Glycaemic index of cereal products explained by their content of rapidly and slowly available glucose

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Glycaemic index: Insulinaemic index: Dietary carbohydrate: Carbohydrate quality:
Cereal products

Since its development in 1981, the glycaemic index (GI) has had a pivotal role in highlighting the variation in physiological responses associated with different carbohydrate-containing foods (Jenkins et al. 1981). This ranking of foods by the glycaemic responses elicited when equi-carbohydrate portions are consumed has provided a unique, and at times controversial, perspective on the issue of carbohydrate quality (Wolever, 1997; Bellisle, 2001). Low-GI diets have been successfully applied as a dietary therapy in diabetes mellitus and other conditions exhibiting derangements in carbohydrate and lipid metabolism (Brand-Miller, 1994). In these studies, the major dietary alterations were to the starch-containing foods, with the substitution of slowly digested low-GI products, such as pasta, wholegrain cereal and legumes, for rapidly digested high-GI products, such as bread, breakfast cereals and potatoes.

The relationship between the rate of starch digestion and GI has been established by investigations of in vitro amylolytic hydrolysis (O’Dea et al. 1981; Jenkins et al. 1982; Heaton et al. 1988; Bornet et al. 1989; Englyst et al. 1992; Granfeldt et al. 1992). The rate and extent of starch digestion is influenced by botanical origin as this determines the amylose:amylopectin ratio and the structural type of the starch granule (Gallant et al. 1992). The other important factor is food processing, which determines the extent of starch gelatinisation, particle size and the integrity of the...
plant cell wall (Heaton et al. 1988; Holm et al. 1988; Colonna et al. 1992; Holt & Brand-Miller, 1994; Heijene et al. 1995). These physico-chemical variables of starch-containing foods are difficult to characterise in a quantitative manner that relates to their likely physiological fate. Instead, the influence of such physico-chemical characteristics on the rate and extent of carbohydrate digestion can be measured, and this can then be used to provide a description of this nutritionally important aspect of the food.

In conjunction with studies on human subjects, we have developed analytical procedures that characterise dietary carbohydrates with regard to chemical composition and likely gastrointestinal fate (Englyst et al. 1992, 1999; Englyst & Hudson, 1996). The glycaemic carbohydrate fraction that is available for absorption in the small intestine is measured as the sum of sugars and starch, excluding resistant starch (RS). We have divided the glycaemic glucose fraction (sum of glucose in the glycaemic carbohydrate fraction, but excluding lactose) into rapidly available glucose (RAG) and slowly available glucose (SAG), to reflect the likely rate of release and absorption of glucose.

In addition to the rate of carbohydrate digestion, food-mediated effects on both gastrointestinal events and post-absorptive metabolism can influence the GI. Gastric emptying is affected by food particle size (Thomsen et al. 1994) and fat content (Gannon et al. 1993), as well as by viscous fibre, which also limits enzymatic hydrolysis in the small intestine by restricting access to the food bolus (Jenkins et al. 1978). Post-absorptive factors that can influence GI include the identity of the sugar moieties, which are metabolised differently (Lee & Wolever, 1998), and the insulinotropic effect of protein, which can increase the clearance rate of circulating glucose (van Loon et al. 2000). This emphasises the fact that GI values do not represent a direct measure of carbohydrate absorption from the small intestine. Rather, the GI values are determined by the combined effect of all the properties of a food that influence the rate of influx and removal of glucose from the circulation. A better understanding of the mechanisms involved should provide insight into the concept of GI, and help to establish whether different types of low-GI diets are equally beneficial to health.

As cereal grains are the largest contributor to carbohydrate intake, it follows that altering the carbohydrate quality of cereal products is likely to have the most tangible effect on this aspect of nutrition. Considerable choice and flexibility exists for the consumer in their selection of cereal products based on the grain type, degree of refinement and type of processing (Prochaska et al. 2000). Unfortunately, with a few notable exceptions, the food-processing techniques employed in the manufacture of cereal products tend to result in the disruption of the food matrix and the gelatinisation of starch granules, thereby making them readily digestible and consequently they generally have high GI values. The challenge is to identify techniques for cereal processing that result in starch that is slowly digested, thereby achieving low-GI products.

Previously, we have shown that for a limited number of predominantly starchy foods, the division between RAG and SAG has physiological significance with regard to glycaemic response (Englyst et al. 1999). In the present paper, we extend our investigations on the correlations between the chemical and carbohydrate digestibility characteristics of cereal products and their GI and insulinemic index (II) values. These relationships need to be established in order to achieve a greater understanding of the role of carbohydrate quality in nutrition.

Subjects and methods

Test meals

Twenty-three cereal products, selected for investigation on the basis that they may be interchangeable within a breakfast meal or snack, were collected from different countries (Table 1). The products encompass a range of ingredients and processing techniques that contribute to defining their physico-chemical properties. The predominant cereal present in these products is wheat, with a few exceptions where maize (cornflakes), rice (Special K) or oats (Alpen) represent the sole or major cereal component. Apart from the Alpen muesli, which contained steamed rolled oat kernels (and some dried fruit and nuts), the products were produced from flour and did not contain dense matrices. The other products in the breakfast-cereal group (BC) comprised flakes of extruded cereals, which are low in fat, with a moderate content of sugar. The bakery-product and crackers group (BP&C) included two baguette meals and two brioche-type products where fat and milk are present during the baking process. The crackers were included within group BP&C, as these are also produced by baking in the presence of fat and moisture. The biscuit group (Bi) was on average moderately high in fat and sugar, and in contrast with the other groups, several of the biscuits were baked under low-moisture conditions, which restricts the extent to which starch granules are gelatinised.

Glycaemic index determination

The GI values of the selected cereal products were determined using the previously described standard protocol (Wolever, 1991; Food and Agriculture Organization/World Health Organization, 1998). The results were obtained from a series of seven sets of GI determinations, each of which included between eleven and fourteen healthy subjects. Briefly, subjects who had maintained an overnight fast were fed the test product in a portion size that was calculated to contain 50 g carbohydrate (as determined by an initial analysis of total starch and sugar content). Within the individual sets, each subject was given the products being tested once in a randomised order. In addition, on three separate occasions each subject also consumed a 50 g glucose solution as the reference meal (anhydrous glucose (dextrose); Sigma Chemical Company, St Louis, MO, USA). Blood samples were taken before and 15, 30, 45, 60, 90 and 120 min after each meal commenced. Plasma glucose concentrations were measured in duplicate using an enzymatic method (Roche Diagnostica, Basle, Switzerland) and insulin was measured by radioimmunoassay. Incremental areas under the blood glucose response curves were calculated using the trapezoid rule, with only the area above the
### Table 1. Nutritional composition and physiological characteristics of twenty-three cereal products

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Glycaemic indices of cereal products

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‡The nutritional composition of the cereal products is expressed per portion size used in the physiological studies. The carbohydrate component was characterised according to a number of different chemical and digestibility characteristics used to investigate which best described the variance in the physiological responses.

§ GI and II values were determined according to standardised methodology based on test meals of portion sizes calculated to provide 50 g carbohydrate. A solution of glucose was used as the reference meal, to which the responses to the other test meals were related (hence without units).

{Mean values and standard deviations for comparison between product groups.}
baseline being included. For each subject, a GI value for the test food was calculated as the incremental area under the blood glucose response curve for the test food, expressed as a percentage of the average incremental area under the blood glucose response curve for the three glucose reference test meals. The GI value of a product was calculated as the average of the GI values for that product obtained from the individual subjects. Calculation of II values for the test foods followed the principles utilised in the GI calculations.

**Dietary carbohydrate analysis**

Carbohydrate analysis of the products was performed using previously described procedures (Englyst et al. 1994, 2000). For the measurement of RAG, SAG and starch fractions, samples were minced using the specified procedure to simulate buccal mastication. Portions of the samples containing approximately 500 mg carbohydrate were weighed into 50 ml centrifuge tubes together with an internal standard. After an initial treatment with pepsin, the samples were incubated with a mixture of hydrolytic enzymes under controlled conditions of pH, temperature, viscosity and mechanical mixing. Subsamples were taken from the incubation mixture exactly 20 and 120 min after the commencement of the hydrolysis. RAG and SAG values were calculated as the glucose released at 20 min and between 20 and 120 min respectively. Any starch remaining in the main incubation tube was dispersed and hydrolysed, with the increase in released glucose calculated as the RS fraction. Free sugar glucose and fructose (including that derived from sucrose) were determined after extraction procedures and incubation with invertase.

Five models describing different chemical and digestibility properties of the carbohydrate components were constructed from the analytical fractions. These were developed in a progressive fashion, with each successive carbohydrate model based on the previous one: model 1, starch and sugar; model 2, starch, with the sugar fraction divided into total fructose and free sugar glucose components; model 3, fructose and the total glucose fraction calculated as the sum of free sugar glucose and glucose from starch; model 4, fructose, with the total glucose fraction divided into glycaemic glucose and RS; model 5, fructose and RS, with the glycaemic glucose divided into RAG and SAG.

To investigate the relationships between the physicochemical characteristics of foods and their in vitro digestibility profiles, three products from each group had their extent of starch gelatinisation determined by scanning differential calorimetry (Biliaderis et al. 1980). The results were expressed as the starch gelatinisation index (extent to which starch is gelatinised, with 100 representing total gelatinisation).

**Statistical analysis**

Student’s t test was used for comparisons between cereal product groups; a P value < 0.05 was considered significant. The relationships between nutrients and with the GI and II were investigated by univariate correlations. The total variance in GI and II that each of the carbohydrate models 1–5 could explain was investigated by ANOVA. The variance explained by fat, protein and NSP was also investigated. From these findings, simplified models were developed that best described the variance in GI and II values of the cereal products by their chemical and digestibility characteristics. Statistical analysis was performed with SPSS (version 9; SPSS Inc., Chicago, IL, USA).

**Results**

**Characteristics of the cereal products**

Table 1 shows the nutrient compositions of the twenty-three foods that were investigated in the physiological studies, expressed per portion size, and with the corresponding GI and II values. There were significant differences between the three groups of products in their chemical and digestibility characteristics. There was less fat in BC than the other groups, and less protein and starch and more sugar in Bi than in BP&C. The differences between groups extended to the detailed carbohydrate fractions, where Bi contained less RAG and more SAG than BC, and Bi contained more total fructose than BP&C. Bi had significantly lower GI values than BP&C and BC. There was no significant difference in II values between groups. There was a positive correlation between GI and II values (Fig. 1), although II values were significantly higher (P < 0.001) than GI values.

The starch gelatinisation indices for the subset of products in the BC group were: Energy Mix 99, Chocapic 100, Special K 100. For the BP&C group, the values were: French baguette with butter + jam 98, French baguette with chocolate 98, pain au lait 100. For the Bi group...
the values were: P’tit déjeuner miel et pépites 56, Principe megamanana vanilla 50, Véritable petit beurre 40. Fig. 2 shows that the Bi group had the lowest mean GI value, with the highest mean SAG content and the lowest mean starch gelatinisation index compared with the BC and BP&C groups.

**Relationships between nutrients**

In order to understand the statistical models that explain the variance in GI and II values, it is essential to establish the relationships that exist between the different carbohydrate fractions and the other nutritional components. Due to the
design of the present study, with the carbohydrate content of the test meals fixed at 50 g carbohydrate, there is necessarily an interaction between those carbohydrate fractions described in the five models where a reciprocal relation with total carbohydrate exists, e.g. starch:sugar (model 1), total glucose:fructose (model 3). In addition, because the models build progressively on one another, a number of other relationships between carbohydrate fractions could be explained by their close links with other associated analytical fractions in different models, e.g. sugars:free sugar glucose:fructose (models 1 and 2), starch:total glucose:glycaemic glucose:RAG (models 1–5).

A summary of the significant univariate correlations between nutrients is shown in Table 2, which focuses on the RAG fraction in order to demonstrate its relative associations with other carbohydrate components. The positive correlation between starch and RAG in these products can be explained by the high proportion of the starch that is included in the RAG fraction (mean value 79·1% (range 55·3–94·3)%), compared with the SAG (mean value 17·5% (range 2·2–41·2)%), or RS (mean value 3·4% (range 0·0–8·9)%) fractions. The total glucose and glycaemic glucose fractions, which combine the glucose components from starch and sugar, have strong positive correlations with RAG, reflecting the fact that an average of 83·5% total glucose in the products is included in the RAG fraction.

In model 5, RAG was correlated with total fructose, SAG and RS, with the SAG fraction demonstrating the strongest relationship. No significant relationships were observed between SAG and either starch or sugar, indicating that the SAG content of the products must be determined by the food-processing technique. Whilst the positive correlation between RS and starch is to be expected, there was a negative correlation between RS and NSP. There was no significant correlation between NSP and starch contents, and this finding remained even after the exclusion of Alpen, with its exceptionally high NSP value (in part due to its fruit and nut components), from the analysis.

The correlation of fat with RAG and SAG in these products is of particular interest. To some extent, this relationship reflects the different characteristics of the product groups, with BC having significantly lower fat and SAG contents than Bi, and with BP&C intermediate in SAG but relatively high in fat content. Even so, positive correlations between fat and SAG were apparent within each group, but did not reach significance due to the small sample sizes. RAG was also positively correlated with protein content.

### Relationships with the glycaemic and insulinaemic indices

In order to explain the variance in GI and II values, their correlation with the chemical and digestibility characteristics of the cereal products was investigated. The relationships between the carbohydrate fractions and the GI values are shown in Fig. 3, together with the total variance explained by each of the models 1–5. Neither starch nor sugar in model 1 were significantly correlated with GI, as was the case for free sugar glucose (model 2) and fructose (models 2–5). The positive correlations of GI with total glucose (model 3) and RS (models 4 and 5) can be explained by the analytical association that exists between these carbohydrate fractions and the RAG fraction (Table 2). GI was strongly correlated with both RAG and SAG contents in model 5.

The variance in GI that could be explained by the different carbohydrate models ranged from 33·8% for model 1 to 68·8% for model 5 (Fig. 3). The combined effect of fat, protein and NSP was to describe 59·1% of the variance in GI, but of these, only the fat content was significant ($r^2$ 0·52, $P<0·01$). The simplified model that best described the variance in GI combined the strongest variables, SAG and fat, and accounted for 73·1% (Fig. 4). The partial correlations of fat and SAG with GI within this model suggest that SAG is the dominant factor in comparison with fat.

The relationship between the carbohydrate fractions and the II values is summarised in Fig. 3, together with the variance in II explained by each of the models 1–5. Total fructose exhibited the strongest negative correlation and, through association, explained the correlations observed with sugars and free sugar glucose. Positive correlations were observed for total starch, total glucose and glycaemic glucose, all of which can be related to their association with RAG, which had the strongest positive correlation. A negative correlation between II and SAG was observed.

The variance in II that could be explained by the different carbohydrate models ranged from 27·6% for model 3 to 41·1% for model 5 (Fig. 3). Fat, protein and NSP together accounted for 46·4% of the variance, the majority of which was explained by the positive correlation with protein ($r^2$ 0·33, $P<0·01$). The simplified model that best described the variance in II combined RAG and protein, and accounted for 45·0% (Fig. 4), with RAG and protein having similar partial correlations within this model.

### Discussion

The present study has investigated the relationship between the chemical and digestibility characteristics of a selection of cereal products and their physiological properties (GI and II). Various models were investigated, which

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**Table 2. Selected relationships between the nutritional components of the cereal products investigated**

<table>
<thead>
<tr>
<th>Protein</th>
<th>Starch</th>
<th>RS</th>
<th>RAG</th>
<th>SAG</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fat</td>
<td>0·462*</td>
<td>−0·620**</td>
<td>−0·465*</td>
<td>0·578**</td>
</tr>
<tr>
<td>Protein</td>
<td>0·462*</td>
<td>0·458*</td>
<td>0·706**</td>
<td></td>
</tr>
<tr>
<td>Starch</td>
<td>0·469*</td>
<td>0·465*</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sugar</td>
<td>−0·567**</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fructose</td>
<td>−0·573**</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FSG</td>
<td>−0·558**</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total glucose</td>
<td>0·567**</td>
<td>0·816**</td>
<td>−0·427*</td>
<td></td>
</tr>
<tr>
<td>Glycaemic glucose</td>
<td>0·803**</td>
<td>−0·443*</td>
<td></td>
<td></td>
</tr>
<tr>
<td>RS</td>
<td>0·513*</td>
<td></td>
<td>−0·419*</td>
<td>−0·866**</td>
</tr>
<tr>
<td>RAG</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

RS, resistant starch; RAG, rapidly available glucose; SAG, slowly available glucose; FSG, free sugar glucose.

$P<0·05$. $^{**}P<0·01$.

† For details of products and procedures, see Table 1 and p. 330.
characterised the carbohydrate fraction progressively from solely chemical attributes in model 1 (starch and sugars) to a profile of the rate and extent of release in model 5 (fructose, RAG, SAG, and RS). The division of the glycaemic glucose fraction into RAG and SAG fractions (model 5) was found to describe 68.8% of the variance in GI, compared with 33.8% explained by the division into starch and sugars.

The twenty-three products investigated encompass a range of ingredients and processing techniques that contribute to defining their physico-chemical properties. Although starch is the main constituent of the products investigated in the present study, they also had a range of sugar, fat, protein and NSP contents. Each of these components has been found to influence glycaemic response, with much of this evidence based on observing the effects of addition of the component of interest to carbohydrate meals (Jenkins et al. 1978; Gannon et al. 1993; van Loon et al. 2000). The few studies that have specifically investigated the relationship between nutrient composition and GI value can be difficult to interpret as they incorporate a varied range of food groups.

For example, Trout et al. (1993) identified a negative relationship between protein content and GI, but this disappeared when the legume group, with its high-protein content and low GI value, was considered separately from the rest of the products. The present study was limited to the investigation of processed cereal products, thereby eliminating any interference associated with a wider range of food groups. This focus on cereal products did not restrict the range of GI and II values, but rather illustrated that the type of cereal product consumed can have a marked influence on physiological responses.

For the products investigated, the determinants of the digestibility profile of the carbohydrate fraction are the starch:sugar ratio used, the cereal type and the degree of food processing. The production of the breakfast cereals and bakery products investigated involves heating in the presence of moisture, which results in the gelatinisation of starch and its consequent rapid digestion. In contrast, several of the biscuits investigated were produced by baking under very-low-moisture conditions, which reduces the extent of starch gelatinisation and results in partially intact starch granules that are less susceptible to

![Fig. 3. Relationships between (A) glycaemic index and (B) insulinaemic index of twenty-three cereal products and their carbohydrate composition expressed as five models describing different chemical and digestibility characteristics. FSG, free sugar glucose; RS, resistant starch; RAG, rapidly available glucose; SAG, slowly available glucose. For details of subjects, cereal products and procedures, see Table 1 and p. 330. The univariate correlation coefficients (r) for the individual carbohydrate fractions are provided alongside their labels *P < 0.05, **P < 0.01. The variance explained by each full model is derived from multiple covariate regression analysis (r^2 values).](https://doi.org/10.1079/BJN2002786)
Fig. 4. Simplified models of the variance in glycaemic index (GI; (A), (B), (C)) and insulinaemic index (II; (D), (E), (F)) of twenty-three cereal products, explained by their chemical and carbohydrate digestibility characteristics. SAG, slowly available glucose; RAG, rapidly available glucose. (A), GI model with SAG + fat; GI = 85.8 – 2.335 × SAG + 1.493 × fat, $r^2 = 0.731$; (B), SAG (within GI model), $r^2 = 0.445$; (C), fat (within GI model), $r^2 = 0.272$; (D), II model with RAG + protein; II = 35.8 + 0.821 × RAG – 2.624 × protein, $r^2 = 0.450$; (E), RAG (within II model), $r^2 = 0.178$; (F), protein (within II model), $r^2 = 0.184$. For details of subjects, cereal products and procedures, see Table 1 and p. 330.
the action of amylolytic enzymes (Bornet et al. 1989). This relationship between a low extent of starch gelatinisation and high SAG values was confirmed in the present study by the low starch gelatinisation index for the Bi group compared with the BP&C group (Fig. 2).

Several of the relationships between nutrients identified in the present study are of interest, both in characterising the overall physico-chemical profile of the products, and in explaining observed correlations with GI and II values. For instance, the positive correlation between RS and GI is difficult to explain on its own, as the RS fraction is not absorbed in the small intestine and therefore cannot elicit a glycaemic response. Indeed, a negative relationship between RS content and GI has been reported, though the high RS products in that study contained 30–50 % starch as RS (Björck et al. 2000), compared with an average RS content of 3–5 (range 0–9/0) % starch in the present study. Our present finding of a positive correlation between RS and GI is explained by the positive relationship that exists between RS and RAG in these products. This is typical of products where starch has been gelatinised, with the majority being readily digestible, except for the small amount of retrograded starch that resists digestion.

In the present study, the inter-relationship between RAG and SAG makes it difficult to identify which of these factors explains the strong correlations that they have with GI. However, we have previously shown that it is RAG, by virtue of its rapid digestion and absorption in the small intestine, that is responsible for the postprandial rise in blood glucose concentrations (Englyst et al. 1999). It follows that SAG exerts its reductive effect on GI values by replacing RAG in the test meal. An increase in the proportion of total fructose in a test meal will also result in a reduction in RAG and explains why foods high in fructose or sucrose have been reported to have only moderate GI values (Brand-Miller et al. 1995). In the present study, a correlation between fructose and GI was not observed, despite the fact that it replaced part of the RAG fraction. Consequently, this is likely to explain why the RAG fraction described only 55 % of the variance in GI compared with the 63 % explained by the SAG fraction. Despite its low GI value, high intakes of fructose are generally not associated with adverse effects on blood glucose concentrations (Englyst et al. 1999). This seems probable, therefore, that the greatest health benefits of low-GI diets will accompany those that contain carbohydrates that are slowly digested and absorbed.

The 51.5 % variance in GI described by fat content is not fully explained by the association between fat and SAG, which together described 73 % of the variance in GI (Fig. 4). Numerous studies have demonstrated that fat can lower the glycaemic response to foods (Welch et al. 1987; Collier et al. 1988; Gannon et al. 1993). However, a recent study found that while the addition of 40 g fat to a 75 g carbohydrate meal of pasta delayed the appearance of exogenous glucose in blood, this effect was either inconsistent or masked by other factors, as no significant correlation between fat and GI was observed. The finding that the two high-fat baguette meals had moderately high GI values lends further support to the suggestion that fat per se only has a minor effect on GI in the present study.

Several studies have included measures of II values, which are of interest due to the role of insulin in glucose homeostasis and its regulatory effects in lipid metabolism. In addition, the large insulin demand associated with high-GI diets has been proposed to be involved in the aetiology of diabetes (Salmeron et al. 1997; Wolovere, 2000). The present study confirms the relationship between GI and II values for starchy foods, although the correlation is not as strong as reported previously (Björck et al. 2000). It is possible that this rather weak association, and the observation that II values were higher than GI values, could be explained by the combined insulinoergic effects of protein, fat and possibly of other undetermined properties of the foods. This is supported by the investigation into the glycaemic and insulinaemic responses to 1000 kJ portions of a range of foods (Holt et al. 1997). Holt et al. (1997) could only explain 23 % of the variance in the insulin score by the glycaemic score of the foods, and only a further 10 % could be accounted for by the macronutrient composition. Of the carbohydrate fractions investigated in the present study, RAG demonstrated the strongest correlation with II, but still only explained 32 % of the variance. The observed positive correlation between protein and II is in agreement with previous findings (Kabadi, 1991; Trout et al. 1993; Brand-Miller et al. 1995), although the association between RAG and protein may in part explain this relationship. The model incorporating RAG and protein accounted for 45 % variance in II, considerably less than could be identified for GI.

The present study has demonstrated that RAG and SAG, describing the rate of carbohydrate release from foods, are the carbohydrate fractions that best describe the variability in physiological attributes of the cereal products investigated. Although there appeared to be independent effects of fat on GI and of protein on II, these relationships may in part be explained by their correlations with RAG and SAG. The effect of protein and fat should not be completely dismissed, and indeed their presence in the foods investigated is probably responsible for some of the variance observed in GI and especially II values. This acts to highlight further the consistency of the effect of RAG and SAG on the physiological responses, in spite of the complexity of factors that can have an influence. It is apparent from the present study that food processing is the major determinant of the state of gelatinisation of starch and the SAG content of cereal products. The Bi
group had the highest mean SAG content, although this group also exhibited the greatest range in the proportion of SAG in the starch fraction, reflecting the diversity in processing methods used in biscuit production. This demonstrates that limiting the extent of starch gelatinisation, as occurs with some types of biscuit manufacture, represents a feasible method by which to lower the GI values of cereal products.

In conclusion, the present paper has shown that carbohydrate identity and food processing largely determine the variation in GI values of cereal products, and that this is adequately reflected by the classification scheme describing the rate of carbohydrate release from foods. Specifically, the SAG measurement allows the identification of those low-GI foods containing carbohydrates that are slowly digested and absorbed, for which health benefits are likely to be associated. We suggest that the proposed classification scheme would be valuable in the further elucidation of the mechanisms by which carbohydrate quality can influence health.

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References


