Cardiology in the Young

cambridge.org/cty

Brief Report

Cite this article: Ehsan L, Thoe JA, Parent JJ, and Fakhoury JD (2024) Chylothorax related to acute SARS-CoV-2 infection in a patient with Noonan syndrome with prior uncomplicated cardiac surgeries. *Cardiology in the Young* 34: 448–451. doi: 10.1017/S1047951123004171

Received: 15 January 2023 Revised: 4 August 2023 Accepted: 17 November 2023

First published online: 22 December 2023

Keywords:

Chylothorax; chylous effusion; SARS-CoV-2 infection; Noonan syndrome

Corresponding author:

J. D. Fakhoury; Email: fakhourj@bronsonhg.org

© The Author(s), 2023. Published by Cambridge University Press. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted re-use, distribution and reproduction, provided the original article is properly cited.



Chylothorax related to acute SARS-CoV-2 infection in a patient with Noonan syndrome with prior uncomplicated cardiac surgeries

Lubaina Ehsan¹, Jessica A. Thoe², John J. Parent² and Joseph D. Fakhoury^{1,3}

¹Department of Pediatric and Adolescent Medicine, Western Michigan University Homer Stryker M.D, School of Medicine, Kalamazoo, MI, USA; ²Division of Pediatric Cardiology, Indiana University School of Medicine, Indianapolis, IN, USA and ³Pediatric Hospital Medicine, Bronson Children's Hospital, Kalamazoo, MI, USA

Abstract

SARS-CoV-2 is a novel coronavirus that has rarely been associated with chylothorax. Patients with Noonan syndrome are at risk for developing chylothorax, especially after cardiothoracic interventions. We present the case of SARS-CoV-2 infection triggering the underlying tendency of a patient with Noonan syndrome to develop chylothorax who did not develop it even after prior cardiothoracic interventions. Patient presented in respiratory distress without hypoxia and was found, on imaging, to have a large right-sided pleural effusion, which was eventually classified as chylothorax. The patient was then started on a low-fat diet. Chest tube drainage substantially reduced the effusion in size, and it remained stable. Our report highlights that SARS-CoV-2 infection can cause the development of a chylothorax or a chylous effusion in patients with Noonan syndrome or among populations with a similar predisposition. A high index of suspicion in vulnerable patients or those not responding to traditional therapy should exist with providers, thus leading to the testing of the fluid to confirm the diagnosis.

SARS-CoV-2 has shown to result in a spectrum of symptoms ranging from mild to severe respiratory failure, as well as life-threatening pneumonia and multi-organ failure. A few case reports have been published showing that SARS-CoV-2 is also rarely associated with chylothorax. Chylothorax results from thoracic duct damage with chyle leakage from the lymphatic system into the pleural space. SARS-CoV-2 is known to cause a cytokine storm with consequent hyper-inflammation and an arterial and venous vasculopathy associated with a prothrombotic state. The theory is that a prothrombotic state leads to venous circulation endothelial damage and then thoracic duct damage ultimately causing chylothorax. Patients with Noonan syndrome are known to have chylothorax either congenitally or as a spontaneous event that can also occur after a surgical intervention, typically a cardiac repair. Even though seemingly spontaneous, chylothorax in patients with Noonan syndrome is usually due to underlying lymphatic dysplasia in approximately 20% of the patients and involves the pulmonary and intestinal lymphatics. We present a case where SARS-CoV-2 infection triggered the underlying tendency of a patient with Noonan syndrome to develop chylothorax.

Case report

An 11-year-old male with a history of Noonan syndrome and pulmonary valve replacement (5 years prior to presentation) presented with 7 days of worsening cough, nasal congestion, shortness of breath, fever, and right-sided chest pain. A clinical diagnosis of Noonan syndrome was made based on the facial and cardiac phenotype at birth. Eventually, genetic confirmation was obtained by panel testing at 18 months of age and showed a pathogenic variant in RIT1: c.246T > A (p. Phe82Leu), a variant previously reported in other patients with Noonan syndrome. 10 He reported that his chest pain worsened with coughing and taking deep breaths. He had a positive at-home SARS-CoV-2 rapid antigen test, which was obtained 5 days before admission. On physical exam, he was haemodynamically stable without any additional respiratory distress. On auscultation, he had decreased breath sounds in the right lower lobe and a grade 2 out of 6 crescendo-decrescendo systolic murmur at the left upper sternal border. SARS-CoV-2 real-time polymerase chain reaction done at the hospital was positive. Chest X-ray showed extensive right middle lobe consolidation and right pleural effusion. The differential diagnosis included viral pneumonia secondary to SARS-CoV-2 infection with concern for secondary bacterial pneumonia, empyema, chylothorax, pulmonary embolism, or pulmonary tuberculosis. He was started on ceftriaxone due to concern for secondary bacterial pneumonia.

Infectious diseases, cardiology, and intensivist services were consulted. Chest CT with contrast showed a large right pleural effusion, right lower lobe opacity, and multiple bilateral pulmonary nodular opacities with ill-defined margins and ground-glass attenuation (Fig. 1).

Cardiology in the Young 449

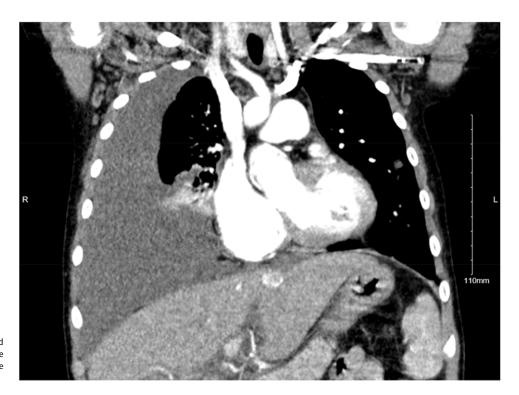


Figure 1. Chest CT coronal view obtained before chest tube placement showing a large right pleural effusion with right lower lobe capacity.

Other pertinent initial serum laboratory test results are as follows: normal procalcitonin (0.08 ng/mL, reference: <0.5 ng/mL), normal leukocytes (3.2 10*9/L, reference: 4–11 10*9/L), normal albumin (4.3 g/dL, reference: 3.5–5.0 g/dL), peripheral blood culture with no growth, and disseminated intravascular coagulation screen within normal limits.

A chest tube was then placed, which initially drained bloodtinged cloudy fluid and it turned milky over 3 days; this raised concern for chylothorax. Repeat chest CT with angiography showed decreased pleural effusion but with loculations for which the patient received pleural tissue plasminogen activator. Of note, chest CT with angiography did not show a pulmonary embolism. Serum laboratory tests performed within a day of chest tube placement showed pleural fluid to serum (pleural fluid: serum) total protein and lactate dehydrogenase ratios of 0.98 and 0.64, respectively. An echocardiogram was not concerning for an acute cardiac aetiology. It showed mild to moderate residual pulmonary stenosis with moderate to severe insufficiency (pulmonic valve mean gradient: 22 mmHg, peak, and peak gradient: 47 mmHg). The echocardiogram also noted a left anterior mitral valve leaflet with mild stenosis and no insufficiency, as well as systolic anterior motion of mitral valve. It also revealed normal left ventricle size with hyperdynamic function and mild subaortic thickening. There was no evidence of elevated right atrial pressures; as evidence by these findings, the right atrium was normal sized and the inferior vena cava was not dilated and had more than 50% collapse with respiration. The patient did not have a central line placement to permit a measurement of central venous pressure. Chest fluid cytology was negative for malignant cells. Chest fluid analysis also showed hypertriglyceridaemia (687 mg/dL). Chest fluid bacterial culture and acid-fast bacilli smear were negative. Pleural effusion met Light's criteria for being exudative based on pleural fluid:¹¹ serum total protein and lactate dehydrogenase ratios being greater than 0.5 (value: 0.98) and 0.6 (value: 0.64), respectively. It was further determined to be chylothorax given pleural hypertriglyceridaemia and chest fluid drainage with milky appearance.

The patient then received tissue plasminogen activator for 3 days to break up the loculations. He was also started on a low-fat diet. Based on repeat chest X-rays, right pleural effusion showed interval improvement in the initial 7 days after chest tube placement and then remained stable. After 9 days, the chest tube was removed when a goal of less than 35-mL drainage within 12 hours was reached. On outpatient follow-up, the pleural effusion remained stable for the next 4 months, and it was labelled as moderate-sized on chest X-ray (Fig. 2), without any new signs of respiratory distress.

Discussion

Chylothorax is a rare type of pleural effusion in the paediatric population, wherein the lymphatic fluid is found in the pleural space. Chylothorax can be caused by several broad categories, including congenital chylothorax associated with lymphatic malformations or syndromes, traumatic chylothorax following invasive procedure or other trauma, chylothorax related to elevated central venous pressure, chylothorax associated with tumours, or other miscellaneous causes. 12-14 Chylothorax is associated with many syndromes, including Down syndrome, Noonan syndrome, and Turner syndrome. ^{5–8,15} In Noonan syndrome, chylothorax may present as a congenital or spontaneous event and is also commonly seen in patients with Noonan syndrome after cardiothoracic surgical intervention.^{5–8} In one study of paediatric cardiac surgery admissions, patients with Noonan syndrome were 90% more likely to have postoperative chylothorax. 16 It is also interesting that our patient only had right-sided effusion, which may suggest a regional lymphatic abnormality or lymphatic system leak that occurred in the setting of SARS-CoV-2 infection. Recent literature using magnetic resonance lymphangiography has shown that the side of abnormal pleural perfusion does not always correlate with the side of chylothorax, which highlights the complex pathophysiology of lymphatic anatomy and physiology in patients with Noonan syndrome.¹⁷

450 L. Ehsan et al.



Figure 2. Follow-up chest X-ray in 4 months showing a moderate right-sized pleural effusion.

Chylothorax is identified by testing pleural fluid for triglycerides, like our case study. While not available for our patient, dynamic contrast magnetic resonance lymphangiography has been shown to be useful for patients with Noonan syndrome or chylous effusions to further assess the lymphatic system. 17-19 Assessment of abnormal pulmonary lymphatic perfusion via magnetic resonance lymphangiography allows the detection of pulmonary lymphangiectasia, which may be present in patient with chylothorax, although there have been patients with a chylothorax with normal underlying pleural lymphatic perfusion.¹⁷ Once identified, chylothorax can be treated with a low-fat diet, diuresis, and, if respiratory status dictates, fluid drainage.4 Octreotide use is also practised at paediatric centres, ⁴ although it was not used for our patient. In some circumstances of persistent chylothorax, lymphatic interventions may be indicated. In other cases, like Noonan syndrome, refractory chylothorax may be approached with a more targeted therapy like mitogen-activated protein kinase inhibition with trametinib. 18-20 Notably, due to the underlying complex lymphatic dysplasia and chylothorax pathophysiology among patients with Noonan syndrome, treatments such as lymphatic drainage, low-fat diet, and diuretics can be particularly ineffective, which is precisely why mitogen-activated protein kinase inhibition has been trialled in this patient population. 19,20

SARS-CoV-2 is a novel coronavirus that has caused a broad clinical spectrum from asymptomatic to critical illness with acute respiratory distress syndrome and even death. SARS-CoV-2 infections are rarely associated with chylothorax. In one case report, a young infant with spontaneous chylothorax was subsequently diagnosed with acute SARS-CoV-2 infection and died at 2 months of age due to pulmonary hypertensive crisis and respiratory failure. In another published case, a 78-year-old man diagnosed with acute SARVS-CoV-2 infection was found to have right pleural effusions later diagnosed as chylous. He had developed partial thrombosis in the superior vena cava, and the

distal right subclavian vein was thought to be the underlying cause of his chylothorax with resultant poor lymphatic drainage.¹

To our knowledge, this is the first reported case of a patient with Noonan syndrome having chylothorax related to acute infection with SARS-CoV-2. It is also important to note that our patient had normal estimated right atrial pressure based on an echocardiogram. While patients with Noonan syndrome can have chylothorax congenitally or spontaneously related to acute infectious processes or cardiothoracic surgery, it has not been associated previously with this specific infectious disease. Also curious in this case is that this patient has had cardiothoracic surgeries without prior history of chylothorax. There is no clear history of thrombus impairing lymphatic drainage, but the lymphatic malformations associated with Noonan syndrome coupled with systemic inflammation and cytokine storm shown to be underlying SARS-CoV-2 may have been enough to lead to chylothorax. This case demonstrates that while patients with Noonan syndrome are known to have a higher risk of chylothorax, the exact reason why a patient with Noonan syndrome will have a chylothorax in response to one process and not others is still not predictable.

Conclusion

Spontaneous chylothorax can occur in patients with a predisposition, including those with Noonan syndrome, during an acute respiratory infection such as SARS-CoV-2. An index of suspicion for chylous effusion should exist in Noonan syndrome patients with respiratory infections such as SARS CoV2 who develop effusions. Those not responding to traditional therapy that includes diuretic, low-fat diet, and octreotide should consider addition of mitogen-activated protein kinase inhibitor, particularly with ongoing need for chest tube drainage and respiratory support.

Acknowledgements. None.

Author contributions. Lubaina Ehsan and Jessica A. Thoe are contributed equally.

Drs. Ehsan and Fakhoury conceptualised the brief report.

Drs. Ehsan, Thoe, Parent, and Fakhoury were involved in the acquisition and interpretation of data for the report.

Drs. Ehsan, Thoe, Parent, and Fakhoury were involved in drafting the article and revising it critically for important intellectual content.

Drs. Ehsan, Thoe, Parent, and Fakhoury provided final approval of the version to be published.

Dr Fakhoury supervised the manuscript.

All authors approved the final manuscript as submitted and agree to be accountable for all aspects of the work.

Financial support. None.

Competing interests. None.

References

- 1. Satriano F, Scioscia G, Cagnazzo MG, et al. Chylothorax found in a patient with COVID-19. Respirol Case Rep 2022; 10: e0836.
- 2. Bezirganoglu H, Okur N. SARS-CoV-2 associated with death in an infant with congenital chylothorax. J Paediatr Child H 2022; 58: 536–538.
- 3. McGrath EE, Blades Z, Anderson PB. Chylothorax: aetiology, diagnosis and therapeutic options. Resp Med 2010; 104: 1–8.
- Agrawal A, Chaddha U, Kaul V, Desai A, Gillaspie E, Maldonado F. Multidisciplinary management of chylothorax. Chest 2022; 162: 1402–1412.

Cardiology in the Young 451

- C.H.A.N. DKL, Ho NK. Noonan syndrome with spontaneous chylothorax at birth. J Paediatr Child H 1989; 25: 296–298.
- Schlüter G, Steckel M, Schiffmann H, et al. Prenatal DNA diagnosis of Noonan syndrome in a fetus with massive hygroma colli, pleural effusion and ascites. Prenat Diagn 2005; 25: 574–576.
- Smith S, Schulman A, Weir E, Beatty D, Joffe H. Lymphatic abnormalities in Noonan syndrome: a case report. S Afr Med J 1979; 56: 271–274.
- Fisher E, Weiss E, Michals K, DuBrow I, Hastrieter A, Matalon R. Spontaneous chylothorax in Noonan's syndrome. Eur J Pediatr 1982; 138: 282–284.
- Roberts AE, Allanson JE, Tartaglia M, Gelb BD. Noonan syndrome. Lancet 2013; 381: 333–342.
- Aoki Y, Niihori T, Banjo T, et al. Gain-of-function mutations in RIT1 cause Noonan syndrome, a RAS/MAPK pathway syndrome. Am J Hum Genet 2013; 93: 173–180.
- 11. Light RW, Macgregor MI, Luchsinger PC, B.A.L.L. JR, WC. Pleural effusions: the diagnostic separation of transudates and exudates. Ann Intern Med 1972; 77: 507–513.
- Dubin PJ, King IN, Gallagher PG. Congenital chylothorax. Curr Opin Pediatr 2000; 12: 505–509.
- 13. Rocha G, Fernandes P, Rocha P, Quintas C, Martins T, Proença E. Pleural effusions in the neonate. Acta Paediatr 2006; 95: 791–798.

- Van Aerde J, Campbell AN, Smyth JA, Lloyd D, Bryan MH. Spontaneous chylothorax in newborns. Am J Dis Child 1984; 138: 961–964.
- Yamamoto T, Koeda T, Tamura A, et al. Congenital chylothorax in a patient with 21 trisomy syndrome. Pediatr Int 1996; 38: 689–691.
- Kriz C, Flores S, Villarreal EG, Bronicki RA, Loomba RS. Impact of Noonan syndrome on admissions for pediatric cardiac surgery. Minerva Pediatr 2019; 74: 461–467.
- 17. Pieper C, Wagenpfeil J, Henkel A, et al. MR lymphangiography of lymphatic abnormalities in children and adults with Noonan syndrome. Sci Rep-UK 2022; 12: 11164.
- Dori Y, Smith C, Pinto E, et al. Severe lymphatic disorder resolved with MEK inhibition in a patient with Noonan syndrome and SOS1 mutation. Pediatrics 2020; 146: 146.
- Nakano TA, Rankin AW, Annam A, et al. Trametinib for refractory chylous effusions and systemic complications in children with Noonan syndrome. J Pediatr 2022; 248: 81–88.e1.
- Gordon K, Moore M, Van Zanten M, et al. Case report: progressive central conducting lymphatic abnormalities in the RASopathies. Two case reports, including successful treatment by MEK inhibition. Front Genet 2022; 13: 2720.