# Metabolism of vitamin E in sheep\*

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1. The metabolism of tritium-labelled tocopherol was investigated in thirty-six wethers and forty-two ewes fed on hay known to produce nutritional muscular dystrophy (NMD).

2. In one experiment, the effect of selenium or vitamin E or both on the metabolism of tritium-labelled tocopherol was studied. No difference was found in the incorporation rate of radioactivity in tissues due to the addition of either of these components.

3. In a second experiment, significantly more radioactivity was found in the tissues of sheep dosed intramuscularly than of those dosed by stomach tube into the rumen. Excretion of radioactivity was rapid when labelled tocopherol was given into the rumen.

4. In a third experiment, the efficiency of vitamin E in preventing NMD in lambs was tested. Supplementation with vitamin E of the diet consisting of NMD-producing hay and given to ewes was ineffective as a prophylactic treatment. Direct administration of vitamin E to the lambs completely prevented the occurrence of NMD. Plasma vitamin E concentrations in the treated lambs were significantly higher than in untreated lambs at weekly sampling over a 4-month period regardless of whether the lambs were born from ewes treated with the vitamin or not.

The metabolic interrelationships of selenium and vitamin E have received considerable attention during the last few years. Calvert, Nesheim & Scott (1962) have suggested a synergistic effect between vitamin E and selenium in the prevention of muscular degeneration in the chick. Desai & Scott (1965) reported that Se is effective in increasing the retention of radioactive tocopherol in the plasma of chicks. However, Cheeke & Oldfield (1969) did not confirm these results in rats and suggested further investigation to determine if there are species differences in the effect of Se on vitamin E metabolism. Thus, the work now reported was undertaken to study the relation of Se to vitamin E retention in sheep.

The absorption and retention of vitamin E administered to sheep either by stomach tube into the rumen or by injection also appeared to merit investigation. Investigations on monogastric animals by Overman, McNeely, Todd & Wright (1954), Gloor, Weber, Wunsch & Wiss (1963) and Imbesi (1958) indicated that oral administration of tocopherol was more effective than the parenteral route in increasing the concentration of tocopherol in the plasma. In contrast, Caravaggi, Gibbons & Wright (1968) reported that the plasma tocopherol concentrations in sheep were higher after intramuscular than after oral administration, and Brüggemann & Niesar (1954) reported that there was no increase of plasma tocopherol in sheep 14 d after intramuscular injection of vitamin E.

\* Contribution no. 365 Animal Research Institute: contribution no. 119 Analytical Chemistry Research Service.

## EXPERIMENTAL

#### Analytical methods

The tissues were homogenized in 25 ml chloroform-methanol (2:1) per g of tissue and the lipids were extracted several times with a mixture of chloroform-methanol (2:1). After evaporation of the solvent, the lipids were dissolved in 10 ml of scintillation solution (60 mg 2,5-diphenyloxazole and 1 mg 1,4-di-2-(5-phenyloxazolyl) benzene in toluene) and counted in a Packard liquid scintillation counter 518. All values were converted into disintegrations/min (dpm) by internal standardization.

*Expt* 1. Twenty 4-month-old wethers were given hay from an area in northern Ontario where nutritional muscular dystrophy (NMD) is common. Because the hay was low in both energy and nitrogen, 2% urea and 6% sucrose were added to each day's feed supply by spraying in a water solution over the hay. All wethers received vitamin A (10 000 i.u.) and vitamin D (1750 i.u.) at weekly intervals. The wethers were divided for experimental purposes into four groups, each of five animals, and received the NMD-producing hay either unsupplemented, or were given weekly, by stomach tube into the rumen, a dose of 1 mg sodium selenite, or 1 g  $\alpha$ -tocopherol, or 1 mg sodium selenite plus 1 g  $\alpha$ -tocopherol. After 1 year on these treatments, all sheep, taken one per treatment at a time, were given by stomach tube into the rumen a dose of  $4.93 \times 10^7$  dpm/kg body-weight [ ${}^{3}\text{H}$ ]DL- $\alpha$ -tocopherol-(5-methyl-T) in aqueous ethanol solution. The radioactive tocopherol (430 mCi/m-mole) was purchased from Radiochemical Centre, Amersham, England. Blood samples were taken from each sheep 2, 4, 6, 10 and 24 h after administration of the radioisotope. All tissue samples were frozen and stored at  $-20^{\circ}$  until analysed.

*Expt* 2. Sixteen Shropshire wethers were fed for 9 months on a NMD-producing hay. The animals were then placed in metabolism cages a few days before dosing and allowed hay and water *ad lib*. The tritium-labelled  $\alpha$ -tocopherol, in aqueous ethanol solution, was administered once, either via stomach tube into the rumen or intramuscularly into gluteal muscle, at doses that varied between  $8.88 \times 10^8$  dpm and  $2.22 \times 10^9$  dpm from trial to trial. At each time of dosing, a pair of sheep of equal weight was dosed; one sheep intraruminally and the other intramuscularly with the same amount of radioactivity.

From the four sheep killed at 24 h, blood was collected hourly up to 12 h and then at 24 h after dosing. Urine and faeces, if any, were collected at the same time as blood. In the other trials, blood, faeces and urine were collected at daily intervals. The collected material was handled for counting as in the previous experiment.

*Expt* 3. Forty-two ewes fed on a NMD-producing hay for 3 years were used for this experiment. At 2 months of gestation, twenty-three of them (group B) were given, by stomach tube into the rumen, at weekly intervals a dose of 1 g of DL- $\alpha$ -tocopherol; the remaining nineteen ewes (group A) were given no treatment. Four ewes in group B and five in group A produced twin lambs. From birth to weaning, eleven lambs from each of groups B and A were treated each week orally with 500 mg of DL- $\alpha$ -tocopherol, while the remaining sixteen of group B and thirteen from group A were untreated.

Serum vitamin E concentrations were determined weekly by the method of Desai (1968) and serum aspartate aminotransferase (L-aspartate: 2-oxoglutarate aminotransferase; EC 2.6.1.1) activities were determined every 2 weeks by the method of Reitman & Frankel (1957).

The carcasses of all lambs, whether the animals died or were killed, were examined for gross and microscopic lesions of NMD.

#### **RESULTS AND DISCUSSION**

Expt 1. The results showed a wide variation in the distribution of radioactivity in the tissues, even between animals on the same treatment. Since the standard deviation was largely proportional to the mean, it was necessary to use the logarithms of the values for the analysis of variance. After transformation, the standard deviations were more nearly equal and uncorrelated with the means. The level of statistical significance selected was P = 0.05.

# Table 1. Expt 1. Radioactivity of the tissues of the wethers 26 h after administration of a single oral dose of $[^3H]_{DL-\alpha}$ -tocopherol

(In all experimental treatments, NMD-producing hay was given to four groups of five animals (weights 52-63 kg) for 1 year and was either unsupplemented (control), or supplemented weekly with an oral dose of selenium, or  $DL-\alpha$ -tocopherol, or  $Se+DL-\alpha$ -tocopherol. The amount of radioactivity given to each sheep was  $4.93 \times 10^7$  dpm/kg body-weight. Values are mean logarithms of dpm/g fresh tissue with standard errors)

Tissue	No supplement	Se supplement	DL-a-Toco- pherol supplement	DL-a-Toco- pherol + Se supplement	SE
Liver	3.8270	3.6226	3.8267	3.9523	0.2162
Spleen	3.2996	3.2037	3.7173	3.3316	0.2249
Vena cava	3.2878	3.0794	3.3147	3.2690	0.1282
Abomasum	3.5446	3.6107	3.4917	3.2037	0.3765
Duodenum	3.2723	3.1046	3.3537	3.3712	0.2960
Adrenal	3.9858	3.7368	4.0076	3.8830	0.2042
Heart	3.0120	3.1879	2.9467	3.1852	0.1940
Lung	3.4830	3.4641	3.4209	3.3952	0.1800
Kidney	3.1678	3.1392	3.4856	3.3699	0.1303
Muscle	2.9865	2.7677	2.2174	2.7719	<b>0</b> ·1749

NMD, nutritional muscular dystrophy; dpm, disintegrations/min.

The presence or absence of Se or tocopherol in the diet of the sheep during the 12 months before administration of radioactive tocopherol had no significant effect on the distribution of radioactivity in any of the tissues examined (Table 1). These results are in agreement with the findings of Diplock, Bunyan, McHale & Green (1967) and Green, Diplock, Bunyan, Muthy & McHale (1967) that Se does not affect the metabolism of tocopherol in rat tissues.

According to Cheeke & Oldfield (1969), plasma radioactivity after administration of tritiated vitamin E was lower in rats given a Se supplement than in untreated animals. Scott & Thompson (1968) found that Se increased the absorption and raised blood concentrations of vitamin E in chicks. On the other hand, Mukhtar (1966) reported that Se supplementation of diets for steers caused an increase in vitamin E in blood

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# Table 2. Expt 1. Radioactivity of the blood of wethers after administration of a single dose of $[^{3}H]$ DL- $\alpha$ -tocopherol

(In all experimental treatments, NMD-producing hay was given to four groups of five sheep (weights 52-63 kg) for 1 year and was either unsupplemented (control), or supplemented weekly with an oral dose of selenium, or  $DL-\alpha$ -tocopherol, or  $Se+DL-\alpha$ -tocopherol. The amount of radioactivity given to each sheep was  $4.93 \times 10^7$  dpm/kg body-weight. Values are mean logarithms of dpm/g fresh blood with standard errors)

Sample time- interval (h)	No supplement	Se supplement	DL-a-Toco- pherol supplement	DL-a-Toco- pherol + Se supplement	SE
I	2.4799	2.8949	2.9194	2.8387	0.1142
2	2.8870	3.0129	3.1120	3.1496	0.1428
4	<b>2·</b> 8986	3.2254	3.3348	3.1677	0.1103
6	2.8583	3.0023	2.9966	3.1343	0.1337
12	3.1092	3.3022	3.3917	3.2607	0.0994
24	3.0987	3.3472	3.3336	3.3847	0.1006

NMD, nutritional muscular dystrophy; dpm, disintegrations/min.

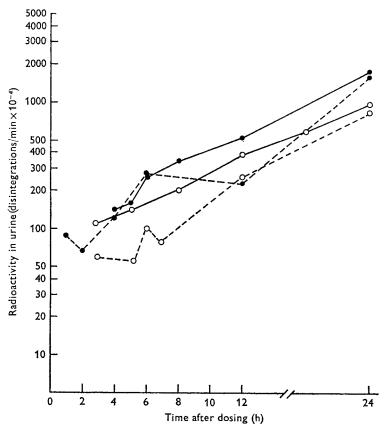


Fig. 1. Urine clearance of radioactivity after administration to sheep, intramuscularly or by stomach tube into the rumen, of a single dose of [<sup>3</sup>H]DL-α-tocopherol. •••, sheep 27  $(8.88 \times 10^8 \text{ dpm}, \text{ by stomach tube}); \bullet -- \bullet, \text{ sheep 2 } (8.88 \times 10^8 \text{ dpm}, \text{ intramuscularly}); O-O \text{ sheep 48 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ by stomach tube}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 73 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 74 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 74 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 74 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 74 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 74 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 74 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 74 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 74 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 74 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 74 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 74 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 74 } (1.11 \times 10^9 \text{ dpm}); O-O, \text{ sheep 74 } (1.11 \times 10^9$ intramuscularly). dpm, disintegrations/min.

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and a decrease in the liver. Mukhtar (1966) suggested that the reduction of vitamin E in the liver may be due to its transfer from liver to blood caused by Se functioning as a tocopherol carrier.

In our experiment, the presence or absence of Se or vitamin E, or of both, in the diet of sheep before administration of [<sup>3</sup>H]tocopherol had no significant effect on the radioactivity recovered in blood at any of the sampling times (Table 2).

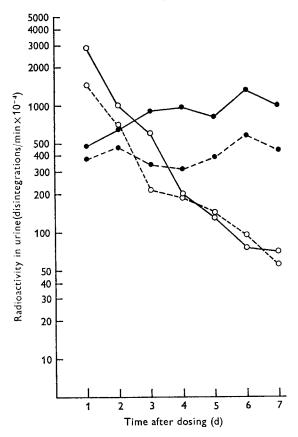


Fig. 2. Urine clearance of radioactivity after administration to sheep, intramuscularly or by stomach tube into the rumen, of a single dose of  $1.11 \times 10^9$  dpm [<sup>3</sup>H]DL- $\alpha$ -tocopherol. -, sheep 44 (intramuscularly); - - -, sheep 26 (intramuscularly); --0, sheep 56 (by stomach tube); 0 - - 0, sheep 37 (by stomach tube). dpm, disintegrations/min.

From blood measurements taken immediately before slaughter (24 h after dosing) and from liver measurements at slaughter, regression analyses were calculated, using logarithmic transformations, to evaluate the possible relationships between these variables. The linear regression of dpm in blood on dpm in the liver (r = -0.407) was not significant. In order to study the possibility that the relationship might be curvilinear, the analyses were again made after a second logarithmic transformation of the logarithms of dpm in the liver. The fit of the curve was not improved (r = -0.413).

*Expt 2.* Maximum radioactivity in blood was recorded after 6 and 8 h for the intramuscular and intraruminal treatments respectively, followed by a slight decrease and

Radioactivity of fresh blood, and of the daily faeces and urine from wethers nos.* 70a, 55a, 83b and 88b	whe into the rumen (IR) or intramuscularly (IM), a single dose of $1.11 \times 10^9 dpm$ [ <sup>3</sup> H]DL- $\alpha$ -tocopherol after	hay for 9 months
Table 3. Expt 2. Radioactivity of fresh blood, and of the	given, by stomach tube into the rumen (IR) or intramusci	having received NMD-producing hay for 9 months

le	Total dɛ	aily faecal e	Fotal daily faecal excretion (dpm $\times$ 10 <sup>-4</sup> )	1 × 10 <sup>-4</sup> )	Total dail	y urinary e	xcretion (6	otal daily urinary excretion $(dpm \times 10^{-4})$		Blood activity $(dpm \times 10^{-2}/g)$	(dpm×1c	o <sup>−2</sup> /g)
losing (d)	70a IR	55a IM	$^{8_3b}_{IR}$	88b IMI	70a IR	55a IM	83b IR	88b IMI	70a IR	55a IM	83b IR	88b IM
	64376	2522	59732	3800	1 387	2416	1 792	1285	84	107	38	421
	26572	4142	23376	6738	1314	2819	507	1 509	65	381	33	418
	5 688	3034	4495	5 204	161	2 949	204	1188	42	132	14	358
	1188	2407	I 394	3450	822	1827	°,	824	60	234	20	239
	351	2 0 6 2	562	3073	133	1 030	75	622	24	268	II	245
	186	1518		2 209	62	854	45	562	23	192	17	189
	151	1 440	441	1729	67	969	13	430	71	207	27	184
	61	1 229	330	I 420	42	570	20	333	17	169	IO	161
	38	1157	250	1 566	47	478	19	256	II	171	9	158
	59	o26	120	1 560	38	447	28	202		165	10	138
	53	843	51	1149	35	853	19	295	13	157		711
	44	851	49	1 309	34	582	18	293	12	175	II	6 <b>0</b> 1
	30	1021	21	1 223	33	515	22	308	10	160	7	122
	II	522	×	653	26	774	15	240		102	0	III

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NMD, nutritional muscular dystrophy; dpm, disintegrations/min. \* Sheep whose numbers are followed by the same letter were dosed at the same time: (a) 34–36 kg body-weight; (b) 37–38 kg body-weight.

		Tissue ra	Tissue radioactivity		Sample		Faeces radioactivity	ioactivity	
Tissue	27 IR	48 IM	48 IR	13 IM	ume interval (h)	27 IR	4 MI	84 RI	13 IM
Liver	21044	36517	35 921	61 687	, v	195	30	03	II
Adrenal	10 594	21561	3555	20385	<i>م</i> '	251	68	;	32
Spleen	4750	12811	4411		7	742	ļ	504	51
Lung	3353	9265	4091	7448	×	2 0 9 9	195	1 710	38
Vena cava	795	4356		1 020	12	6226	]	14986	, S
Kidney	2 061	3 106	4 100	12342	24	58485	645	22 658	1541
Pancreas	898	2 003	1 066	[	Total			3	
Heart	196	2310	1 520	2446	in 24 h	o7 998	948	39951	1 703
Muscle	366	2 001	2186	2395					
Abomasum	859	1 623	1614	2059					
Duodenum	3671	1333	1 220	4 I 86					
Fat	742	2 069	450	1 578					
Testis	000	1434	1681	1072					

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Table 4. Expt 2. Radioactivity of fresh tissues (dpm) and faeces (dpm  $\times$  10<sup>-4</sup>) from wethers nos. 27, 2, 48 and 72, 24 h after a

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then an increase at 24 h. At these times and thereafter, blood radioactivity concentrations were much higher in sheep dosed intramuscularly than in those dosed intraruminally.

Intraruminal and intramuscular doses of [<sup>3</sup>H]tocopherol were both rapidly excreted in the urine of sheep (Fig. 1). In the sheep dosed intraruminally, the urinary radioactivity values were highest at 24 h and thereafter declined sharply (Fig. 2). When sheep were injected intramuscularly, urinary radioactivity increased to a maximum in 3–6 d and subsequently declined steadily (Fig. 2, Table 3). These results show that in the sheep dosed intramuscularly the radioactivity of both blood and urine decreased much more slowly than in those dosed intraruminally (Table 3). This difference could be related to the slow release of [<sup>3</sup>H]tocopherol from the site of injection into the circulatory system, if tocopherol introduced direct into the gastro-intestinal tract is eliminated rapidly. Radioactivity concentrations in the faeces indicate that sheep receiving tritium-labelled tocopherol intraruminally excreted the greater percentage of the dose during the first 3 d; in the intramuscularly injected sheep the excretion rate was much slower (Table 3).

There were significant differences (at the 1% level) in the metabolism of the tritiated vitamin E given by either method. One or more days after treatment, retention of radioactivity in the tissues was much greater in the sheep treated intramuscularly than in those dosed intraruminally (Tables 4 and 5). There was no substantial difference in the pattern of tissue distribution due to the mode of administration.

As reported previously (Hidiroglou, Jenkins & Carson, 1969), high uptakes of radioactivity were recorded at 24 h in some tissues, such as liver and adrenal, and lower uptakes in others, such as muscle (Table 5). However, at 2 weeks, these differences were less. Adipose tissue, testis and heart accumulated large amounts of radioactivity and they could be considered among the accumulator tissues. In these tissues, localization of tritiated vitamin E might be related to the physiological role of vitamin E. Rindi & Perri (1958) reported a significant increase in plasma tocopherol concentration in human beings after the parenteral administration of an aqueous solution of vitamin E. According to these authors, the ineffectiveness of intramuscular dosing reported by other investigators was due to the very slow absorption of the oil carrier which was used.

Our results are in agreement also with those of Johnson (1955) who reported that, following oral or intramuscular administration of [14C]tocopherol, the percentage of the administered radioactivity recovered in the carcasses of rats 48 h after dosing was higher when the dose was given intramuscularly.

Differences in the ability of individual sheep to metabolize tocopherol, as reported in Expt 1, were found also in this experiment, presumably because there was variation in the rate of excretion between different sheep. The rate of excretion differed markedly with the two routes of administration of the radioactive tocopherol; vitamin E given orally in aqueous solution was excreted rapidly, which could explain the much lower residual radioactivity in the tissues of intraruminally treated than of intramuscularly treated sheep.

Table 5. Expt 2. Effect of mode of administration, by stomach tube into the rumen (IR) or intramuscularly (IM) of  $[^{3}H]$ DL- $\alpha$ -tocopherol on the distribution of radioactivity in the tissue of wethers after having received NMD-producing hay for 9 months

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141. 1	IDI	RUU	TL	<i>,</i> 0	А	.11	υ	0	11-	1E.	ĸS					
th 500 mg ambs) and I B₀)		SE	0.20350	0.25754	0.35656	0.29852	0.32305	0.27749	0.33175	0.34416	0.23586	0.20247	o.11541			ular dystrophy.
tted weekly wi group B (B <sub>0</sub> 1 d) amin E (A <sub>0</sub> and		Healthy	(2) 8866.1	(8) 61 <b>7</b> 10	o.1085 (8)	0.0043 (8)	$\overline{1}.9914$ (8)	<u>1</u> .8281 (8)	<u>1</u> -6013 (8)	ī-6812 (8)	<u> </u>	(8) 1007.I	ī.7371 (7)			NMD, nutritional muscular dystrophy.
eleven from group A ( $A_1$ lambs) were treated weekly with 500 mg vitamin E; the remaining sixteen lambs of group B ( $B_0$ lambs) and thirteen of group A ( $A_0$ lambs) were untreated) Lambs not given vitamin E ( $A_0$ and $B_0$ )	With	Very sugar NMD lesions	0.0705 (10)	0.0462 (8)	(6) 996o.o	(6) 0960.0	o·1064 (9)	0.0955 (8)	$\overline{1.9478}(7)$	$\overline{1}.7492(5)$	$\overline{1.6353}$ (5)	ī-6161 (5)	ī·74o3 (4)			NMD, n
group A (A <sub>1</sub> <sup>1</sup> the remaining s roup A (A <sub>0</sub> lamh Lar		WILLI SEVERE NMD lesions	0.1060 (12)	<u>1</u> .9415 (10)	1.8403 (10)	ī-8298 (10)	$\overline{1}.6458(9)$	1.6729(8)	[1.5109 (7)	$\overline{1}.2858(5)$	$\overline{1}.3228(5)$	ī.6440 (4)	<u>1</u> .4717 (4)			mbers of lambs.
eleven fron vitamin E; thirteen of		sD*	0.21330	0.26168	0.30942	00962.0	0.33428	01002.0	0.31472	0.32506	0.24166	0.18232	0.17055	0.21723	0.22065	eses are the nui
1p B) were dosed 1, with I g DL-α- eceived no treat- B (B <sub>1</sub> lambs) and		B1	0.8645 (11)	(11) 1227.0	0.6556 (11)	0.4995 (11)	0.3941 (11)	0.3322 (9)	0.4559 (8)	0.3011 (8)	0.1977 (8)	0.1128 (8)	<u>1</u> .9715 (8)	ī-9066 (8)	ī.9774 (8)	Figures in parentheses are the numbers of lambs.
(At 2 months of pregnancy twenty-three ewes (group B) were dosed at weekly intervals, by stomach tube into the rumen, with 1 g DL- $\alpha$ - tocopherol; the remaining nineteen ewes (group A) received no treat- ment. From birth to weaning eleven lambs from group B (B <sub>1</sub> lambs) and	All lambs	A1	(11) 5120.1	(11) 4066.0	0.6841 (II)	0.7747 (11)	(11) E0170	0.4982 (11)	0.3718 (10)	0.4667 (10)	0.2569 (9)	(6) 190E.o	0.1220 (6)	ī.9527 (6)	ī 8894 (6)	atments.
		B	0.1316 (16)	0.0800 (14)	0.0497 (15)	I.9880 (15)	1.9575 (15)	ī·8641 (13)	ī.7268 (11)	I.6684 (7)	ī.5934 (7)	$\overline{1.6459}(7)$	(9) 01 19 <u>1</u>	ī·6075 (4)	1.6981 (4)	* Pooled standard deviations within treatments.
et 2 months of eekly intervals pherol; the ren t. From birth to		$\mathbf{A_0}$	0.0047 (13)	0.0033 (12)	$\overline{1}.9364 (12)$	0.0084 (12)	I-8492 (11)	ī·8672 (11)	<u>1</u> .6388 (11)	ī-5419 (11)	1.5550 (11)	1.6729 (IO)	1.7046 (9)	1.5714 (8)	ī-5153 (8)	led standard de
(/ at w toco men	Δ	(weeks)	I	61	ς	4	ŝ	9	7	œ	6	OI	II	12	13	* Poo

Table 6. Expt 3. Mean logarithms of the plasma DL- $\alpha$ -tocopherol concentrations ( $\mu g/ml$ ) of lambs from eves that had

received NMD-producing hay for 3 years

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# Metabolism of vitamin E in sheep

Expt 3. DL-a-Tocopherol given direct to lambs protected them completely from death due to NMD and elevation of serum aspartate aminotransferase. No such protection, however, was obtained by the untreated lambs born from ewes given 1 g  $DL-\alpha$ -tocopherol weekly, six of which died from NMD. In this experiment, no vitamin E determinations were made on the milk of the ewes; however, an appreciable increase of tocopherol concentrations in the milk of cows following supplementation with vitamin E has been reported by Parrish (1949) and Merk & Crasemann (1961). On the other hand, Safford, Swingle & McRoberts (1956) found no correlation between NMD incidence and tocopherol concentrations in ewe's milk. The fact that NMD occurs mostly in the spring, when the young grass is rich in vitamin E, suggests that the tocopherol provided by ewe's milk may be biologically unavailable to their lambs. This is supported by the present results, in which plasma vitamin E concentrations in the unsupplemented lambs from either treated or untreated ewes were of the same magnitude (Table 6). It is possible that either the availability to the lambs of vitamin E from ewe's milk was impaired by the presence of an antagonistic substance or its retention was greatly diminished by the absence of some dietary component not present in the milk.

The concentrations of vitamin E in the plasma were much higher in lambs dosed with vitamin E than in those untreated (Table 6). Although this difference was initially large, it diminished thereafter, in spite of the fact that the treated lambs continued to receive 500 mg vitamin E weekly. Such decrease of plasma tocopherol concentrations with age could be related either to the diminishing capacity of a component of the plasma or to some defect in the absorption by the lambs of this lipid-soluble vitamin. The variability in the vitamin E concentrations of the plasma was great and was more pronounced in the vitamin E-treated lambs.

As reported previously, none of the vitamin E-treated lambs died from NMD or showed SGOT values of more than 200 Frankel Units. The percentages of untreated lambs with elevated serum aspartate aminotransferase values (> 200) at 1, 3, 5, 7 and 9 weeks of age were 33, 66, 50, 45 and 40 for the lambs born from the untreated ewes, and 15, 36, 37, 33 and 33 for those born from vitamin E-supplemented ewes.

According to Culik, Bacigalupo, Thorp, Luecke & Nelson (1951), NMD in lambs is characterized by low tocopherol concentrations in blood, with critical values below 0.081 mg of total tocopherol/100 ml of blood. In our experiment, death from NMD was associated with such low plasma tocopherol concentrations, and similarly low tocopherol concentrations were recorded in the plasma of untreated lambs that were healthy (Table 6) but had high serum aspartate aminotransferase values.

All the results obtained in this experiment, including those for serum tocopherol concentrations, clinical and post-mortem examinations and serum aspartate amino-transferase determinations, confirm the findings of Hogue, Proctor, Warner & Loosli (1962) that  $\alpha$ -tocopherol is utilized for the prevention of NMD when given direct to the lambs but not when given to the ewes.

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