4 for each group. * Mean values were significantly different from those of wt intestinal fragments (P < 0.05). Insets: Western blots of protein expression profiles in different intestinal fragments ((1) duodenum, (2) proximal jejunum, (3) medium jejunum, (4) distal jejunum).

added with three lipids (2-oleoyl-1-palmitoyl-sn-glycero-3-phosphocholine, 1-palmitoyl-sn-glycero-3-phosphocholine and monoolein). We clearly demonstrated that specific absorption of [3H]cholesterol or tocopherol is mediated by SR-BI and NPC1L1, respectively, in the presence of oleic acid, SR-BI and NPC1L1 seem to be involved in a common absorption pathway that ensures both cholesterol and tocopherol uptake.

In conclusion, this Caco-2 cell study shows that, in the presence of oleic acid, SR-BI and NPC1L1 seem to be involved in a common absorption pathway that ensures both cholesterol and tocopherol uptake.

In vivo studies were then conducted to pinpoint the respective and physiological contributions of SR-BI and NPC1L1 to cholesterol and tocopherol absorption, using wt mice, iSR-BI tg mice or wt mice treated with ezetimibe. SR-BI mRNA over-expression was found in all four intestinal fragments, as already observed in rats(1) and human subjects(26). This differs from the inhibition of SR-BI expression observed in NPC1L1-knockdown Caco-2 cells(31).
In contrast to what was observed on Caco-2 cells, the two cholesterol analogues, especially [3H]cholesterol, displayed very distinct uptake profiles compared to tocopherol in wt mice as well as in ezetimibe-treated or iSR-BI tg mice. [3H]-cholesterol uptake increased progressively along the duodenal–jejunal axis and decreased in the distal jejunum, in correlation with NPC1L1 expression. NBD-cholesterol uptake was similar in the duodenum and the proximal and medium jejunum, and slightly decreased in the distal jejunum fragment. In contrast, tocopherol uptake increased very sharply along the intestinal axis. None of the profiles correlated to SR-BI expression in wt mice. Ezetimibe treatment led to a drastic decrease of [3H]cholesterol uptake. These results confirm the major in vivo involvement of NPC1L1 in this process. The significant increase in [3H]cholesterol uptake, specifically in the distal jejenum in our iSR-BI tg mice, is in agreement with other reports suggesting a moderate collateral role of scavenger receptors in cholesterol absorption and with the increased absorption of cholesterol that we had previously observed in this model. This increase was actually more pronounced at 4 h after oral administration, i.e. during digestion and after transport and secretion into the blood.

Fig. 3. Comparison of [3H], 22-(N-(7-nitrobenz-2-oxa-1,3-diazol-4-yl)amino)-23,24-bisnor-5-cholen-3-ol (NBD)-cholesterol and tocopherol uptakes along the duodenal–jejunal axis of mouse intestine in wild-type (wt), scavenger receptor class B type I-overexpressing transgenic (iSR-BI tg) and ezetimibe-treated mice. Mice were force-fed with either [3H]- and NBD-cholesterol- or γ-tocopherol-enriched emulsions. After 2 h, the first 20 cm of the mouse intestine were harvested downstream of the biliary duct, rinsed and cut into four fragments: the duodenum (4 cm), and proximal (P, 4 cm), medium (M, 6 cm) and distal (D, 6 cm) jejunum. Results are expressed in nmol of compound/cm of each intestinal fragment, given as mean values with their standard errors represented by vertical bars. * Mean values were significantly different from those of control (wt) fragment (P ≤ 0.05). (a, d, g) wt mice (n 9); (b, e, h) wt (v) iSR-BI tg mice (g); n 4–6/group; (c, f, i) wt mice (i) treated with ezetimibe (g), n 4–6/group. (a–c) [3H]Cholesterol; (d–f) NBD-cholesterol and (g–i) γ-tocopherol.
Conversely, tocopherol absorption showed a different pattern to [3H]-cholesterol absorption in different mice models. Ezetimibe treatment had a small effect on vitamin E absorption in the proximal to medium intestine, but induced a significant 30% decrease in the distal jejunum compared to wt mice, in agreement with the maximum uptake and the high expression of NPC1L1 observed in this fragment. This confirms that NPC1L1 is also a contributor to in vitro tocopherol absorption. These findings are consistent with a previous study that reported ezetimibe-sensitive absorption of tocopherol by rat intestine(7). The overexpression of SR-BI led to a significant apparent increase in \( \gamma \)-tocopherol uptake in the proximal part of the intestine, in accordance with our previous data(6). However, when compared to the basal uptake values in wt mice (8 and 10 \( \mu \)g/cm for duodenum and proximal jejunum, respectively; Fig. 3(c)), these 3.2- and 24.4-fold increases in iSR-BI \( \text{tg} \) correspond to 25.6 and 24.4 \( \mu \)g/cm for both fragments, which is much lower than the uptakes observed in wt medium- and distal jejunum (58 and 130 \( \mu \)g/cm, respectively). These results show for the first time that tocopherol intestinal absorption mainly occurs in the distal part of mouse intestine. SR-BI primarily contributes to vitamin E low-level absorption in the proximal intestine, whereas NPC1L1 principally contributes to its high-level distal absorption. However, the drastic increase in tocopherol absorption found in the medium and distal jejunal fragments strongly suggests a regulated process, almost certainly involving another specific unknown receptor.

Finally, based on the experiments in Caco-2 cells and as expected(11), neither overexpression of SR-BI nor ezetimibe treatment decreased the absorption of NBD-cholesterol, showing that neither SR-BI nor NPC1L1 are implicated in NBD-cholesterol uptake. NBD-cholesterol absorption was even stimulated when the ezetimibe-dependent pathway was inhibited. Our results further underline that the chemical modification from cholesterol to NBD-cholesterol has a real impact in apical receptor recognition and uptake by the enterocyte. These in vitro results, summarised in Fig. 4, refine the data obtained in Caco-2 cells. NBD-cholesterol does not appear to be a suitable cholesterol tracer of the uptake process by SR-BI or NPC1L1. Moreover, although both NPC1L1 and SR-BI are involved in the uptake of cholesterol and vitamin E in vitro, NPC1L1 is definitely critical for the absorption of cholesterol in vivo and contributes, although to a lesser extent, to vitamin E uptake. The iSR-BI transgenic model strengthens the argument for a collateral role of SR-BI. The difference observed between Caco-2 cells and the mouse model could be partly explained by the fact that Caco-2 cells present a different protein pattern to in vitro enterocytes. In particular, Caco-2 cells highly express both SR-BI(77) and NPC1L1(200), whereas other lipid transporters may not be expressed at all(80). The route of lipid delivery, i.e. artificial \( \gamma \)-physiological micelles, could also be important, along with the transporter locations along the intestine and inside the enterocytes.

Thus, despite the fact that cholesterol and vitamin E share common uptake pathways through SR-BI and NPC1L1 in vitro, their uptake profiles along the intestine are highly different, suggesting that other mechanisms co-occur during the digestion–absorption process. Further investigations are needed to understand the delayed tocopherol absorption to the distal part of the intestine compared to cholesterol. Finally, the identification of additional intestinal transporters of fat nutrients remains an essential issue.

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