

REVIEW ARTICLE

Association between human papillomavirus (HPV) and oesophageal squamous cell carcinoma: a meta-analysis

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SUMMARY

The oncogenic potential of human papillomaviruses (HPV) is well known in the context of cervical carcinoma; however, their role in the development of oesophageal squamous cell carcinoma (OSCC) is less clear. We aimed to determine the extent of the association between HPV infection and OSCC. A comprehensive literature search found 132 studies addressing HPV and OSCC in human cases, and a meta-analysis was performed using a random-effects model. There was evidence of an increased risk of OSCC in patients with HPV infection [odds ratio (OR) 2·69, 95% confidence interval (CI) 2·05–3·54]. The prevalence of HPV in OSCC was found to be 24·8%. There was an increased risk associated with HPV-16 infection (OR 2·35, 95% CI 1·73–3·19). Subgroup analyses showed geographical variance, with Asia (OR 2·94, 95% CI 2·16–4·00), and particularly China (OR 2·85, 95% CI 2·05–3·96) being high-risk areas. Our results confirm an increase in HPV infection in OSCC cases.

Key word: Human papilloma virus (HPV).

INTRODUCTION

Globally, cancer of the oesophagus is the eighth leading cause of cancer mortality [1]. The US government spends an estimated \$1.1 billion [2] on treating over 30 000 [3] Americans suffering from the disease each year. Barrett's oesophagus, the premalignant metaplasia of oesophageal tissue related to gastro-oesophageal reflux disease, is a major risk factor for oesophageal adenocarcinoma [4]; however, the causes of squamous cell carcinoma are less clear.

Oesophageal squamous cell carcinoma (OSCC) has traditionally been associated with risk factors such as tobacco and alcohol [5], although given the vast

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geographical differences in incidence, the aetiology of the disease remains poorly understood. Countries such as the People's Republic of China (China), Singapore, Iran, Chile, Brazil, South Africa and France have all been identified as high-risk regions for the development of OSCC where incidence rates are markedly higher than in other areas [6]. Given this variance, many possible risk factors have been studied in an attempt to determine the cause of OSCC – including nutritional deficiencies (e.g. vitamins A, B, C) [6], physical factors (e.g. poor oral hygiene, hot drinks) [7], chemicals (e.g. nitrosamines, tobacco, alcohol, opium) [7], and viruses [e.g. Epstein–Barr virus, herpes simplex virus, human papillomavirus (HPV)] [6].

HPV was first described in 1949 in skin warts; however, it was not until the mid-1970s that it was linked to cervical cancer [8]. The link between HPV and cervical cancer has since been well established,

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given global detection of HPV DNA in over 90% of cervical cancers and the lack of geographical variation [9]. Meanwhile, research was also being conducted into the malignant conversion of other anogenital as well as extragenital papillomatous lesions to squamous cell carcinomas [8]. In 1985, specific HPV types were first found in oropharyngeal carcinomas and it is now thought that over 25–30% of these cancers may be caused by high-risk HPV types [8]. In 1982, Syrjänen *et al.* published the first study into the histological presence of known HPV patterns in 60 Finnish patients with established OSCC and found positive results in 40% [10]. Since then, research has continued into the possible causative role of HPV infection in OSCC pathogenesis.

The aim of this meta-analysis is to combine, examine and quantify the results from studies addressing the association between HPV infection and the development of OSCC on a global scale.

METHODS

Study protocol

We followed the PRISMA guidelines where possible in performing our systematic review [11]. A systematic search was undertaken through Medline (from 1950), PubMed (from 1946), EMBASE (from 1949) and Current Contents Connect (from 1998) and Google Scholar to 3 December 2012, to identify relevant articles where possible. The search used the terms 'oesophageal cancer' OR 'esophageal cancer' AND 'human papillomavirus', which were searched as text words and as exploded medical subject headings. The reference lists of relevant articles were also searched for appropriate studies. No language restrictions were used in either the search or study selection. There were four studies included which were not published in English [12–15]. A search for unpublished literature was not performed.

Study selection

We included cross-sectional, case-control and cohort studies that met the following inclusion criteria: (1) all cases and controls were human and adult; (2) the oesophageal cancer was squamous cell carcinoma; (3) all patients had no other health conditions. We excluded studies that did not meet the inclusion criteria.

Data extraction

The data extraction was performed via a standardized data extraction form, collecting information on publication years, study designs, numbers of cases, numbers of controls, total sample sizes, countries, continents, development statuses, control sources, mean ages, percentage male of total samples, the risk estimates or data used to calculate the risk estimates, confidence intervals (CIs) or data used to calculate CIs, the types of oesophageal carcinoma investigated, the methods of detection of HPV in the samples and the specific types of HPV detected. Where no data was available on the specific types of HPV detected, studies were included in the Nonspecific analysis. Where no control group was used, studies were included in the Prevalence analysis. Quality of the studies was not assessed and authors were not contacted for missing data. Adjusted odds ratios (ORs) were extracted in preference to nonadjusted ORs; however, where ORs were not provided, unadjusted ORs and CIs were calculated. Where more than one adjusted OR was reported, we chose the OR with the highest number of adjusted variables. A sample which was found to be HPV-16 and HPV-18 positive was included in both subgroup analyses but in the overall analysis only once. However, a number of studies did not distinguish in their results between HPV-16 and HPV-18 positive samples and instead gave combined results as samples being positive for HPV-16/18. As such these studies were included in pooled HPV-16/18 analysis.

Four studies did not distinguish between histological types of oesophageal carcinoma and, in order to include them in the meta-analysis, they were assumed to be squamous cell carcinoma (Table 1). Where multiple control groups were studied, we selected the group of separate people over normal mucosal biopsies taken from areas adjacent to the tumour site.

Statistical analysis

Pooled ORs and 95% CIs were calculated for the effect of HPV on the risk of developing OSCC using a random-effects model [16]. We tested heterogeneity with Cochran's Q statistic, with P < 0.10 indicating heterogeneity, and quantified the degree of heterogeneity using the I^2 statistic, which represents the percentage of the total variability across studies which is due to heterogeneity. I^2 values of 25%, 50% and 75% corresponded to low, moderate and high degrees

Table 1. Summary of studies included in the meta-analysis

									Non-	specific HI	v		HPV-1	6			HPV-1	18			HPV-	16/18		
Author	Country	Mean age	Case % male	Data- collection year	Cases	Controls	Detection method	Control source	+ve		OR (95% CI)	Prev		+ve	OR (95% CI)	Prev (%)	+ve	HPV +ve Controls	OR (95% CI)	Prev (%)	HPV +ve cases	+ve	OR (95% CI)	Prev (%)
Abdirad,	Iran†	58-84	57	1991-2005	93		PCR					8-6				3.22				1.07				4.3
2011 [79] Acevedo- Nuno,	Mexico†	63.06	88		17		PCR					88-24												
2004 [80]* Akutsu,	Japan			1989–1993	29		PCR + SBH					0												
1995 [81] Antonsson, 2010 [22]	Australia	65-2	56	2002-2005	222	55	PCR	Other	8	0	2.02 (0.25–16.49)‡	3.6	6	0	1.5 (0.18–12.72)‡	2.7					6	0	1.5 (0.18–12.72)‡	2.7
Astori,	Italy	67	71		14	16	PCR + SBH	ANM	6	4	2·25 (0·48–10·6)	42-86	3	3	1.18 (0.2–7.08)	21.43					3	3	1.18 (0.2-7.08)	21-43
2001 [23] Awerkiew,	Germany		69		23		PCR					0												
2003 [82] Ayshamgul,	China†	50-5		2007-2009	50		PCR					38				38								38
2011 [83] Bahnassy,	Egypt	61.3	68	1996–1998	47	50	PCR + IHC	ANM	24	12	3·3 (1·39–7·85)	51-06				25.53				12.77				38-3
2005 [24] Baines,	USA				16		PCR					0												
2005 [84] Benamouzig, 1992 [26]	France	62	92	1990	12	17	ISH + DBA +histological	Other	1	0	1·45 (0·08–25·81)‡	8-33												
Benamouzig,	France	64	75		75	49	diagnosis PCR+ DBA	ANM	0	0		0												
1995 [25] Bjorge,	Norway					123	ELISA	Other			2·2 (0·9–5·3)§	21			2-9 (0-8-10)				2·2 (0·7–6·7)					
1997 [27] Bognar, 2008 [85]	Germany			1999–2000	16		PCR + SBH					31-25				31.25								31-25
Brandsma, 1989 [28]	USA			1984–1987	3	3	SBH	Other	0	0		0												
Cao, 2005 [29]	China†	50-12	58	1999–2003	265	357	PCR	Other	207	203	2·17 (1·89–3·88)	78-11	182	179	2·18 (1·56–3·04)		11	5	3.05 (1.05–8.88)		186	181	2·29 (1·64–3·2)	
2005 [29] Castillo, 2006 [86]	Colombia and Chile†	63-6-72-2	53	1996–2001	73		PCR + SBH					28-77				15-07				47-62				28-77
2000 [80] Chang, 1992 [90]	China†	54-6			51		ISH+PCR +SBH					49-02												
Chang,	China†	53·3(M)	59	1989–1990	363		ISH					22-31				0.28								
1993 [89] Chang,	China†	57·2(F) 51–57	50		36	21	ISH	ANM	4	2	1.82 (0.18-18-69)	8-33												
1997 [31] Chang, 2000 [91]	China†	55·3–57·4	62		101		ISH + PCR					16-83												
Chang, 2000 [87]	China†	55·1(M) 57·3(F)	58	1989–1990	700		ISH					16-57				2								
Chang, 1990 [30]	China†	-, -(-)			6	9	FISH	Other	3	0	7 (0.69–70.75)‡	66-67												
Chang, 1990 [88]	China†	52.3	51		51		ISH					3.92								1.96				
Chen, 1994 [92]	China†	58-2	88		40		PCR + SBH					60				15								
Damin,	Brazil†	58-4	70	1989–1996	165	26	PCR	Other	26	0	4.68 (0.61–36.04)‡	15-76	25	0	4·46 (0·58– 34·46)‡	15-15	2	0	0.31 (0.03-3.51)‡	1.21	26	0	4.68 (0.61–36.04)‡	15.76
2006 [32] de Villiers,	China†	53-9-57-8	49		117		PCR					17-09			5 4.40) ‡									
1999 [94] de Villiers,	Germany	60	81		21		PCR					66-67				47-62				4.76				52-38
2004 [93] Dillner,	Finland			1968–1991	39	78	ELISA	Other			13-1 (1-6-108)§	24-14				24.14								24.14
1995 [33] Ding,	China†	58	71	2005-2008	17		PCR					47-06												

Table 1 (cont.)

									Non-	Non-specific HPV	>		HPV-16			HPV-18			HPV-16/18		
Author	Country	Mean	Case %	Data- Case % collection male year	Cass	ss Contro	Detection Cases Controls method	Control	HPV +ve cases	HPV +ve controls	OR (95% CI)	Prev +	HPV HPV +ve +ve Cases Controls	OR \$ (95% CI)	Prev (%)	HPV HPV +ve +ve Cases Cont	HPV +ve OR Controls (95% CI)	Prev +	HPV HPV +ve +ve cases controls	OR ols (95% CI)	Prev (%)
Dreilich,	Sweden			1990–2000	71		PCR					14.08			14.08						14.08
Emadian,	Iran†			2001-2008	40	9	PCR	ANM	15	S	4·2 (1·35– 13·06)	37.5									
Erol,	Hungary	57	99		4		PCR					25									
2009 [13] Far,	Iran†		49		140		PCR					23-57									
2007 [97] Farhadi,	Iran†	\$.2	28	1996-2001	38	38	PCR	Other	4	Ś	3-85 (1-22-12-14)	36.84 5	0	5.61	13.16	3 5	0.57 (0.12–2.56)	7.89	Ś	1.67 (0.52–5.97)	21.05
Eidalgo,	Portugal	62	92		16	∞	PCR	ANM	6	∞	0.18 (0.02–1.86)	56-25 8	S	0.37 (0.07-1.95)	115	3	0.28 (0.05-1.56)	18.75 9	∞	0.48 (0.09-2.52)	56.25
Furihata,	Japan		79	1981–1992	38		ISH					36.84			13·16			23.68			36.84
1993 [98] Furihata,	Japan		87	1981–1992	71		ISH					33.8			14.08			19.72			33.8
Gao, 2007	China†	40-50		2002	4	475	Hybrid	Other	2	373	0.27 (0.04–1.97)	90									
2006 [36] Goto,	Japan,	62			181		Capture II PCR + ISH					9.39									7.18
2011 [100]	China,																				
Hale,	South Africa†		45	1983-1988	20		Modified					9									
1989 [101] Han,	China†		70	1995	90	121	Feulgen ELISA	Other			4.5 (1.8–11.9)	24									
1996 [37] Haswgawa,	Japan				48		PCR + ISH					41.67			35.42			20			41.67
2002 [102] He,	China†		88	1987–1994	152	152	PCR + SBH	ANM	37	0	48-58	24·34 31	0	38.69	20.39	0 8	8:39	5.26 37	0 4	48.58	24:34
Herrera- Goepfert,	Mexico†	62·7(M) 61·2(F)	78	2000-2008	09		PCR				*(0c.4cc-1c.0)	25		†(ot.107-17.6)	0		*(76.10+0.1)	8:33		*(oc.466-16.0)	18-33
Hille,	South Africa†				24		Histological					33-33									
Hille,	South Africa†	53	100		70		Histological criteria					33-86									
Hippelainen,	Finland	89	49	1979–1991	61		Histological					18.03									
Hussain, 2011 [39]	India†	57.5	57		75	75	PCR	ANM	4	0	16.98	18-67	0 41	16.98	18-67			-	0 14	16.98	
Junquera, 2009 [14]	Spain	62	80	2007–2008	Ξ		PCR					54.54									
Kamangar,	China†	<u>¥</u> ∞	49	1985–1991	66	381	FISH	Other	33	106	1.3 (0.18-2.08)	33-33 1	12 29	1.67 (0.82–3.14)	12.12	8 35	0.87 (0.39–1.94)	8.08 20	0 64	1.25 (0.72–2.19)	20.2
Kamath,	USA	9	50	1988-1998	22		PCR					0									
Z000 [107] Katiyar, 2005 [41]	India†		73		101	26	PCR	Other	19	_	5·79 (0·74-45·48)	18-81	17 1	5.06 (0.64-39.92)	16.83	2 0	0.5 (0.04-5.8);	1.98	19 1	5.79 (0.74-45.46)	18-81
Kawaguchi,	China and			1993–1998	75	75	PCR	ANM	17	17	1 (0.47–2·15)	22-67						1	17 17	1 (0.47–2.15)	
Khurshid,	Japan	66.2	74		27	12	PCR	Other	17	6	5-1 (1-11-23-37)	62-96 3	0	1.37	11:11	0 4	1.91 (0.19–19.2);	14.81 7	0	7-7 (0-8-74-05)‡	25-93
Kiki,	Turkey		43		30	10	PCR	Other	10	0	3·1 (0·35–27·66);	33-33 9	0	3.86	30	0 4	2:74	23-33 1	10 0	4.5 (0.5-40.66);	33-33
Kim,	Koreat				24	21	PCR	ANM	16	2	19 (3-52-102-58)	1 29.99	16 2	19 (3.52–101.58)	29.99		*(+c.cz-cz.o)	_	16 2	19 (3-52-101-58)	29-99
1992 [15] Kiyabu, 1989 [108]	USA			1950–1987	13		PCR + DBA					0									

									Non-sp	Non-specific HPV	^	_	HPV-16			HPV-18			HPV-16/18	18	
The part	Author Country	Mean		Data- % collection year	Cases	s Controls	Detection method	Control			OR (95% CI)			PV e OR antrols (95% CI)	Prev		vv b OR atrols (95% CI)	Prev (%)	HPV H +ve + cases cc	PV ve OR ontrols (95% CI)	Prev (%)
Newforkshiely should be some a single should be some a		62-3	95	1998–1999		40	PCR	ANM		0		0									
Mandanial Annotation of the Mandaman Annotation	Τ		76		63		PCR					0									
Anamatist Anamatist Probability	Ö			2006 2007	267		PCR					0.37									
seeding 45 model 3 model 3 model 3 model 3 model 4 4 model 4				1973–1985		10	FISH	ANM		0	9 (0.81–100-14)‡	50									
Houghous St. St. Houghous St.	<u> </u>				39		hisFISH					23.08									
Hare Keing Sign Sign Sign Sign Sign Sign Sign S		2	74	1994–1997		302	ELISA	Other		62	0.77 (0.44–1.34)				11.57			4.96			16.53
Maintain			84	1992–1994			PCR + SBH					8.57									
Such Substitution (Control of Control of Con					21	19	PCR	ANM		0	0.9 (0.05–15-47)‡	4.76									
Misch Misch Misch Misch 4 No. 1-10 (1) 4 Solid-1-9 (1) 33.3 3.3 3.4 4 No. 1-10 (1) 3.4	- E	57–61			63		PCR					30·16									
Seeding Hamiltonian Hamiltonia	٦				12	12	ISH	Other		0	5.5 (0.51–59.02)‡	33-33									
Climate Line 4 6 CRR + NRI CRR + NRI </td <td></td> <td></td> <td></td> <td></td> <td>10</td> <td></td> <td>PCR</td> <td></td> <td></td> <td></td> <td></td> <td>0</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td>					10		PCR					0									
Chinaty 4 109 200 6 5 FCR+NIT ANA 3 4 413 (13-1553) 6 3 4 413 (13-1553) 6 3 4 413 (13-1553) 6 3 4 413 (13-1553) 6 3 4 413 (13-1553) 6 3 4 413 (13-1553) 6 3 4 413 (13-1553) 6 3 4 413 (13-1553) 6 3 4			40		3	72	PCR + ISH	Other		43	0.67 (0.04–11.22)				23						
Ching Light 70 1997-200 40 15 CRC HIIC ANM 2 4 413(1/3-1525) 6 4 412(1/3-1525) 6 5 441(1/3-1525) 6 5 4 413(1/3-1525) 6 5 441(1/3-1525) 6 5 5 4 413(1/3-1525) 6 5 441(1/3-1525) 6 5 5 4 413(1/3-1525) 6 5 4 413(1/3-1525) 6 5 6 413(1/3-1525) 6 5 6 413(1/3-1525) 6 7 413(1/3-1525) 6 7 413(1/3-1525) 6 7 413(1/3-1525) 6 7 1<				1999–2000		36	PCR + ISH	Other		17	1.9 (0.82-4:36)	65.9									65.9
Claiming 35 104 CRR ANM 61 172 (1994) 2-301 288 41 172 (1994) 2-301 288 41 172 (1994) 2-301 288 41 172 (1994) 2-301 288 41 61		ad	70	1997–2000			PCR + IHC PCR + IHC ISH + SBA	ANM ANM ANM			4-13 (1-12–15-25) 15-44 (3-42–69-7)			4·12 (1·12–15·2: 15·44 (3·42–69·5	8 E						
Closure 61 1999-2004 67 CRAN AM 14 6 690-060-7-48 567 1 448 6 203-255 209 7 7 3 Greece 67 87 1895-2002 3 27 CARA 17 6 488 (143-1459) 566 7 1 960-05-1599 33 14 6 444 (152-1479) 566 7 1 660 7 444 (152-1479) 567 1 1 660 7				1999–2003		104	PCR	ANM		14	1.72 (0.99–2.99)										
Greece 67 87 1989-2002 30 27 FCR ANM 17 6 458 (143-1459) 5667 1 69 0045-1507) 33 13 15 44 (132-1477) 30 10 10 44 (132-1477) 30 10 10 44 (132-1477) 30 10 20		53	61	1999–2004		<i>L9</i>	PCR	ANM		9	2.69 (0.69–7.48)			1.72 (0.39–7.51)			2:03	2.99			
South Africat 67 84 1095-1008 60 10 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 66 (18-17) 67 (18-18) 67 (18-18) 7 8 8 9 <td>5</td> <td><i>L</i>9</td> <td>87</td> <td>1989-2002</td> <td></td> <td>27</td> <td>PCR</td> <td>ANM</td> <td></td> <td>9</td> <td>4.58 (1.43–14.59)</td> <td></td> <td></td> <td>0.9 (0.05–15-07)</td> <td>3.33</td> <td></td> <td>(0.16–22.93)‡ 4·4 (1·32–14·7)</td> <td>50</td> <td></td> <td></td> <td>53-33</td>	5	<i>L</i> 9	87	1989-2002		27	PCR	ANM		9	4.58 (1.43–14.59)			0.9 (0.05–15-07)	3.33		(0.16–22.93)‡ 4·4 (1·32–14·7)	50			53-33
South Africart 575 S	7 F	19	87	1989–2002		32	PCR	Other		9	5-66 (1-8-17-79)	26-67									
Nouth Africary South Africany South Africany South Africary South Africany South			28	1995-1998			PCR					94			4						
USA 66 64 1958-1988 22 PCR+ISH ANM 6 6 6 ASS-1988 73 ASS-1988 732-527) 732-527) 732-527) 732-527) 732-527) 732-527) 732-527) 732-527) 732-737		ca†			114	41	PCR	Other		3	10·62 (3·1–36·41)			2.22(0.26-19.04)							
Indiary Any	<u> </u>	99	64	1958-1988			PCR + ISH					45.45			7						
Irant 64 2005-2007 45 PCR 49-42 27-06 27-06 235 235 Irant 1.43 (0.53-20)7 45 18 0.71 (0.31-1-62) 25 7 1-43 (0.55-3-74) 8 0 3-12 (0.37-26)‡ 31 7 UK 68 34 34 12 1-43 (0.31-1-62) 0 1-43 (0.55-3-74) 8 0 3-12 (0.37-26)‡ 31 7				2001–2005		62	PCR PCR + SBH	ANM		3	1 (0·2–5·27)	0 7·32					1 (0·19–5·27)	7:32			
Irant 62 1997–1998 85 PCR Other 42 18 0.71 (0.31–1·G2) 25 7 1-43 (0.55–3·74) 8 0 3-12 (0·37–26); 31 7 UK 68 34 22 PCR+SBH 0 0 3-12 (0·37–26); 31 7	- ·	2		2005-2007			PCR					37.78									
	E :		62	1997–1998			PCR					49.42			27.06			2.35			29-41
UK 68 34 22 PCR+SBH						31	PCR	Other		18	0.71 (0.31–1.62)	44		1.43 (0.55-3.74)	_		3.12 (0.37–26)‡				
		89	34		22		PCR + SBH					0									

Table 1 (cont.)

									Non-	Non-specific HPV	1		HPV-16		Н	HPV-18		1	HPV-16/18		
Author	Country	Mean	Case % male	Data- Case % collection male year	Cas	es Contro	Detection Cases Controls method	Control	HPV +ve cases	HPV HPV +ve +ve cases controls	OR (95% CI)	Prev +	HPV HPV +ve +ve Cases Controls	OR ols (95% CI)	HPV Prev +ve (%) Case	HPV HPV +ve +ve Cases Controls	OR (95% CI)	Prev +	HPV HPV +ve +ve cases controls	OR (95% CI)	Prev (%)
6861	Japan				46	11	IHC	Other	∞	0	2·1 (0·23–18·86)‡	17-39									ĺ
	Japan	63	79	1983–1991	61		IHC					8.2									
Ogura,	Japan			1981–1991	4		PCR + SBH					7.14									
1993 [120] Ono, 1994	Japan	41–83	98	1982–1992	45		ISH					30.95			16.67			19.05			30-95
[121] Ostrow,	USA				5		ISH					0									
Pantelis,	Germany	28	96		53	53	PCR	ANM	6	6	1 (0·36–2·76)	16-98 6	9	1 (0·3–3·33)	11.32 3	8	1 (0·19–5·19)	5.66 9	6	1 (0.36–2.76)	16-98
2007 [03] Paz, 1997	USA			1982–1994	Ξ		PCR + SBH					0									
œ.	China†	57	47	1986	32	57	PCR	Other	2	4	0.88 (0.15–5.11)	6.25 1	- 1	1.81 (0.12–29.9)	3.12 0	-		_	-	1.81 (0.12–29.9)	3.12
2001 [64] Poljak,	Slovenia	8.85	100		20		PCR					0.1			10						10
	Slovenia	9-69		1985–1995	1117		PCR					0									
	China† Australia	58.1	08	2004-2005	60	99	ISH +IHC DBA	ANM	24	6	9.07 (3.48–23.64)	04									
1989 [126] Rugge,	Italy	19	100		18		PCR					0									
	Japan			1989–1994	92	18	PCR	ANM	0	0		0									
Shen,	China†			1994-1997	55	25	PCR	ANM	36	33	1.26 (0.58–2.74)	65.45						1	19 6	4.31 (1.56–11.88)	65-45
_	Japan				72		PCR + SBH					20.83			20.83						20.83
Shimada,	Japan			1981–1992	31		PCR + SBH					16·13									
	China†	19	81	1994-2005	59		PCR + SBH					32.2			25.42			5.08			30.51
	Hong Kong				319		PCR					13.48			12:23			1.88			13.48
Si, 2004 [131] Sitas, 2012 [149]	China† Australia, South Africa,	99	49		87 1561	1 2502	PCR ELISA	Other				20-69	13-3		20-69 35-0 29-9	φ.		3.45			20.69
	Brazil, Iran, China†				;		Ş					c									
Smits, 1995 [133]	The inemediands				60		PCK					0									
Sobti,	India†				52		PCR + ELISA					73.08									
Strickler,	USA	62	88	1975–1991	19		ELISA					5.26									
Suzuk,	China and	57-6-61-6			93		PCR + ISH					4.3			3.23						3.23
Syrjanen,	Finland		45	1969–1980	09		Histological					40									
	Japan			1982–1996	123		PCR + ISH					30-08									
	Italy	63	91	1989–1996	45		PCR					0									

Table 1 (cont.)

									Non-	specific HF	v		HPV-1	16			HPV-	-18			HPV-	16/18		
Author	Country	Mean age	Case % male	Data- collection year	Cases	Controls	Detection method	Control source	+ve		OR (95% CI)		HPV +ve Cases	+ve	OR (95% CI)	Prev (%)	+ve	HPV +ve Controls	OR (95% CI)		+ve		OR (95% CI)	Prev (%)
Togawa, 1994 [68]	Italy, France, Japan, Iran, USA, South Africa	35–91	81		72	33	PCR + SBA	ANM	10	0	5·16 (0·63–42·13)‡	13-89									10	0	5·16 (0·63–42·13)‡	13-89
Toh, 1992 [139]	Japan			1990–1990	45		PCR					6.67				2.22				4.44				6-67
Tornesello, 2009 [140]	Italy	61-3		1999–2003	36		PCR					27-78				1								1
Turner, 1997 [141]	Canada and USA	67	63		51		PCR					1.96				1-96								1.96
Van Doornum, 2003 [69]	The Netherlands	64-9	61	1989–1999	56	100	ELISA	Other	8	18	0.76 (0.31–1.88)	14-29	8	18	0.76 (0.31–1.88)	14-29					8	18	0.76 (0.31–1.88)	14-29
Van Rensburg, 1993 [70]	South Africa†				10	10	ISH	ANM	3	5	0.43 (0.07–2.68)	30												
Wang, 2010 [142]*	China and USA		71		435		PCR					56-09												
Weston, 2003 [71]	Brazil†	63	75	1999–2000	40	10	Hybrid Capture II	Other	1	1	0.23 (0.01-4.05)	2.5												
White, 2005 [143]	Kenya†	58	69		29		PCR					0												
Williamson, 1991 [72]	South Africa†	56			11	11	PCR	ANM	5	1	8·33 (0·78–89·47)	45-45												
Woo, 1996 [144]	Korea†			1991–1994	25		ISH + histological criteria					44												
Yang, 2008 [73]	China†		59	2000-2005	435	550	PCR	Other	308	253	2.38 (2.18-3.72)	70.8									308	253	2·85- (2·18–3·71)	70-8
Yao, 2006 [74]	China†	55·7–58			82	40	ICH+ISH	ANM	23	0	15-2 (1-97–117-23)‡	28.05									23	0	15·2 (1·97–117·23)‡	
Zhang, 2010 [75]	China†	57-32	70	2002-2006	70	60	PCR + IHC	ANM	35	20	2 (0.98-4.08)	50	21	13	1.55 (0.7–3.44)	30	8	3	2·45 (0·62–9·69)	11-43				
Zhang, 2011 [76]	China†	57-3	70	2000-2006	70	100	PCR	Other	28	8	7-67 (3-22–18-23)	40	28	8	7-67 (3-22–18-23)	40					28	8	7-67 (3-22–18-23)	40
Zhang, 2011 [77]*	China†	57-3	67	2000-2006	106	100	PCR	Other	82	33	6.94 (3.47–12.86)	77-36	61	22	4.81 (2.61–8.85)	77-36								77-36
Zheng, 1999 [145]	China†	46	67		3		PCR					0												
Zhou, 2003 [78]	China†	57-4	75		48	23	PCR + ISH + IHC	ANM	31	8	3-42 (1-21-9-69)	64-58	31	8	3·42 (1·21–9·69)	64-58					31	8	3·24 (1·21–9·69)	64-58

ANM, Adjacent normal mucosa; Other, other patients; PCR, polymerase chain reaction; SBH, Southern blot hybridization; ISH, *in-situ* hybridization; ELISA, enzymelinked immunosorbent assay; FISH, fluorescence *in-situ* hybridization; ICH, immunohistochemistry; DBA, dot blot analysis; SBA, slot blot analysis; hisFISH, histopathological-fluorescence *in-situ* hybridization.

^{*} All types of esophageal cancer.

[†] Classified as a developing country [146].

[‡] Insufficient data to calculate an OR as the number of HPV positive controls was 0 therefore in order to be able to calculate an OR, we added one HPV positive control sample.

[§] Adjusted ORs – Bjorge et al. [27] adjusted for continine and Dillner et al. [33] adjusted for smoking.

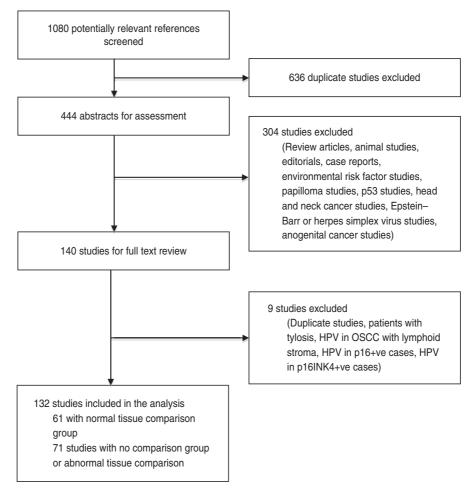


Fig. 1. Results of the literature search.

of heterogeneity, respectively [17]. We quantified publication bias using Egger's regression model [18], with the effect of bias assessed using the fail-safe number method. The fail-safe number was the number of studies that we would need to have missed for our observed result to be nullified to statistical non-significance at the P < 0.05 level. Publication bias is generally regarded as a concern if the fail-safe number is less than 5n+10, with n being the number of studies included in the meta-analysis [19]. We performed a sensitivity analysis based on sample size using different sample sizes (n < 10, n < 20, n < 50) and calculated the respective ORs. All analyses were performed with Comprehensive Meta-analysis version 2.0 (Biostat, USA).

Population attributable risk percentage (PAR%) was calculated using the formula:

$$PAR\% = \frac{HPV - positive \, cases \, (OR - 1/OR)}{all \, OSCC \, cases}.$$

The input data was taken from this meta-analysis.

RESULTS

From the literature search, we screened 1080 potentially relevant references, of which 444 abstracts were extracted for assessment (Fig. 1). Of these, 140 were selected for a detailed full-text review, and four studies were excluded - one included patients with tylosis, one studied HPV DNA presence in only p16-positive cases, one studied HPV DNA presence in only p16INK4-positive cases and one studied HPV DNA presence in OSCC with lymphoid stroma. There were five duplicate publications, we used the most recent in each case and the others were excluded. In total, 132 studies were included for analysis. There were two studies published by Moradi and colleagues [20, 21] that included the same case group; however, one study was a case-control study and was included in the analysis of risk while the other was a crosssectional study and was used in the prevalence analysis. We excluded the case-control study [21] from the prevalence analysis due to repetition of data. In the

Table 2. Prevalence of human papillomavirus in oesophageal squamous cell carcinoma

Factor	Prevalence, % (95% CI)
Global	
All studies	24.8 (21.2–28.8)
Region	
Africa	42.4 (34.0–51.3)
Asia	27.9 (22.6–33.8)
Europe	18.8 (12.8–26.7)
North America	15.2 (6.5–31.7)
Oceania	15.7 (3.6–48.2)
South America	16.3 (7.4–32.2)
Socioeconomic status	
Developing	31.1 (25.5–37.4)
Developed*	18.9 (14.8–23.8)
Country	
Australia	15.7 (3.6–48.2)
Brazil	8·1 (1·3–36·9)
China	32.8 (25.6–40.9)
Finland	24.8 (12.9–42.3)
France	2.8 (0.0–27.1)
Germany	26.2 (8.0–59.1)
India	26.6 (12.9–46.9)
Iran	26.9 (13.2–47.2)
Italy	16.6 (6.8–35.0)
Japan	20.7 (14.2–29.2)
Korea	26.5 (5.0–71.4)
Mexico	59.4 (6.5–96.9)
The Netherlands	2.9 (0.0–25.0)
South Africa	40.4 (28.5–53.5)
USA	8.7 (3.5–20.0)
HPV type	
16	16.8 (12.9–21.7)
18	8·1 (5·7–11·4)
16/18	23.0 (17.9–28.9)
Detection method	
Polymerase chain reaction	25.4 (20.9–30.4)
In-situ hybridization	22.6 (16.7–29.9)

CI, Confidence interval.

event that the same authors published multiple studies within a similar time period, we used information from the Materials, Methods and Results sections to confirm that these were from different sample populations. Sixty-one studies had control groups comprising of normal tissue samples and were used in both the risk and prevalence analyses [12, 15, 21–78]. The 71 other studies contained either no control group or samples from controls with benign or structural oesophageal disease and were used only in the analysis of the prevalence of HPV DNA in OSCC [10, 13, 14, 20, 79–145]. In total, 13 795 oesophageal samples were

included in the analysis, with 9291 as cases with established OSCC and 4504 as controls with normal oesophageal mucosa. The mean age of the patients was 59·36 (range of study means 45–67 years) and 70·1% were male (although data was only available in 82 studies) (Table 1).

Prevalence

From the 138 studies that assessed prevalence (n=12037 cases), 27.4% (95% CI 21.2-28.8) of OSCC samples were infected with HPV (Table 2). These studies contained cases from 30 countries and six continents (Table 1), which during subgroup analyses, demonstrated geographical differences.

When comparing the prevalence according to economic development, developing countries had a $12 \cdot 2\%$ higher rate of HPV DNA in OSCC than developed countries [31·1 (95% CI $25 \cdot 5$ –37·4) and $18 \cdot 9$ (95% CI $14 \cdot 8$ – $23 \cdot 8$), respectively]. We found that when analysing the prevalence by region, there was a $25 \cdot 9\%$ difference between the lowest, North America and the highest, Africa (Table 2). The largest population of people studied was Chinese (46 studies, n=5859 cases), where the prevalence was found to be $32 \cdot 8\%$ (95% CI $25 \cdot 6$ – $40 \cdot 9$), ranking third highest globally behind only Mexico and South Africa (Table 2).

Of the 54 studies that reported data on the prevalence of HPV-16 (n=4737 cases), 16·8% (95% CI 12·9–21·7) of cases contained HPV DNA, while of the 30 studies that reported data on HPV-18 (n=2272 cases), only 8·1% (95% CI 5·7–11·4) of cases contained HPV DNA. However, 56 studies looked at high-risk HPV types, 16 and/or 18, (n=4351 cases) and the combined prevalence was found to be 23·0% (95% CI 17·9–28·9).

There were 12 methods used to detect HPV in the samples (Table 1), the most commonly used being polymerase chain reaction (PCR) (94 studies) followed by *in-situ* hybridization (ISH) (24 studies), Southern blot hybridization (16 studies), immunohistochemistry (10 studies), enzyme-linked immunosorbent assay (8 studies), histological criteria (5 studies), dot blot analysis (4 studies), fluorescence ISH (3 studies), hybrid capture II (2 studies), slot blot analysis (2 studies), histopathological-fluorescence ISH (1 study), and the modified Feulgen technique (1 study). Thirty-seven studies used multiple detection methods, all of which included either PCR or ISH. In the 32 studies that included PCR, these were the results selected for analysis. In the remaining five studies, the ISH results

^{*} Countries classified as developing are identified in Table 1.

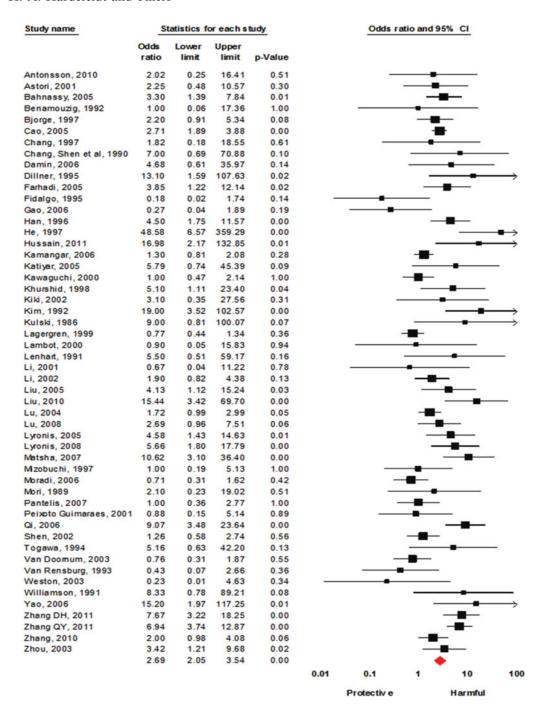


Fig. 2 [colour online]. Forest plot of association between the presence of human papillomavirus DNA and the development of oesophageal squamous cell carcinoma. There were six case-control studies that had no positive samples in either the case or control groups. Therefore these are not included in the forest plot.

were selected. The authors acknowledge that the use of histological criteria to diagnose HPV infection is a technique no longer employed. Two of the studies using histological criteria also utilized other detection methods in which cases the latter results were included; however, the remaining studies were included for a historical perspective.

Association between HPV and OSCC

Using the 61 studies (n=3970 cases) that contained a normal tissue comparison group, we calculated a pooled OR of 2·69 (95% CI 2·05–3·54) (Fig. 2), demonstrating an increased risk of developing OSCC in the presence of HPV infection. There was a moderate

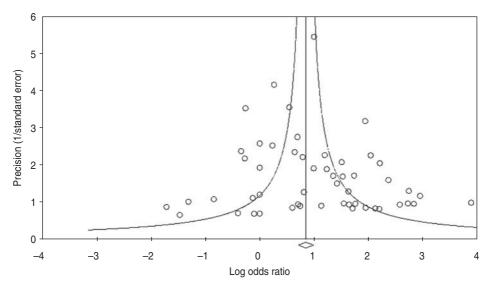


Fig. 3. Funnel plot of precision by log odds ratio.

level of heterogeneity ($I^2 = 63.82\%$, P < 0.001), and with a fail-safe n of 1675 (P = 0.15) there was no evidence of publication bias (Fig. 3). We performed a number of subgroup analyses, looking at the risk of specific types of HPV, the effect of differing control sources, geographical variance, and the method of HPV detection (Table 3).

There were 35 studies (n = 2942 cases) that provided information on specific types of HPV detected. We performed a subgroup analysis investigating the risk of HPV-16 and HPV-18, as well as combined HPV-16/18, in the development of OSCC. We found an increased risk of developing OSCC in patients with HPV-16 (OR 2·35, 95% CI 1·73–3·19) rather than HPV-18 (OR 1·28, 95% CI 0·87–1·88); however, a combined analysis of patients with HPV-16, HPV-18, or both resulted in a risk point estimate of 2·46 (95% CI 1·85–3·28). The heterogeneity was significantly high in all three analyses (Table 3).

Samples making up the control groups used in the studies were either biopsies of normal mucosa taken from an area of oesophgus adjacent to the tumour or from separate patients. We performed a subgroup analysis and found that studies that used separate patients as controls yielded a pooled OR of 2.98 (95% CI 1.95–4.57), which was higher than those studies that used adjacent samples (OR 2.51, 95% CI 1.72–3.68).

In concordance with the data assessing prevalence, there was a greater risk in developing countries of developing OSCC in the presence of HPV infection than in developed countries (Table 3).

Asia was the most predominantly studied region (36 studies, n=2738 cases) and we calculated a risk point estimate of 2.94 (95% CI 2.16–4.0). There was a significant difference between this and the second most studied region, Europe (13 studies, n=526), where the OR was found to be 1.64 (95% CI 0.90–2.99).

There were nine methods used in detecting HPV DNA in the samples (Table 1). PCR was the most commonly used method (41 studies), followed by ISH (10 studies), immunohistochemistry (8 studies), enzyme-linked immunosorbent assay (6 studies), Southern blot hybridization (4 studies), fluorescence ISH (2 studies), hybrid capture II (2 studies), dot blot analysis (2 studies) and slot blot analysis (2 studies). Sixteen studies used multiple detection methods, all of which included either PCR or ISH. In the 12 studies that included PCR, these were the results selected for analysis. In the remaining four studies, the ISH results were selected. We performed a subgroup analysis on the different detection methods, and studies using ISH resulted in the highest risk (OR 5.98, 95% CI 3.46-10.33), which was higher than the pooled risk of studies using PCR (OR 2.05, 95% CI 1·45–2·90).

PAR%

Using the formula discussed in the Methods section, the PAR% was calculated as a global figure as well as for differing regions. The results (Table 3) demonstrated that 15.7% of OSCC cases globally are

Table 3. Subgroup analysis of case-control studies

Factor	OR (95% CI)	I ² (P value)
All studies	2.69 (2.05–3.54)	63.82 (<0.001)
True Ors*	2.44 (1.82–3.27)	69·27 (<0·001)
Approximate ORs*	5·12 (2·67–9·81)	9.33 (0.35)
SES		
Developed	2.28 (1.40–3.73)	57.73 (0.001)
Developing	2.92 (2.08–4.11)	67.77 (<0.001)
Detection method		
Enzyme-linked immunosorbent assay	2.23 (0.76–6.53)	61.66 (0.02)
<i>In-situ</i> hybridization	5.98 (3.46–10.33)	0.00 (0.92)
Polymerase chain reaction	2.05 (1.45–2.90)	58.79 (<0.001)
HPV subtype		
16	2.35 (1.73–3.19)	64·14 (<0·001)
18	1.28 (0.87–1.88)	31.35 (0.11)
True 18 (excl. approximate)	1.14 (0.63–2.08)	51.46 (0.04)
16/18	2·46 (1·85–3·28)	61.04 (<0.001)
Control source		
Adjacent normal mucosa	2.51 (1.72–3.68)	65.47 (<0.001)
Other patients	2.98 (1.95–4.57)	64.17 (<0.001)
Region		
Africa	3.47 (1.02–11.84)	65.46 (0.03)
Europe	1.64 (0.90–2.99)	60.44 (0.005)
Oceania	3.84 (0.79–18.67)	0 (0.36)
South America	1.28 (0.07–23.82)	62.23 (0.1)
Asian analysis		
Asia	2.94 (2.16–4.00)	66.01 (<0.01)
All other continents	2·20 (1·34–3·61)	58·44 (<0·001)
China	2.85 (2.05–3.96)	67·20 (<0·001)

OR, Odds ratio; CI, confidence interval.

attributable to being HPV positive. Regional analysis revealed a higher percentage in Asia (19·0%) than in Europe (6·3%). When analysed by socioeconomic status, developing countries were found to have a higher PAR than developed countries.

DISCUSSION

The results of our meta-analysis highlight an almost threefold increased risk of OSCC in the presence of HPV infection. The study population included cases from a range of geographical areas, high- and low-incidence areas for oesophageal cancer and high- and low-prevalence areas for HPV infection.

We found HPV-16 to be both the most prevalent type detected as well as the highest risk in terms

of oncogenic potential. Of the studies with data on the various HPV types, we found HPV-16 to make up 65% of all HPV infections, and with an OR of 2.47 it was shown to be associated with the development of oesophageal cancer. HPV-16 infection was found to be twice as prevalent and almost double the risk for oesophageal cancer than HPV-18. Analysis of HPV-16 and/or HPV-18 showed that together these two types of HPV make up over 80% of HPV in infected samples, although the increase in risk is similar to that of HPV-16. These figures are in accordance with HPV infection in cervical squamous cell carcinoma, where HPV-16 is the most common type, followed by HPV-18 [9]. However, it has also been found that HPV-18 is the most predominant type found in cervical adenocarcinoma [146], and this

^{*} Approximate ORs refers to case-control studies that had insufficient data to calculate an OR as the number of HPV-positive controls was 0 (Table 1). In order to be able to calculate an OR, we added one HPV-positive control sample. True ORs refers to case-control studies that had sufficient data to calculate an OR and therefore these are an absolute representation of the study.

may also be the case for oesophageal adenocarcinoma although more research is needed to explore this potential link.

Control samples were either taken from normal oesophageal mucosa adjacent to the tumour site or completely different participants. A pooled subgroup analysis found studies using other participants to be have a higher risk than studies with adjacent normal mucosa as the normal control. One study included in our analysis, published by Benamouzig et al. in 1992 [26], included two control groups, 17 separate patients with normal oesophageal musoca as well as 12 biopsies of normal mucosa taken from adjacent to the tumour site of the case patients. Out of the total 29 control biopsies, only one of the adjacent normal mucosa samples tested positive for HPV DNA by ISH. However, three such biopsies were found positive by histological diagnosis. This was hypothesized as possibly being due to paraneoplastic impairment of the local oesophageal mucosal immunity [26]. As this effect can not be excluded, theoretically, separate patients as controls should yield a truer estimate of the risk. However, further research should be undertaken to evaluate this.

While Oceania and Africa were found to have the highest ORs, both groups had small numbers of studies (n=2 and n=4, respectively) and in terms of both the risk and prevalence analyses, the CIs overlapped considerably. This could be due to inter-region differences in baseline HPV prevalence or small sample sizes within the analysis. Therefore, these results are difficult to compare with regions such as Asia and Europe, which had much larger numbers of studies and while the CIs again overlapped, it was to a smaller degree. With this in mind, comparison of the results from Asia and Europe still demonstrated geographical variance. The risk for Asian populations was found to be higher than European populations. However, sample size may also play an important role again as it is important to note that the average sample size of cases in Asian studies is 76, compared to the average 40 of European studies. Interestingly, the geographical variance is in direct opposition to that of cervical cancer, which has a higher HPV prevalence in Europe, North America and Australia than Africa and Asia [9]. Given the questionable validity of these results without further research into general population prevalence of HPV as well as the interregion differences from larger studies, it is difficult to interpret the significance of the geographical differences noted.

Previous research has shown that OSCC is the second most common cause of death in Chinese males [38]. Chinese studies made up a large proportion of our meta-analysis (35 studies, n=3861 cases) and were found to have a pooled OR of 2.85. This put China at a greater risk than the global estimate of 2.69. The prevalence of HPV in OSCC in China was also found to be greater than the global estimate by 8%. A previous meta-analysis published in 2009 on the prevalence of HPV in OSCC in China found an average prevalence of 46.9% (95% CI 43.8-50.0) [147], which is considerably higher than that of the present analysis, where the results showed a prevalence of 32.8%. However, the 2009 meta-analysis had a number of limitations – it included only 15 studies, compared to the 23 of this study, included studies only using PCR in the detection of HPV and excluded all studies that were not in Chinese.

This study attempted to analyse the general population prevalence utilizing the separate control group data; however, only 33 studies could be included (as opposed to normal mucosa adjacent to the tumour site)—less than a third of the included studies. On a global scale 22.8% of controls were HPV positive. Regional analysis demonstrated that the estimated general population prevalence in Asia was 35.2%, Europe 19.8%, Africa 7.7% and 0% in Oceania. Given the number of studies utilized to gain these results (Asia n=19, Europe n=3, Africa n=2, Oceania n=1), it is difficult to assess the reliability of these results. A wider study of the general population is required before an accurate prevalence of HPV can be determined.

Due to the multiple detection methods used, a subgroup analysis was also performed to evaluate for any differences. Previous research has shown that in terms of sensitivity of HPV detection, ISH is almost equivalent to PCR, and in terms of reproducibility it is the superior detection method [148]. Therefore it is interesting to note that subgroup analysis showed that studies using ISH resulted in a pooled OR of 5.98 (95% CI 3·46–10·33), with minimal heterogeneity $(I^2 = 0.00, P = 0.92)$; however, PCR studies resulted in a much lower risk (OR 2.05, 95% CI 1.45-2.90) with a much higher heterogeneity $(I^2 = 58.79,$ p < 0.001). A possible explanation for this is the individual results in the ISH analysis as there were only seven studies included in the group, compared to the 41 in the PCR group.

The PAR% was also calculated based on data from this meta-analysis. The results showed that globally

15·7% of OSCC cases can be attributed to HPV. A limitation of this analysis is the use of the data from studies in this meta-analysis and a truer PAR% would be gained from using continent-wide statistics. This would require data on the incidence of OSCC throughout each region as well as oesophageal HPV positivity, which is not available at the current time. However, as a preliminary measure, our results demonstrate that 19·0% of OSCC cases in Asia can be attributed to HPV compared to 6·3% in Europe. Furthermore, Africa was found to have the highest PAR% at 28·7%; however, given the small number of studies from this region it is difficult to determine the significance – similarly for Oceania.

This meta-analysis has several strengths. A broad literature search was conducted with no language restrictions, a point which has been a limitation of previous meta-analyses on this topic. The search was also geographically unrestricted, which allowed for a broader insight into the global prevalence and risk associated with HPV and OSCC, as well as enabling the analysis of geographical variances.

This study also had several limitations. First, many of the studies included had small sample sizes - only 22 out of the total 131 studies had sample sizes over 100 cases. However, the risk ratios for different sample sizes (n < 10, n < 20, n < 50) were calculated and there was not statistically significant difference between them, suggesting that this had little impact on the combined estimates. Some analyses showed heterogeneity, which may have been due to small sample size. The use of adjacent normal mucosa as a control group also limited our ability to compare the results throughout the analysis. Only two of the studies provided ORs that had been adjusted for known risk factors for oesophageal cancer (i.e. alcohol and smoking), while the others provided only the raw data and unadjusted ORs were calculated. Therefore while this study was able to demonstrate an increased risk of OSCC in the presence of HPV infection, its independence as a risk factor is yet to be determined.

These results have demonstrated a considerable global association between the presence of HPV infection and the development of OSCC. They have implications for future research into the transmission of the virus to oesophageal mucosa as well as possible future treatment regimens for oesophageal cancer. On a regional scale, in areas such as Asia, where the incidence of OSCC is high, these results may have implications for immunization schemes and programmes

advocating for preventative measures to minimize the transmission of the virus.

In conclusion, this study has confirmed a relationship between OSCC and HPV infection. In addition, it has demonstrated an increased risk in developing countries, particularly China, as well as with HPV-16 infection compared to HPV-18. The prevalence of HPV DNA in OSCC was found to be higher in similar areas, and HPV-16 found to be the most prevalent globally, with the highest risk in comparison to other HPV types.

DECLARATION OF INTEREST

None.

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