Epidemiology of group B streptococcus in Korean pregnant women

B. K. LEE¹, Y. R. SONG¹, M. Y. KIM², J. H. YANG², J. H. SHIN², Y. S. SEO², K. Y. OH³, H. R. YOON⁴, S. Y. PAI⁵, B. FOXMAN⁶ AND M. KI*²

¹ Department of Obstetrics and Gynecology, Eulji University Hospital, Daejeon, Korea
² Department of Obstetrics and Gynecology, Cheil General Hospital & Women’s Healthcare Center, Kwandong University College of Medicine, Seoul, Korea
³ Department of Obstetrics and Gynecology, Eulji Hospital, Seoul, Korea
⁴ Seoul Clinical Laboratories & Seoul Medical Science Institute, Seoul, Korea
⁵ Department of Preventive Medicine, Eulji University School of Medicine, Daejeon, Korea
⁶ Department of Epidemiology, School of Public Health, University of Michigan, Ann Arbor, MI, USA

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SUMMARY
Between January 2006 and May 2008, 2624 pregnant S. Korean women between 35–37 weeks gestation were screened for group B streptococcus (GBS). Resistance to antimicrobials was tested by disk diffusion and serotype determined using co-agglutination assays and microarray methods. Overall, 8% of pregnant women were colonized. Serotype III was the predominant serotype (43.8%), followed by serotypes V (20.3%), Ia (12.1%), and Ib (9.5%). GBS was frequently resistant to clindamycin (54.0%) and erythromycin (25.6%); 3.7% were resistant to cefazolin. More than three-quarters of serotype V were resistant to clindamycin or erythromycin or both, and 71% of serotype III were resistant to clindamycin but only 12% were resistant to erythromycin. GBS prevalence exceeded earlier reports by one-third. This is the first report of cefazolin resistance in Korea. These results underscore the need to establish screening measures and chemoprophylaxis guidelines regarding GBS infections in Korea.

Key words: Antimicrobial resistance, cefazolin, clindamycin, colonization rate, erythromycin, prevalence, serotype.

INTRODUCTION
Group B streptococcus (GBS, Streptococcus agalactiae) was recognized as a major cause of neonatal sepsis and meningitis in the 1970s [1], but the first Korean case of GBS neonatal meningitis was not described until 1984. Since then, the number of reported cases of neonatal GBS disease has increased steadily [2].

Neonatal sepsis usually develops within 3 days after birth, and the most frequent causative organism is GBS [3]. Maternal colonization is the primary risk factor; but only 1–2% of babies that become infected develop GBS disease. GBS serotype, neonatal birth weight, and the immune status of the neonate or of the mother during pregnancy modify disease risk [4]. Asymptomatic colonization with GBS is common worldwide, with estimates from vaginal and rectal sampling ranging from 15–30% depending on the
population [5]. Screening is not standard for care in all Korean hospitals, and the few published reports suggest GBS colonization is considerably lower than elsewhere, ranging from 0.3 to 5.9% [6–8]. However, some of these reports did not use GBS-selective media, and urine specimens were not included.

Penicillin is the intrapartum prophylactic antibiotic of choice for prevention of GBS-induced neonatal sepsis. In pregnant women with penicillin hypersensitivity, erythromycin or clindamycin are recommended; however, since the 1990s, resistance to these two antibiotics has increased significantly [6, 9, 10] exceeding 50% in Korea [7].

The incidence rate of clinically diagnosed neonatal sepsis in Korea of 30.4/1000 live births [11] is significantly higher than the 2/1000 reported for the USA [12]. However, the reported prevalence of GBS in pregnant women in Korea is low. To help understand this discrepancy, we screened 2624 Korean pregnant women at 35–37 weeks gestation for GBS.

METHODS

Subjects
All consenting pregnant women of 35–37 weeks of gestation seen in Daejeon, at the Eulji University Hospital or the Mote Obstetrics and Gynecology (OBGY) Clinic, or in Seoul, at the Eulji General Hospital or Cheil Women’s Hospital between January 2006 and May 2008 who had routine prenatal testing were included. GBS was cultured and identified in the Departments of Laboratory Medicine of the Eulji hospitals in Seoul and Daejeon and, for samples obtained at Cheil Hospital, in the Seoul Clinical Laboratory. The laboratory also carried out antimicrobial resistance testing. The Department of Preventive Medicine of Eulji University performed GBS laboratory work for the Mote OBGY Clinic in Daejeon. The Institutional Review Boards at the Eulji (04-08 and 06-25) and Cheil (SCH-IRB-2005-24) hospitals approved the study protocol. Written informed consent permitting use of the sample materials and medical records for research purposes was obtained from every study participant.

GBS isolates

GBS collection
Physicians collected vaginal mucus or discharge with a swab from the vaginal introitus without inserting a speculum, and placed the swab into Stuart’s transport medium. A swab was inserted through the anal sphincter, rotated two or three times, and placed into a separate container of transport medium. Urine samples were self-collected specimens of the first 20 ml of urine. All participating laboratories used the same protocols for GBS incubation and identification. Media and reagents were purchased by the combined research team and distributed to each participating laboratory.

GBS culture
Todd–Hewitt broth supplemented either with gentamicin (8 μg/ml) and nalidixic acid (15 μg/ml), or with colistin (10 μg/ml) and nalidixic acid (15 μg/ml) was used to repress the growth of microorganisms other than GBS. Urine samples were centrifuged, and 1 ml of the sedimented sample was placed into the selective medium. Rectal and vaginal swabs were removed from the transport medium and used to inoculate the selective broth medium. Cultures were shaken three or four times to ensure adequate mixing of the analyte. The lids of the culture tubes were loosely closed and the cultures incubated, along with a negative control, for 18–24 h at 35–37 °C in ambient air containing 5% CO₂. If the medium in the tubes was still clear after 18–24 h, the cultures were reincubated and inspected again at 48 h. Specimens with evident bacterial growth were subcultured on plates containing sheep blood agar (tryptic soy agar with 5% defibrinated sheep blood; TSAII; KOMED Co., Korea).

GBS identification

GBS was identified using the latex agglutination assay (Streptex; Murex Biotech Ltd, UK) and the catalase test.

Antimicrobial resistance

Positive GBS samples were tested for antibiotic resistance by culturing samples of the bacteria on disks containing penicillin G, ampicillin, erythromycin, clindamycin, cefazolin (Sigma Chemical Co., USA), or vancomycin (Daewoong Lilly, Korea) in Muller–Hinton agar (Muller-BAP: KOMED Co., Korea). The size of the inhibitory zone was observed after 18–36 h incubation. Guidelines of the Clinical and Laboratory Standards Institute (CLSI, formerly NCCLS) were used to interpret disk diffusion test. Since the CLSI has not established a resistance standard for cefazolin, the following criteria were used: S ≥ 28 mm; I = 26–27 mm; R ≤ 25 mm, as previously defined [13].
A 1.8-ml storage tube was filled with 0.75 ml Todd–Hewitt broth, inoculated with two or three pure-cultured GBS colonies, and incubated for 18–24 h at 35–37 °C in ambient air containing 5% CO₂. Then, 0.75 ml of glycerol was added to the samples, which, after adequate mixing, were frozen at −70 °C for storage.

GBS serotyping

GBS serotype was determined using hyperimmune rabbit antiserum (Essum, Sweden) against serotypes Ia, Ib, and II–VIII of the capsular polysaccharide of GBS. Capsular serotypes of strains that were non-typable by antigen-antibody precipitation were determined by microarray methods [14, 15].

Statistical analysis

All statistical analyses were done using SPSS 14.0 statistical software (SPSS Inc., USA). The GBS prevalence rate by area and specimen, the serotype, and the antimicrobial resistance of the bacteria were tested for statistical significance using the χ² test.

RESULTS

Colonization rate

A total of 7710 specimens were collected from the urine, vaginal introitus, and anorectal area of 2624 pregnant women at four hospitals in Seoul and Daejeon. From these, 352 GBS isolates were obtained from 211 women (8.0% of the total, Table 1). Prevalence of colonization varied significantly by hospital (P < 0.001) with the highest prevalence (10.5%) observed at Cheil Hospital in Seoul (Table 1). Colonization occurred most frequently in urine (6.2%), significantly (P < 0.001) higher than in the vaginal introitus (4.1%) and anorectal specimens (3.4%, Table 2). All three specimen types could not be collected from all participants. Of the 2526/2624 (96%) pregnant women who contributed all three
specimens, more than one positive culture was obtained from 204 (8.1%); 36/204 (17.6%) were positive in all three specimens.

Serotype distribution
The most common serotypes were III (43.8%), V (20.3%), Ia (12.1%), Ib (9.5%), with 1.6% being non-typable. None of the isolates belonged to serotype IV. The serotype distribution differed by anatomical site \( (P=0.03, \text{Fig. 1}) \), and study site: serotype V was more common in Seoul than Daejeon (22.9% vs. 13.5%) and serotype III was more frequent in Daejeon (56.9 vs. 40.7%).

Antibiotic resistance
None of the GBS isolates were resistant to penicillin G, ampicillin or vancomycin, and there was no significant difference in resistance between Seoul and Daejeon. Cefazolin resistance was observed in 13 (3.7%) of the isolates. Clindamycin resistance was detected in 54.0% of the isolates and 25.6% of the isolates showed erythromycin resistance. All isolates resistant to erythromycin were also resistant to clindamycin, except in two cases (Table 3).

Resistance was associated with serotype: more than three quarters of serotype V isolates were resistant to clindamycin or erythromycin or both. All isolates susceptible to erythromycin and resistant to clindamycin were of serotype III (Table 4).

DISCUSSION
GBS colonization is reportedly low in Asia, and especially Korea (0.3–5.9%) \([6–8]\). Therefore, even though the incidence of neonatal sepsis in Korea is much higher than in other countries [11] there are no guidelines for GBS prevention. Using media selective for GBS, we found a colonization rate of 8%
(4.6–10.5% depending on the hospital), predominance of the more virulent serotypes (serotypes III and V), and very high levels of antimicrobial resistance (54.0% for clindamycin, 25.6% for erythromycin, 3.7% for cefazolin). The higher prevalence of colonization observed in our study compared to previous reports probably reflects the large sample size (2624 pregnant women in four hospitals), use of selective media, and inclusion of urine samples with vaginal and anorectal swabs for GBS culture. Although the colonization prevalence is still much lower than in USA and Europe, the high rates of resistance suggest that GBS is an emerging problem in Korea, and that screening measures and chemoprophylaxis guidelines regarding GBS infections, especially in pregnant women should be established.

The GBS prevalence rate of 8.0% detected in our study is similar to that in neighbouring countries: 8.2% in Japan, 11.2% in Taiwan and 12% in India and Pakistan [16–18]. However it is lower than the 26% found in the USA and 10–36% in Europe [19]. In a study from The Netherlands prevalence differed by country of origin: 13% in Asia, 21% in Europe, and 29% in Africa [20].

The distribution of GBS serotypes found in our study is similar to that reported for USA and Canada, with Ia, III and V predominating [21, 22], and to Taiwan, although in Japan serotypes VI and VIII are most common [23]. This is in contrast to the serotype distribution reported for Korean isolates collected from general hospital laboratories in the 1990s, when serotypes Ib and III were the most prevalent, and serotype V accounted for only 11% [9, 24].

We found a higher rate of resistance to clindamycin and a lower rate of resistance to erythromycin than reported for isolates collected from general hospital laboratories between 1990–2002 in Korea (54.0% vs. 35.0% for clindamycin, and 25.6% vs. 30.0% for erythromycin, respectively) [9]. The resistance rate reported in the current study is still much higher than found in pregnant women in the USA and Canada [10, 25, 26]. We found no previous reports of resistance to first-generation cephalosporins in Korea [24, 27]. The prevalence found here, of 3.7% to cefazolin is similar to that found in the USA (2–4%) [28, 29] and Japan (5.1%) [30]. Similar to previous reports from the USA, GBS serotypes III and V were more likely to be resistant to clindamycin and erythromycin [31]. However, a previous report from Korea did not observe an association between serotype III and high rates of erythromycin and clindamycin resistance [9].

The frequency of maternal GBS colonization and the incidence of early-onset GBS infection in newborns varies by country, influencing the choice of GBS prevention strategy. In the USA, a screening based strategy is used to prevent as many GBS neonatal infections as possible [5]. However, in many European countries, a risk-factor-based strategy is used, as it avoids the increased risk to the infant of allergic reactions later in life, the overgrowth of penicillin-resistant pathogens, and maternal anaphylaxis caused

<table>
<thead>
<tr>
<th>Serotype (n)</th>
<th>Cefazolin</th>
<th>Clindamycin</th>
<th>Erythromycin</th>
<th>Clindamycin and erythromycin</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ia (38)</td>
<td>1 (2.6)</td>
<td>3 (7.9)</td>
<td>1 (2.6)</td>
<td>1 (2.6)</td>
</tr>
<tr>
<td>Ib (30)</td>
<td>0</td>
<td>4 (13.3)</td>
<td>3 (10.0)</td>
<td>2 (6.7)</td>
</tr>
<tr>
<td>II (14)</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>III (138)</td>
<td>11 (8.0)</td>
<td>98 (71.0)</td>
<td>17 (12.3)</td>
<td>15 (10.9)</td>
</tr>
<tr>
<td>IV (0)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>V (64)</td>
<td>1 (1.6)</td>
<td>50 (78.1)</td>
<td>50 (78.1)</td>
<td>49 (76.6)</td>
</tr>
<tr>
<td>VI (14)</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>VII (3)</td>
<td>0</td>
<td>2 (66.7)</td>
<td>2 (66.7)</td>
<td>2 (66.7)</td>
</tr>
<tr>
<td>VIII (9)</td>
<td>0</td>
<td>7 (77.8)</td>
<td>7 (77.8)</td>
<td>7 (77.8)</td>
</tr>
<tr>
<td>Non-typable (5)</td>
<td>0</td>
<td>1 (20.0)</td>
<td>1 (20.0)</td>
<td>1 (20.0)</td>
</tr>
<tr>
<td>Missing (35)</td>
<td>0</td>
<td>24 (68.6)</td>
<td>12 (34.3)</td>
<td>12 (34.3)</td>
</tr>
<tr>
<td>Total (350†)</td>
<td>13 (3.7)</td>
<td>189 (54.0)</td>
<td>93 (26.6)</td>
<td>89 (25.4)</td>
</tr>
</tbody>
</table>

* All isolates were susceptible to ampicillin, penicillin G and vancomycin.
† Two isolates were missed.
by penicillin allergy [20, 32]. In Korea, no guidelines exist for GBS prevention and no data for the risk of GBS colonization; given the high rates of sepsis, and increasing GBS colonization, it is time for Korea to implement a screening-based strategy to prevent GBS neonatal infections.

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DECLARATION OF INTEREST

None.

REFERENCES


