Analysis of penicillinase-producing Neisseria gonorrhoeae isolates in Madrid (Spain) from 1983-85

By A. FENOLL, S. BERRÓN AND J. A. VÁZQUEZ

Servicio de Bacteriología, Centro Nacional de Microbiología, Virología e Inmunología Sanitarias, Majadahonda (Madrid), Spain

(Accepted 2 March 1987)

SUMMARY

Between April 1983 and December 1985, 576 strains of Neisseria gonorrhoeae were isolated in our laboratory from patients attending Sexually Transmitted Diseases (STD) clinics. Of these, 61 (10.6%) were penicillinase-producing. Studies on these strains by plasmid analysis, auxotyping and serogrouping showed that the predominant type strains harboured the Asian resistance plasmid, were prototrophic, and were of serogroup WII/WIII. About half of the strains, both of the African and Asian type, harboured the transfer plasmid. Strains of serogroup WII/WIII were less sensitive to tetracycline and cefoxitin than serogroup WI strains.

INTRODUCTION

A penicillinase-producing *Neisseria gonorrhoeae* (PPNG) strain was first isolated in Spain in 1979 (Alomar, Gil Sanchez & Arteaga, 1979).

In Spain gonorrhoea was not a reportable disease until 1982, and the available data on the isolation of PPNG was both scarce and incomplete. In 1982 12·5 % of strains in Barcelona were PPNG and during 1983 such strains constituted 9·7 % of gonococcal isolates in Centro Nacional de Microbiología (C.N.M.V.I.S.) from the Madrid region (Vázquez et al. 1985).

In this study the PPNG strains isolated during the period from April 1983 to December 1985 from the Madrid region have been characterized by auxotyping, serogrouping and plasmid analysis.

Minimum inhibitory concentrations (MIC) to five antibiotics (benzylpenicillin, spectinomycin, tetracycline, cefoxitin and ceftazidime) were also determined in order to demonstrate possible relationships between antibiotic susceptibilities and other features of the organisms.

MATERIAL AND METHODS

Population studied

Between April 1983 and December 1985, specimens from 1648 males and 1294 females attending Sexually Transmitted Diseases (STD) clinics were examined for the presence of N. gonorrhoeae.

Isolation and identification

The transport of specimens to the laboratory, the isolation in Thayer-Martin medium and identification of the strains by means of the Phadebact test (Pharmacia Diagnostic, Piscataway, New Jersey) have been described previously (Vázquez, Berrón & Fenoll, 1985).

All isolates were tested for penicillinase production using the acidometric method ('beta-lactamase detection papers', Oxoid Ltd, Basingstoke, England).

Auxotyping

Gonococcal auxotyping was performed by the method of Catlin (Catlin, 1978). Strains of *N. gonorrhoeae* of known auxotype were used as controls: NCTC (National Culture Type Collection) 10928, prototrophic (wild type); NCTC 10930, Pro Hyx (proline, hypoxanthine); NCTC 10931, Pro Met Thi (proline, methionine, thiamine); NCTC 10932, Arg Met (arginine methionine); NCTC 10933 Arg Hyx Ura (arginine, hypoxanthine, uracil).

Serogrouping

Gonococcal serogrouping was performed using reagents against WI and WII/WIII antigens commercially available (Phadebact Monoclonal GC Test-Pharmacia Diagnostic, Uppsala, Sweden).

Plasmid determination

Plasmid DNA was extracted and purified according to the method of Birboim & Doly, (1979), with an additional incubation at 55 °C for 30 min to achieve complete lysis of the strains. Electrophoresis was carried out on 0.6 % agar (Meyers et al. 1976). Escherichia coli V517 (Macrina et al. 1978) containing eight plasmids of known molecular weight was used as control.

Antibiotics and susceptibility testing

Antibiotic susceptibility tests were performed by the agar dilution technique (Thornsberry, Gerlach & Sherris, 1977) with strains of *N. gonorrhoeae* with known antibiotic susceptibilities as controls (provided by Dr Knut Lincoln, University of Goteborg, Sweden).

Gonococci were inoculated using a multipoint inoculator (Microtiter AM 80) on 3.5% chocolate agar (GC medium base, Difco) containing 5% haemolysed human blood and 1% Kellog supplement (Wite & Kellog, 1985) together with twofold dilutions of the drug to be tested. After incubation for 24 h at 37 °C in 5% CO² the MIC was read.

The range of concentrations of the antibiotics tested was 0.007-1280 mg/l benzyl penicillin G, 0.12-8.0 mg/l tetracycline, 0.12-4.0 mg/l cefoxitin, 2.0-64.0 mg/l spectinomycin and 0.007-0.12 mg/l ceftazidime.

Statistical analysis

 χ^2 test with Yate's correction was used (Colton, 1974).

Table 1. Incidence of N. gonorrhoeae isolations during 1983-85 in STD clinics in Madrid region

Year	No. cases	M (males)	F (females)	
		Total cases		
1983	354	178	176	
1984	927	568	359	
1985	1661 902		759	
Total	2942	1648	1294	
	Isolatio	n of N. gonorrh	oeae (%)	
1983	82	55	27	
1001	(23.2)	(30.9)	(15·3)	
1984	224 (24·2)	163 (28·7)	61 (17·0)	
1985	270	194	76	
1000	(16·2)	(21.5)	(10·0)	
Total	576	412	164	
2 2 3	(19.6)	(25.0)	(12.7)	
		PPNG (%)		
1983	10	6	4	
	(12.9)	(10.9)	(14·8)	
1984	23	16	7	
	(10.3)	(9·8)	(11.4)	
1985	28	19	9	
	(10.4)	(9·8)	(11.8)	
Total	61	41	20	
	(10.6)	(10·0)	(12.2)	

RESULTS

Gonococci were isolated from 412 males and 164 females. Of the 576 isolates, 61 (10.6%) were PPNGs (Table 1). During 1983, 1984 and 1985 the prevalence of PPNG strains was about the same.

Of a total of 43 (70.5%) PPNG strains which had no special nutritional requirements (wild type), 6 (14%) belonged to the WII/WIII serogroup. Of the 15 (24.5%) strains requiring proline, 9 (60%) belonged to the WI serogroup and 6 (40%) to the WII/WIII serogroup (Fig. 1). Three strains (5%) belonged to other auxotypes.

The relationships between serogroups and in vitro susceptibility of the PPNG strains to tetracycline, cefoxitin and ceftazidime are presented in Table 2. For tetracycline, 5/16 strains in serogroup WI had an MIC \geq 1 mg/l compared with 31/45 in the WII/WIII serogroup (P < 0.025). For cefoxitin, 3/16 strains in the WI serogroup had an MIC \geq 1 mg/l as did 30/45 in the WII/WIII serogroup (P < 0.005). All strains in both serogroups were sensitive to ceftazidime and to spectinomycin.

All strains harboured the 2.6 MDa plasmid, 8 (13.1%) the 3.2 MDa plasmid (African) and 53 (86.9%) the 4.5 plasmid (Asian). In the 28 strains which harboured the 24.5 MDa plasmid as well, 4 were African type and 24 Asian

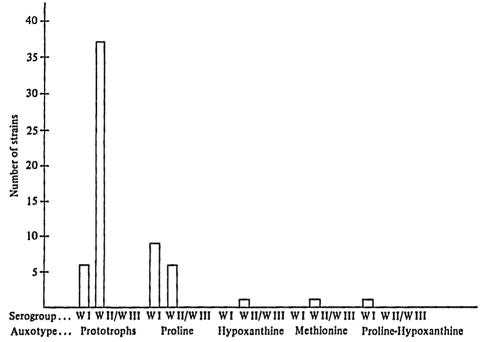


Fig. 1. Incidence of combinations of auxotypes and serogroups in 61 PPNG strains.

Table 2. In vitro susceptibility of PPNG strains to tetracycline, cefoxitin and ceftazidime in relation to serogroups

	Minimum inhibitory concentrations, mg/l				
Serogroup	0.007-0.03	0.06-0.125	0.25-0.50	1.0-4.0	Total
Tetracycline W I		1	10	5	16
WII/WIII	_	_	14	31	45
Total		1	24	36	61
Cefoxitin					
WI		1	12	3	16
WII/WIII			15	30	45
Total	-	1	27	33	61
Ceftazidime					
WI	16		distant		16
WII/WIII	41	4	-		45
Total	57	4	-		61

type (Table 3). A gel loaded with representative DNA preparations is shown in Fig. 2.

All African strains were of the prototrophic auxotype whereas the 53 Asian PPNG were distributed among five different auxotypes (Table 3).

Table 4 shows the MIC for the five antibiotics tested on strains with the African plasmid and with the Asian plasmid. The only difference nearing significance between the two groups was with tetracycline (P < 0.05).

Plasmids (MDa)		Auxotype				
	No. of strains	Proto- trophic	Proline	Hypox- anthine	Meth- ionine	Proline- Hypox- anthine
		African type				
2.6-3.2	4	4/0*		··-		
2.6-3.2-24.5	4	0/4				
Total	8	4/4				
		Asian type				
2.6-4.5	29	1/16	5/5		0/1	1/0
2.6-4.5-24.5	24	1/17	3/2	0/1	<u>.</u>	<u>-</u>
Total	53	2/33	8/7	0/1	0/1	1/0
		•				

Table 3. Plasmid content, auxotype and serogroup of 61 PPNG strains

DISCUSSION

The high incidence of PPNG strains found by us during 1983 in Madrid (Vazquez et al. 1985) has changed little over the 3 years of this study and is similar in males and females. The incidence of PPNG strains resembles that described in the Netherlands (Ansink-Schipper et al. 1984), but is higher than reported from most European countries (Anon, 1983). It is not possible to assess whether the situation in the Madrid region is similar to that in other regions of Spain since data is both sparse and incomplete. The Spanish Weekly Microbiological Bulletin reported an incidence of PPNG strains of 7.3% during 1985 (Anon, 1986) but the number of isolates included was small.

Some information about the epidemiology of PPNG was obtained through the application of a number of typing techniques. The major typing patterns comprised strains of serogroup W11/W111 which were not nutritionally exacting (protoprophs) followed by strains of W1 serogroup, proline dependent. Thereafter strains of W1 serogroup, prototrophs and of W11/W111 serogroup, proline dependent were identified in similar numbers. Other auxotypes were occasionally found (Fig. 1). This dominant incidence of prototrophic strains followed by proline dependent ones is much the same as that reported from other countries (Ansink-Schipper et al. 1984; Yvert et al. 1985). In our study a greater proportion of proline dependent strains were of serogroup W1 than were prototrophic strains (P < 0.001).

Strains of serogroup WII/WIII were less sensitive to tetracycline and cefoxitin than strains of serogroup WI, a correlation that has been described before (Danielson, Bygdeman & Kallings, 1983; Bygdeman et al. 1982). Thus high incidence of serogroup WII/WIII in Spain could well be of clinical importance. Though spectinomycin-resistant PPNG strains have been reported in other countries (Anon, 1983; Pond et al. 1986), none were found in this study.

About 87% of the PPNG isolates possessed the Asian plasmid and half of those harboured the conjugative plasmid (24.5 MDa) as well. The African type was present in only 13% of PPNG strains. Comparable results have been reported from

^{*} No. in serogroup WI/no. in serogroup WII/WIII.



460

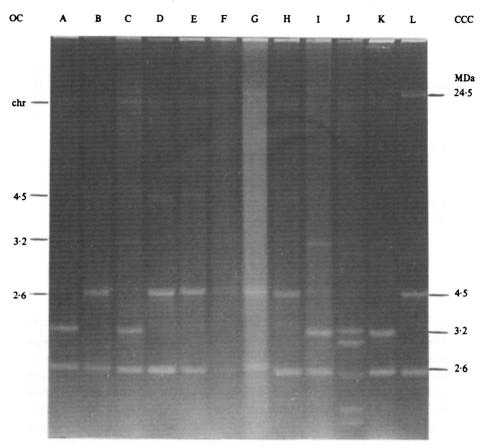


Fig. 2. Agarose gel electrophoresis of plasmid DNA from different PPNG strains containing: 2·6 and 3·2 MDa plasmid (A, K), 2·6, 3·2 and 24·5 MDa plasmid (C, I), 2·6 and 4·5 MDa plasmid (B, D, E, F, H) and 2·6, 4·5 and 24·5 MDa plasmid (G, L). Lane J shows plasmids from *E. coli* V 517.

Table 4. In vitro susceptibility to antibiotic of PPNG strains harbouring the 'African' plasmid and PPNG strains carrying the 'Asian' plasmid

Plasmid type	$MIC_{90} (mg/l)$					
	Penicillin	Spectinomicin	Tetracycline	Cefoxitin	Ceftazidime	
'African'	24.2	14.9	0.6	0.6	0.024	
'Asian'	37.7	14.9	1.9	1.3	0.029	

England and Switzerland (Goh et al. 1985; Arini et al. 1985) but in the Netherlands a higher incidence is recorded (Ansik-Schipper et al. 1984). Half of the isolates with the African plasmid harboured the transfer plasmid as well. The emergence of this combination was associated, in the Netherlands, with an increase in the incidence of PPNG infections (Ansik-Schipper et al. 1984). All African type strains in this study carrying the transfer plasmid were isolated during 1985 which may have a serious epidemiological impact.

Further features of note were a correlation demonstrable between the non-exacting auxotype and the African plasmid already noted elsewhere (Arini et al.

1985; Ansik-Schipper et al. 1984), and the difference between African type and Asian type strains for *in vitro* susceptibility to tetracycline which has also been reported before (Goh et al. 1985; Perine et al. 1977).

Unfortunately, epidemiological data to assess whether the PPNG isolates were from locally acquired or imported infections is not available. However, from 20 PPNG strains isolated in women, 8 were found in prostitutes and 3 in promiscuous women not admitting to working in prostitution. Prostitutes have been described as an important reservoir of PPNG infections in some countries (Ansik-Schipper et al. 1984; Arya et al. 1984) and may well be so in Spain. Perhaps surprisingly no beta-lactamase producing strains were isolated from homosexual men.

The authors thank Isabel Hernandez for her collaboration and technical assistance. We also thank Dra Cecilia Martin for her suggestions during the preparation of the manuscript.

REFERENCES

- Anonymous (1983). World Health Organisation. Surveillance of beta-lactamase producing Neisseria gonorrhoeae (PPNG). World Health Organization Weekly Epidemiological Record 58, 5-12.
- Anonymous (1983). Centers for Disease Control. Spectinomicin-resistant penicillinase-producing Neisseria gonorrhoeae. Morbidity and Mortality Weekly Report 32, 51-52.
- Anonymous (1986). Ministerio de Sanidad y Consumo (Spain). Infecciones bacterianas de transmisión sexual notificadas al BMS en 1985. Boletin Microbiológico Semanal 15, 9-10.
- ALOMAR, P., GIL SÁNCHEZ, J. & ARTEAGA, B. (1979). Aislamiento de una cepa de Neisseria gonorrhoeae productora de beta-lactamasa. Resumen de comunicaciones, VII Congreso Nacional de Microbiología (Cádiz), p. 128.
- Ansink-Schipper, M. C., Van Klingeren, B., Huikeshoven, M. H., Woudstra, R. K., Dessens-kroon, M. & Van Wijngaarden, L. J. (1984). Epidemiology of PPNG infections in the Netherlands: analysis by auxonographic typing and plasmid identification. *British Journal of Venereal Diseases* 60, 141-146.
- Ansink-Schipper, M. C., Huikeshoven, M. H., Woudstra, R. K., Klingeren, B., Koning, G. A. J., Tio, D., Jansen Schoonhoven, F. & Coutinho, R. A. (1984). Epidemiology of PPNG infections in Amsterdam: Analysis by auxonographic typing and plasmid characterisation. *British Journal of Venereal Diseases* 60, 23-28.
- Arini, A., Eichmann, F., Peduzzi, R. & Piffaretti, J. C. (1985). Plasmids in Neisseria gonorrhoeae isolated in Switzerland: relation with the auxotypes. European Journal of Sexually Transmitted Diseases 2, 75-79.
- ARYA, O. P., REES, E., TURNER, G. C., PERCIVAL, A., BARTZOKAS, C. A., ANNELS, E. H., CAREY, P. B., Ghosh, A. K., Jephcott, A. E. & Johnstons, N. A. (1984). Epidemiology of penicillinase-producing Neisseria gonorrhoeae in Liverpool from 1977 to 1982. Journal of Infection 8, 70-83.
- BIRBOIM, H. C. & DOLY, A. (1979). A rapid alkaline extraction procedure for screening recombinant plasmid DNA. Nucleic Acid Research 7, 1513-1523.
- Bygdeman, S., Bäckman, M., Danielsson, D. & Norgren, M. (1982). Genetic linkage between serogroup specificity and antibiotic resistance in *Neisseria gonorrhoeae*. Acta Pathologica Microbiologica Scandinavica Sect. B 90, 243-250.
- CATLIN, B. W. (1978). Characteristics and auxotyping of Neisseria gonorrhoea. In Methods in Microbiology 10 (eds. T. Bergan, and J. R. Norris) pp. 345-380. London: Academic Press.
- COLTON, T. (1965). Statistics in Medicine, p. 174. Boston: Little Brown Co.
- Danielsson, D., Bygdeman, S. & Kallings, I. (1983). Epidemiology of gonorrhoea: serogroup, antibiotic susceptibility and auxotype patterns of consecutive gonococcal isolates from ten different areas of Sweden. Scandinavian Journal of Infectious Diseases 15, 33-42.
- Goh, B. T., Rodin, P., Johnston, N. A. & Wong, H. Y. (1985). Penicillinase-producing

- Neisseria gonorrhoeae: epidemiology, antimicrobial susceptibility and plasmid types. Journal of Infection 11, 63-69.
- MACRINA, F. L., KOPECHO, D. J., JONES, K. R., AYERS, D. J. & McCOWEN, J. M. (1978). A multiple plasmid-containing *Escherichia coli* strain: convenient source of size reference plasmid molecules. *Plasmid* 1, 417–420.
- MEYERS, J. A., SANCHEZ, D., ELWELL, L. P. & FALKOW, S. (1976). Simple agarose gel electrophoretic method for identification and characterization of plasmid deoxyribonucleic acid. *Journal of Bacteriology* 127, 1529-1537.
- Perine, P. L., Thornsberry, C., Schalla, W. & Siegel, M. S. (1977). Evidence for two distinct types of penicillinase-producing *Neisseria gonorrhoeae*. *Lancet* ii, 993-995.
- Pond, E., Batchelor, R. A., Howell, B., Kerry, G., Lake, J., Rice, R. J., Biddle, J. W., & Conwill, D. E. (1986). An unusual case of penicillinase-producing *Neisseria gonorrhoeae* resistant to spectinomic in California. *Sexually Transmitted Diseases* 13, 47–49.
- THORNSBERRY, C., GERLACH, E. H. & SHERRIS, J. C. (1977). New developments in antimicrobial agent susceptibility testing. In *Cumitech 6* (ed. J. D. Sherris). Washington DC: American Society for Microbiology.
- VAZQUEZ, J. A., FENOLL, A., BERRÓN, S., TÉLLEZ, A. & BILBAO, R. (1985). Penicillinase-producing strains of Neisseria gonorrhoeae in Madrid. Genitourinary Medicine 61, 139.
- VÁZQUEZ, J. A., BERRÓN, S. & FENOLL, A. (1985). Rapid diagnosis of Neisseria gonorrhoeae by enzyme immunoassay (Gonozyme). European Journal of Sexually Transmitted Diseases 2, 217-220.
- WITE, L. A. & KELLOG, D. S. (1965). Neisseria gonorrhoeae identification in direct smears by a fluorescent antibody-counterstain method. Applied Microbiology 13, 171.
- YVERT, F., FROST, E., GUIBOURDECHE, M., RIOU, J. Y. & IVANOFF, B. (1985). Auxotypes and serogroups of penicillinase-producing and non-producing strains of *Neisseria gonorrhoeae* isolated in Franceville, Gabon. *Genitourinary Medicine* 61, 100-102.