Reversal of Acute Experimental Cerebral Vasospasm by Calcium Antagonism with Verapamil


SUMMARY: Acute vasospasm of the transclivally exposed basilar artery of anesthetised cats was produced by the subarachnoid injection of platelet-rich plasma (PRP) treated with enough adenosine diphosphate (ADP) to induce platelet aggregation and secretion. Vasorelaxation was produced by the topical application of the calcium antagonist verapamil. Changes in the internal diameter of the basilar artery were determined by measuring the blood column diameter from photomicrographs taken sequentially, at 5 minute intervals, through the operating microscope. Changes in blood vessel diameter are expressed as a plus or minus percentage of the pretreatment diameter. Arterial blood pressure and blood gas values were kept in the physiological range for the cat.

The subarachnoid injection of PRP-ADP produced severe constriction of the basilar artery (mean constriction at 5 minutes after injection: $-40.7\% \pm 2.8$ SEM). Platelet-free plasma, ADP alone and Elliott's A solution had no spasmodogenic effect when injected into the subarachnoid space. The topical application of the calcium channel blocker verapamil (0.1 mg per kg) 30 minutes after the injection of PRP-ADP, with the basilar artery still in spasm (mean constriction: $-23\% \pm 3.5$ SEM), produced prompt and dramatic vasodilation (mean dilation at 5 minutes after application: $+52.7\% \pm 18.1$ SEM). This spasmylytic effect persisted in a decremental fashion for the 60 minute period of observation, by which time the previously constricted vessel had returned to its normal size. These observations indicate that the platelet fraction of whole blood may be involved in the genesis of acute vasospasm following subarachnoid hemorrhage and that this phenomenon can be readily reversed by calcium antagonism.

RESUME: Nous avons produit, grâce à l’injection sous-arachnoïdienne de plasma riche en plaquettes (PRP), traité avec adénosine diphosphate (ADP) en concentration suffisante pour induire l’agrégation et la sécrétion des plaquettes, un vasospasme aigu au niveau de l’artère basilaire exposée chez le chat anesthésié. Une vasoélastique a été produite par l’application topique d’un antagoniste du calcium, le verapamil. Pour mesurer les modifications du diamètre interne de l’artère basilaire, on s’est servi de photomicrographies séquentielles (au 5 minutes), prises au microscope opératoire, qui permettaient d’évaluer le diamètre de la colonne hématoïlogique. Les modifications dans le diamètre du vaisseau sont indiquées en pourcentage ± du diamètre pré-traitement. La pression artérielle et les gaz sanguins furent maintenus dans des valeurs normales pour le chat.

L’injection sous-arachnoïdienne de PRP-ADP a produit une contraction sévère de l’artère basilaire ($-40.7\% \pm 2.8$ SEM, 5 minutes après l’injection). Le plasma libre en plaquettes, l’ADP seul, la solution d’Elliott A, n’eurent aucun effet spasmodique. L’application topique du verapamil (0.1 mg par kg) 30 minutes après l’injection de PRP-ADP, alors que l’artère basilaire était encore en spasme ($-23\% \pm 3.5$ SEM) causa une vasodilatation prompte et dramatique ($+52.7\% \pm 18$ SEM après 5 minutes). Cet effet spasmylytique persistait de façon décroissante pour 60 minutes après quoi le vaisseau contracté avait repris son diamètre initial. Ces observations montrent qu’après une hémorragie sous-arachnoïdienne, les plaquettes peuvent être impliquées dans la genèse d’un vasospasme aigu et que ce phénomène peut être renversé par un antagoniste du calcium.


Cerebral vasospasm is a major cause of mortality and morbidity in patients with aneurysmal subarachnoid hemorrhage (SAH) who survive to reach hospital. Platelet produced spasmodogens, such as serotonin, prostaglandins and thromboxanes, have been implicated as a cause of experimental and clinical cerebral vasospasm (Yamamoto et al., 1972; Latorre et al., 1974; Peterson et al., 1975; Boullin et al., 1976; Hagen et al., 1977; Zervas et al., 1979). These substances may produce vasospasm by altering the permeability of the vascular smooth muscle cell to extracellular calcium or to calcium stored in the sarcoplasmic reticulum, permitting its entry into the cell where it becomes available for actin-myosin coupling (Peterson and Leblanc, 1976). With a constant supply of spasmodogens the constriction would persist and vasospasm would ensue.

Much enthusiasm has recently been generated for the use of calcium channel blockers in the treatment of angina pectoris secondary to coronary vasospasm (Antman et al., 1976; Johnson et al., 1981; Pepine and Conti, 1981). Preliminary reports attest to the usefulness of these agents in the treatment of experimental and clinical cerebral vasospasm and stroke (Peterson et al., 1969; Allen and Bahr, 1979; Allen et al., 1983; Auer et al., 1981; Brandt et al., 1979; Edvinsson et al., 1979; Takagi et al., 1979; Brandt, 1980; Cohen and Allen, 1980; Nagai et al., 1980; Brandt et al., 1981; Gaab et al., 1982; White et al., 1982; Allen et al.,...
This study documents the reversal of acute vasospasm, produced by platelet-rich plasma (PRP) activated to aggregate and release platelet contents by treatment with ADP, by the topical application of the calcium channel blocker verapamil.

**MATERIALS AND METHODS**

Eleven male and female mongrel cats were anesthetised with intraperitoneal pentobarbital, 30-35 mg per kg, and were given atropine, 0.04 mg per kg intramuscularly. A tracheostomy was performed and the tracheostomy tube was connected to a Harvard Small Animal Ventilator which was adjusted to assure constant arterial blood gas values. Catheterization of the femoral artery and vein permitted continuous monitoring of arterial blood pressure and blood gases and the intravenous administration of pentobarbital as required to maintain anesthesia. Prior to any intravenous infusion, 4.5 cc of venous blood was withdrawn for the production of platelet-rich plasma.

The transclival exposure of the basilar artery was performed by a method previously described (Kapp et al., 1968; Echlin, 1971; Peterson et al., 1975). The dura was elevated using the operating microscope. The arachnoid membrane was left intact and kept moist by intermittent application of Elliot’s A solution (mean pH 6.507). Arterial constriction was produced by injecting 0.4 cc of PRP and ADP, prepared as described below, with a number 27 needle and a tuberculin syringe. Vasodilatation was produced by covering the arachnoid membrane with 0.4 cc of verapamil (mean pH 6.609; 0.1 mg per kg) for 5 minutes.

Initial photographs were taken 30 to 60 minutes after the elevation of the dura and immediately before treatment with PRP-ADP. Photographs were again taken at 5 minute intervals after the subarachnoid injection of PRP-ADP or its controls, for a period of 30 minutes. At that time, verapamil was applied to the arachnoid membrane and its effects documented in a similar manner for a period of up to 60 minutes, at which time the experiment was repeated. The operative field was dried using a microsuction device before each photographic exposure and was covered with Elliot’s A solution after each photograph had been taken. The photographic slides were projected at constant magnification with a 35 mm slide projector. The blood column diameter within the basilar artery was measured at 3 or 4 points along its length using a millimeter ruler. This method of quantifying changes in the inner diameter of cerebral blood vessels is widely used and has been found to be accurate and reproducible. (Kapp et al., 1968; Echlin, 1971; Peterson et al., 1973a; Peterson et al., 1975). The changes in the blood column diameter produced

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**Figure 1** — (a) Basilar artery of the cat immediately before the subarachnoid injection of PRP-ADP.
(b) The same vessel 10 minutes after the subarachnoid injection of PRP-ADP, demonstrating vasoconstriction.
(c) The same vessel, 5 minutes after the application of verapamil. There is marked vasodilatation.
by treatment with PRP-ADP and verapamil are expressed as
negative (vasoconstriction) or positive (vasodilation) percentage
changes of the initial control diameter which is taken to be the
blood column diameter at 30 to 60 minutes after the elevation of
the dura, prior to any treatment, and is expressed as 0%.
Statistical analysis was performed using the student t test for
the comparison of 2 means.
PRP was prepared by mixing 0.5 cc of a 3.8% solution of
sodium citrate with 4.5 cc of whole autogenous venous blood in
a glass tube. The mixture was allowed to sediment by force of
gravity for 2 hours (Marcinkiewicz et al., 1978). The PRP was
verified on one occasion to contain 300,000 platelets per microliter.
Just prior to the subarachnoid injection the platelets were
stimulated to aggregate by mixing 0.4 cc of the PRP with 25
micromolar of ADP in a Born aggregometer for one minute.
Visible aggregates were seen in all cases.
Platelet-free plasma (PFP) was prepared after the method of
Milton and co-workers (1978) by mixing 2.25 cc of whole venous
blood with 0.25 cc of 3.8% sodium citrate in a plastic tube and
centrifuging at 1000 RPM (110G) for 15 minutes at room
temperature. The supernatant was centrifuged at 4300 rpm for
15 minutes at room temperature. The new supernatant was
centrifuged at 30,000 rpm for 15 minutes at 4°C. The final
supernatant was incubated at 37°C until ready for injection into
the SAS, approximately 30 minutes after its preparation. The
absence of platelets was verified in all cases. Twenty-five
micromolar of ADP acted as a control for the ADP used for
platelet aggregation. 0.04 cc of Elliott’s A solution served as a
control for the volume of the platelet aggregate injected into the
SAS.

RESULTS
Platelet-free plasma and ADP, Elliot’s A solution or ADP
alone had no significant vasoconstrictive effects on the basilar
artery. Platelet-rich plasma and ADP produced prompt vaso-
constriction of the basilar artery that attained a maximum at 5
minutes after injection (mean constriction: -40.73% ± 2.84 SEM)
and persisted for the 30 minute observation period (mean
constriction: -24.98% ± 3.59 SEM). At that time, the topical
application of verapamil produced rapid and dramatic vasore-
laxation of the blood vessels (mean dilatation at 5 minutes:
+51.84% ± 15.26 SEM) which persisted in a decremental fashion
for the 60 minute observation period, at which time the mean
dilatation was 6.78% ± 7.52 SEM (Table 1, Figure 1 and Figure
2). The reinjection of PRP-ADP, however, again produced
vasoconstriction of a severity similar to that produced by
the first injection. Verapamil did not, therefore, prevent vasocon-
striction following a second injection of PRP-ADP. Reapplication
of verapamil did produce prompt and dramatic reversal of the
vasoconstriction produced by the second injection of PRP-
ADP, in a manner similar to the first application of verapamil
(Table 2, Figure 3). The vasodilatation produced by verapamil
is not ascribed to its pH (mean pH: 6.609) because application of
Elliot’s A solution (mean pH: 6.507) had no such effect.

Slight pial or subarachnoid bleeding was incurred in some
animals with the injection of the PRP-ADP. In 2 animals this
was felt to be of a significant magnitude and these cases are
analysed separately. In these 2 cases subarachnoid hemorrhage
was produced by the nicking of a side branch and produced
spasm, of a similar magnitude and time course as that produced
by PRP-ADP, which was readily reversed with verapamil (Table
3). A statistically insignificant increase in the mean arterial
blood pressure (MABP) was noted at 5 and 30 minutes after the
subarachnoid injection of PRP-ADP (Table 4). This was not
seen with verapamil. There were no significant changes in the
pCO₂ values during the course of the experiments (Table 5).

<p>| Table 1: Effects of a first treatment with PRP-ADP and verapamil on the diameter of the basilar artery |</p>
<table>
<thead>
<tr>
<th>Time (min)</th>
<th>n</th>
<th>PRP-ADP % Change (SEM)</th>
<th>Verapamil % Change (SEM)</th>
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<tbody>
<tr>
<td>5</td>
<td>8</td>
<td>-40.73 (2.84)</td>
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</tr>
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<td>10</td>
<td>8</td>
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<tr>
<td>90</td>
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<td>+6.78 (7.52)</td>
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n = number of observations
SEM = standard error of the mean
PRP = platelet-rich plasma
ADP = adenosine diphosphate

Figure 2 — Effects (mean and SEM) of the subarachnoid injection of PRP-ADP (closed arrow) and of the topical application of verapamil (open arrow) on the basilar artery of the cat. The negative percentages indicate vasoconstriction and the positive percentages indicate vasodilatation. Vasoconstriction is produced by the injection of PRP-ADP and persists until it is reversed by the application of verapamil.
Table 2: Effects of PRP-ADP and verapamil on the diameter of the basilar artery 60 minutes after a prior application of verapamil

<table>
<thead>
<tr>
<th>Time (min)</th>
<th>n</th>
<th>PRP-ADP % Change (SEM)</th>
<th>Verapamil % Change (SEM)</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>6</td>
<td>-29.52 (19.17)</td>
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</tr>
<tr>
<td>10</td>
<td>6</td>
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<td>15</td>
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<td>-30.74 (14.88)</td>
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<td>60</td>
<td>4</td>
<td>+1.63 (10.67)</td>
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</tr>
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<td>75</td>
<td>1</td>
<td>+11.60</td>
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</tr>
<tr>
<td>90</td>
<td>1</td>
<td>-0.89</td>
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PRP = platelet-rich plasma  
ADP = adenosine diphosphate

Table 3: Effects of subarachnoid hemorrhage and verapamil on the diameter of the basilar artery

<table>
<thead>
<tr>
<th>Time (min)</th>
<th>n</th>
<th>SAH % Change (SEM)</th>
<th>Verapamil % Change (SEM)</th>
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</thead>
<tbody>
<tr>
<td>5</td>
<td>2</td>
<td>-35.34 (14.66)</td>
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<tr>
<td>10</td>
<td>2</td>
<td>-38.95 (2.94)</td>
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<td>-44.18 (1.18)</td>
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</tr>
<tr>
<td>20</td>
<td>2</td>
<td>-35.40 (5.53)</td>
<td></td>
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<tr>
<td>25</td>
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<td>-32.35 (2.65)</td>
<td></td>
</tr>
<tr>
<td>30</td>
<td>2</td>
<td>-28.89 (3.50)</td>
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<tr>
<td>35</td>
<td>2</td>
<td>+63.98 (12.57)</td>
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<td>55</td>
<td>1</td>
<td>+57.89</td>
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</tr>
<tr>
<td>60</td>
<td>1</td>
<td>+47.36</td>
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</table>

SAH = subarachnoid hemorrhage  
PRP = platelet-rich plasma  
ADP = adenosine diphosphate

Table 4: Values of mean arterial blood pressure during the initial PRP-ADP and verapamil experiments

<table>
<thead>
<tr>
<th>PRP-ADP</th>
<th>Verapamil</th>
</tr>
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<td>Mean</td>
<td>SD</td>
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<tr>
<td>5 min. before treatment</td>
<td>138.75</td>
</tr>
<tr>
<td>5 min. after treatment</td>
<td>148.25</td>
</tr>
<tr>
<td>30 min. after treatment</td>
<td>156.57</td>
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PRP-ADP = Platelet-rich plasma and adenosine diphosphate  
SD = Standard Deviation

Table 5: Values of the pCO₂ during the initial PRP-ADP and verapamil experiments

<table>
<thead>
<tr>
<th>PRP-ADP</th>
<th>Verapamil</th>
</tr>
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<tbody>
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<td>n Mean</td>
<td>SD</td>
</tr>
<tr>
<td>5 min. before treatment</td>
<td>6</td>
</tr>
<tr>
<td>5 min. after treatment</td>
<td>4</td>
</tr>
<tr>
<td>30 min. after treatment</td>
<td>6</td>
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</table>

DISCUSSION

Despite more than twenty years of laboratory experimentation and numerous trials of treatment of the clinical syndrome with a variety of drug regimens (Sundt et al., 1973; Zervas et al., 1979; Flamm, 1980) there is no universally effective pharmacological treatment of cerebral vasospasm. One of the major factors responsible for this is the difficulty of producing a reliable model for the production of delayed vasospasm that mimics the clinical condition. Most of the advances in our understanding of vasospasm, therefore, have come from experimentation on acute experimental models. The present study uses Echlin's model (Echlin, 1971) as modified by Kapp and his co-workers (Kapp et al., 1968) to insure reproducibility by stabilizing physiological conditions. We have further refined this model so as to avoid the mechanical trauma incurred with the resection of the arachnoid membrane and to eliminate the risk of drying of the blood vessels, shown by Kapp et al. to have a potent vasoconstrictive effect (Kapp et al., 1968). This is accomplished by leaving the arachnoid membrane intact and injecting the spasmogen directly into the SAS with a fine needle. The presence...
of a relatively intact arachnoid membrane also prevents any ambient blood from coming into direct contact with the vessels under investigation. Besides eliminating these potential sources of artifactual vasoconstriction, the injection of the spasmogen into the SAS more closely reproduces the acute phase of SAH. The subarachnoid injection of PRP-ADP and subarachnoid hemorrhage produced by nicking of a side branch of the basilar artery produced a statistically insignificant elevation of the mean arterial blood pressure (MABP). Espinosa and co-workers noted a 25% increase in MABP in 50 to 70% of their monkeys during the creation of chronic experimental SAH (Espinosa et al., 1982). They found no relationship between this increase in MABP and the development of vasospasm at any time after SAH. In our experiments the increase in MABP may represent a reflex response to brainstem ischemia secondary to the constriction of the basilar artery. The choice of ADP-activated PRP to cause acute experimental vasospasm is pertinent in 3 ways: (1) Platelet aggregation has been noted to be increased in patients with spontaneous coronary artery vasospasm (Robertson et al., 1980). By analogy abnormally increased platelet aggregation may be a factor in the genesis of cerebral vasospasm. (2) The platelet fraction of whole blood contains a number of vasoactive compounds which may be involved in the genesis of the acute phase of cerebral vasospasm. (3) Most of the putative spasmogens associated with platelets are produced by brain tissue (Wolfe, 1981). Endogenous biosynthesis of PG-E2, PG-F2 alpha, and PG-D2 has been demonstrated in the cat cerebral cortex, hippocampus and cerebellum and synthesis of these compounds and of thromboxane B2 has been demonstrated in the rat and guinea pig cerebral cortex (Wolfe, 1981). Predominant amongst these, in the cerebral cortex of most of the species studied, is PG-F2 alpha, which has been demonstrated to be a potent vasoconstrictor. Prostaglandins F2 alpha and E2 are seen in increased amounts in the CSF of patients with subarachnoid hemorrhage, stroke, meningoencephalitis and following brain surgery, all conditions which are often associated with vasospasm (Wolfe and Mamer, 1974; Carasso et al., 1977; Hagen et al., 1977).

The final common pathway leading to vasospasm must involve calcium-mediated actin and myosin coupling (Peterson and Leblanc, 1976). Interference with this process by chelation with EDTA or by promotion of cyclic-AMP-mediated calcium ion sequestration has been demonstrated to reverse experimentally induced vasospasm (Peterson et al., 1969; Peterson et al., 1973a; Peterson et al., 1973b; Peterson et al., 1975). Our experiments document that the topical application of a calcium antagonist results in consistent and dramatic reversal of acute experimental vasospasm. They also demonstrate that the effects of this drug are transient, and that vasospasm can be induced in the basilar artery 60 minutes after treatment with verapamil. This indicates that as long as blood vessels are exposed to spasmogens, a continuous supply of calcium blockers may be required to assure ongoing antagonism of their action. This may limit the usefulness of verapamil itself, given systemically, in view of its known hypotensive effects (Lewis, 1980). Nonetheless, calcium channel blockers have marked spasmytic properties and therefore show promise for the treatment of cerebral vasospasms (Allen et al., 1983).

ACKNOWLEDGMENT

We are grateful to Dr. Leon S. Wolfe, head of the department of neurochemistry, of the Montreal Neurological Institute and McGill University for fruitful discussions and advice. We are also grateful to Mrs. Janet Arts for her able technical assistance. This work was supported by the Medical Research Council of Canada and by the William Cone Laboratory for Neurosurgical Research.

REFERENCES


