The validation of using serum iron increase to measure iron absorption in human subjects

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The objectives of the present study were to study the correlation between the change in serum Fe and Fe absorption when administering 100 mg Fe (as FeSO4) orally, and to study the correlation between the absorption from a 3 mg and a 100 mg Fe (as FeSO4) dose. The study was conducted in a group of eleven male blood donors, without any evident infection, who had given blood 8 weeks before the study. On three consecutive mornings the subjects were served a wheat roll fortified with Fe. On the first 2 d the roll was fortified with 3 mg Fe labelled with 59Fe; on day 3 the roll was fortified with 100 mg Fe labelled with 55Fe. The serum Fe response to the 100 mg dose was followed for 6h. Fe absorption was measured by whole-body counting. High correlations were seen between the absorption of Fe and the change in serum Fe after 100 mg Fe ($r^2 0·94, P, 0·001$), between the absorption from 3 mg and 100 mg Fe ($r^2 0·88, P, 0·001$), and between the absorption from 3 mg Fe and change in serum Fe after 100 mg Fe ($r^2 0·90, P, 0·001$). This strengthens the evidence that it is possible to use the change in serum Fe as a measure of Fe absorption, e.g. when establishing the relative bioavailability for Fe powders. The results also imply that the induced serum Fe increase following 100 mg Fe added to a food could predict the Fe absorption of a small dose of Fe added to the same meal.

Serum iron: Iron absorption: Relative bioavailability: Elemental iron

Several methods can be used to measure Fe absorption from Fe compounds. Chemical Fe balance requires long balance periods and great care in analysing oral intake and faecal output of Fe. Thus, it is not feasible to use this method in practice. The introduction of radioisotopes has facilitated the measurement of Fe absorption especially when combined with whole-body counting to measure the total amount of retained radiolabelled Fe. A condition for such measurements is that the Fe compounds in the meal should have a bioavailability equivalent to the extrinsic radiolabelled Fe tracer and therefore that the meal can be homogeneously labelled, thus reflecting the true absorption. This homogeneously labelling ability is valid for most foods and Fe salts. Elemental Fe powders (e.g. metallic Fe), which are insoluble in water and thus have unknown levels of bioavailability, are impossible to label homogeneously with an extrinsic radiolabelled Fe tracer.

Another approach in evaluating the bioavailability of different Fe compounds is to monitor the change in serum Fe after a standardised meal. Ekenved and colleagues (1976) found that when using radiolabelled Fe salts it was possible to relate the induced serum Fe increase to the true absorption of Fe measured by whole-body counting (Ekenved, 1976; Ekenved et al. 1976b). It was found that in a group of subjects there was a good agreement between the amount of absorbed Fe and the serum Fe increase.

However, since the amount of Fe present in common meals will give a low response in serum Fe, higher Fe doses must be used. This begs question of the usefulness of this method when predicting the outcome in an actual fortification situation when low Fe doses are used. In the studies by Ekenved and colleagues FeSO4 was administered as solutions and tablets, which may differ from a meal-based situation. The first aim of the present study was to investigate the correlation between the change in serum Fe and the absolute Fe absorption measured with radioisotopes following administration of 100 mg Fe as FeSO4 in a meal. A second aim was to study the amount of Fe absorbed from a physiological dose of Fe (3 mg) in relation to the absorption from a pharmacological dose of Fe (100 mg). Finally we also wanted to examine the absorption from a physiological dose of Fe (3 mg) and to compare it with the serum Fe increase from a pharmacological dose of Fe (100 mg).

Material and methods

Study design

On three consecutive mornings eleven male subjects were served a wheat roll fortified with FeSO4. On the first 2 d the roll was fortified with 3 mg Fe labelled with 59Fe, whereas on day 3 the roll was fortified with 100 mg Fe labelled with 55Fe. The Fe absorption from both these doses as well...
as the serum Fe response for 6 h from the 100 mg dose was studied. Variability in Fe status between subjects was controlled by using male subjects, without any evident infection, who had given blood 8 weeks before the study.

Subjects and procedures

Eleven healthy male subjects (aged 29–59 years), who had been regular blood donors for many years, took part in the present study. The subjects had given 450 ml blood at a blood donor centre 8 weeks before attending the study. Before the study the subjects denied the use of medicine or Fe medication. All volunteers had been informed about the aim and procedures of the study, both verbally and in writing, before giving written consent to participate. The Ethics Committee of the Medical Faculty of Göteborg University approved the study. The subjects came to the laboratory on three consecutive mornings between 08:00 and 09:00 hours after fasting overnight. The subjects were not allowed to eat anything after 22.00 hours or drink anything after 24.00 hours the evening before. Each morning the subjects were served a meal consisting of one wheat roll fortified with Fe and 150 ml water. The rolls, which were kept frozen until used, were baked from 40 g low-extraction wheat flour, 28:0 g water, 2.6 g yeast, 1.3-g sugar and 0.4 g NaCl at our laboratory. The native Fe content in each roll before fortification was 0.15 mg. The weighed amounts of Fe were added through an incision in the roll.

On days 1 and 2 the subjects were served the wheat roll fortified with 3 mg Fe (as FeSO₄) labelled with ⁵⁹Fe. In order to reduce the influence from the day-to-day variation in absorption the mean absorption from these 2 d was later calculated. The subjects were then instructed not to eat or drink for the next 3 h.

On day 3 an oral Fe tolerance test was performed and an absorption test using radioisotopes. The subjects were served the wheat roll fortified with 100 mg Fe (as FeSO₄) labelled with ⁵⁵Fe. Before serving the meal, a plastic catheter was inserted into an anticubital vein and initial blood samples were obtained without minimising the effect of haemolysis on the serum Fe analysis, serum ferritin and total Fe-binding capacity. In order to compare the effect of haemolysis on the serum Fe analysis, the serum Fe blood samples were obtained without using a vaccutainer and the blood was thus allowed to run freely into the test-tubes (Venoject®II, Autosep, Gel + Clot Act. Z; Terumo Europe NV, Leuven, Belgium). The handling of the blood samples and the serum Fe analysis was conducted as previously described (Hoppe et al. 2003). In order to follow the effect of oral Fe on serum Fe concentration, blood samples were drawn hourly for the first 6 h following the administration of the roll. Following the fifth blood sample, i.e. after 4 h, two unsweetened rusks were served with 150 ml coffee or tea. Between the blood sampling the subjects remained in our laboratory resting in a seated position. Whole-body counting was performed 10–14 d later and a blood sample was drawn in order to calculate the whole-body retention of both isotopes as previously described (Rossander, 1987).

The total received radioactivity for each subject was 74 kBq from ⁵⁵Fe and 37 kBq from ⁵⁹Fe.

Statistical analyses

After determining the increase in serum Fe following the meal, the area under the curve for the 6 h time period (AUC₀–₆h) was calculated using the trapezoidal rule. In a previous study we examined the increase in the basal serum Fe concentration in thirty-two subjects under the same conditions as this present study (Hoppe et al. 2003). It showed that the basal diurnal variation (AUC₀–₆h 18.7 (SEM 1.7) μmol·h/l) included in the serum Fe increase achieved by an oral dose of Fe must be subtracted from the primary data for serum Fe increase. Hence the AUC₀–₆h for each individual in this present study was adjusted for diurnal variation earlier seen in blood donors during exactly the same study conditions. The method used for comparing correlation was the Pearson correlation test. All P values are two-tailed. The statistical program used was SPSS for Windows 10.0.5 (SPSS Inc., Chicago, IL, USA).

Results

Exclusions

From the present results three of the subject’s data were excluded from further calculation. One subject was excluded because of the influence of infection. The other two subjects exceeded the total Fe-binding capacity after administration of FeSO₄; this excluded their data from the calculation of the serum Fe change, but not of absorption. When the total Fe-binding capacity is exceeded, a part of the absorbed Fe will be deposited in the liver during the first passage (Wheby & Umpierre, 1964).

The iron absorption v. change in serum iron after 100 mg iron

When 100 mg Fe as FeSO₄ was administered together with the bread roll and water, the mean serum Fe AUC₀–₆h was 201.1 (SEM 33.8) μmol·h/l and the mean absorption was 14.7 (SEM 2.0) %. The squared correlation coefficient (r²) between Fe absorption from 100 mg and the AUC₀–₆h for the serum Fe change was 0.94 (P<0.001, n 8) (Fig. 1).

The iron absorption from 3 mg iron v. absorption from 100 mg iron

The mean absorption from 3 mg Fe was 55.4 %. When comparing this absorption with the mean absorption from 100 mg Fe, r² was 0.88 (P<0.001, n 10) (Fig. 2).

The iron absorption from 3 mg iron v. serum iron change from 100 mg iron

The r² between Fe absorption from 3 mg and the serum Fe AUC₀–₆h after administering 100 mg Fe was 0.90 (P<0.001, n 8).
The haematological variables and Fe absorption results are presented in Table 1.

**Table 1.** Initial haematological variables and iron absorption from 100 mg iron and 3 mg iron as FeSO₄*

<table>
<thead>
<tr>
<th>Subject no.</th>
<th>100 mg Fe</th>
<th>3 mg Fe</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serum-Fe (µmol/l)†</td>
<td>TIBC (µmol/l)</td>
<td>Ferritin (µg/l)</td>
</tr>
<tr>
<td>Time (h)…</td>
<td>0</td>
<td>1</td>
</tr>
<tr>
<td>1</td>
<td>0.0</td>
<td>39.2</td>
</tr>
<tr>
<td>2‡</td>
<td>0.0</td>
<td>26.2</td>
</tr>
<tr>
<td>3</td>
<td>0.0</td>
<td>30.4</td>
</tr>
<tr>
<td>4</td>
<td>0.0</td>
<td>9.3</td>
</tr>
<tr>
<td>5</td>
<td>0.0</td>
<td>25.7</td>
</tr>
<tr>
<td>6</td>
<td>0.0</td>
<td>5.5</td>
</tr>
<tr>
<td>7‡</td>
<td>0.0</td>
<td>41.6</td>
</tr>
<tr>
<td>8</td>
<td>0.0</td>
<td>33.7</td>
</tr>
<tr>
<td>9</td>
<td>0.0</td>
<td>38.8</td>
</tr>
<tr>
<td>10</td>
<td>0.0</td>
<td>33.2</td>
</tr>
<tr>
<td>11§</td>
<td>0.0</td>
<td>9.3</td>
</tr>
<tr>
<td>Mean</td>
<td>0.0</td>
<td>26.99</td>
</tr>
<tr>
<td>SD</td>
<td>0.0</td>
<td>12.87</td>
</tr>
<tr>
<td>SEM</td>
<td>0.0</td>
<td>3.88</td>
</tr>
</tbody>
</table>

TIBC, total Fe-binding capacity; AUC₀₋₆ₘ, area under the curve for 0–6 h.
* For details of subjects and procedures, see p. 486.
† Increase in serum Fe concentration for 6 h following administration of 100 mg Fe; values were corrected for diurnal variation in serum Fe.
‡ Subject was excluded from calculation of mean serum Fe AUC due to exceeded TIBC.
§ Subject was excluded from calculation of mean serum Fe AUC and absorption from 100 mg Fe due to infection.

**Discussion**

The present findings strengthen our earlier conclusion that monitoring serum Fe increase may be a reliable and simple method to determine the relative bioavailability of various elemental Fe powders in relation to FeSO₄ (Hoppe et al. 2003). However, in order to induce a serum Fe response that differs significantly from the basal variation, a pharmacological dose of Fe is needed. This could give rise to arguments against using this method to determine relative bioavailability, since the response from 100 mg Fe perhaps cannot be translated into an actual fortification situation when smaller physiological doses are used. So, is the relative bioavailability for a fortification Fe powder obtained with this pharmacological dose of Fe really the same when a smaller dose is used? To investigate this we conducted an experiment where the absorption from a 3 mg and a 100 mg Fe dose, as well as the serum Fe response from the 100 mg dose, were studied. The correlations between the absorption and the change in serum Fe from 100 mg, the absorption from 3 mg Fe and the 100 mg Fe dose, and also between the absorption from 3 mg Fe and the change in serum Fe after 100 mg, were good.

When using the double-isotope technique and whole-body counting, a relative bioavailability based on absorption from 100 mg Fe is as valid as a relative bioavailability based on absorption from 3 mg Fe. The present results show that the relative bioavailability based on serum Fe AUC₀₋₆ₘ when administering 100 mg Fe also is valid, since AUC₀₋₆ₘ correlates with absorption from 100 mg Fe (Fig. 1). Furthermore, the absorption from 100 mg Fe, as well as the AUC₀₋₆ₘ, correlates with absorption from 3 mg Fe (Fig. 2 and p. 486). These results make it highly probable that a relative bioavailability...
obtained with the serum Fe method, using 100 mg Fe, is equivalently valid in a fortification situation using lower levels of Fe, e.g. 3 mg.

Subject no. 7 was an exception from the strong correlation between the absorption from the two Fe doses. This subject had the highest absorption from the 3 mg dose (82.4%). Nevertheless in order to end up on the correlation line, the absorption had to be even greater. One possible explanation for this is that when approaching 100% absorption the dose–response relationship deviates. The reason for this could be that when absorption proceeds the remaining Fe in the lumen decreases. This in turn makes it more and more difficult for the brush border Fe transporters (e.g. the divalent metal transporter 1) on the mucosa cells to detect, attract and transport the Fe atoms inside the enterocytes.

The Fe tolerance test, i.e. the serum Fe method, has earlier mostly been used to study Fe absorption from Fe preparations for pharmaceutical purposes and for diagnosis of Fe deficiency (Ekenved, 1976a; Ekenved et al. 1976a; Nielsen et al. 1976; Kelsey et al. 1991). When the purpose has been to evaluate the usefulness of this method in discriminating between normal and Fe-deficient individuals, the small-dose Fe tolerance test (using Fe doses of 5–20 mg) is the most commonly used (Crosby & O’Neil-Cutting, 1984; Costa et al. 1991; Joosten et al. 1997; Jensen et al. 1998a, 1999). The small-dose Fe tolerance test is based on the fact that low doses of Fe do not have the potential to induce any changes in serum Fe in subjects with normal Fe status. In order to do this, the subjects must have some degree of Fe deficiency and hence a higher Fe absorption ability. The utilisation of the serum Fe method on determining the relative bioavailability of elemental Fe used for fortification has to our knowledge only been done in human subjects once before (Gonzalez et al. 2001). However, in our hands the serum Fe method has recently, and for the first time, been used in a meal situation to determine the relative bioavailability of elemental Fe powders (Hoppe et al. 2003).

In summary, the present results strengthen our earlier conclusion that monitoring changes in serum Fe as a measure of Fe absorption is possible. Furthermore, the induced increase in serum Fe following 100 mg Fe added to a food could predict the Fe absorption of a small dose of Fe added to the same meal.

References


