Effects of intraruminal sodium chloride infusion on rumen and renal nitrogen and electrolyte dynamics in sheep

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1. Sheep were given 800 g low-protein roughage/d at 2 h intervals and infused intraruminally with 0, 500, 750, 1000, 1250, 1500 or 2000 mmol sodium chloride/d in 436 ml water. The digestibility of various food fractions and rumen ammonia, volatile fatty acids (VFA) and liquid turnover rate were measured, along with renal haemodynamics and the renal excretory patterns of nitrogen and electrolytes. Ad lib. food intake was determined during the infusion of 0 and 2000 mmol NaCl/d.

2. Infusion of NaCl up to 750 mmol/d had virtually no effect on the indices measured, except water intake and water excretion. Infusion of greater amounts caused a step-wise decrease in the digestibility of organic matter (OM) and N. Rumen liquid turnover rate was increased substantially and rumen NH_3 and VFA concentrations were decreased. Ad lib. food intake was not different when either 0 or 2000 mmol NaCl/d were infused into the rumen.

3. The glomerular filtration rate and effective renal plasma flow (ERPF) were substantially increased after the infusion of 1250 mmol or more NaCl/d. Extracellular fluid volume was also increased. The renal excretion of urea and uric acid+allantoin (URAL) were decreased at the higher infusion rates but the fractional excretions of both these substances were enhanced. The excretion of sodium, chloride, calcium and magnesium were markedly increased with increasing salt infusion.

4. The results suggest that high NaCl inputs into the rumen increase the rumen turnover rate, which in turn decreases the digestibility of OM, particularly N. This causes lower rumen NH_3 and VFA concentrations. Plasma urea and URAL concentrations are also decreased and this causes lower renal excretion of these substances despite a much higher fractional excretion resulting from the greatly enhanced urine flow rate.

5. When roughages low in N are given, NaCl intake should be kept below 20 mmol/kg body-weight per d to prevent a decline in the digestibility of the food and any consequent reduction in protein available to the sheep.

In many areas of the world sheep consume plants or drinking water which have a high sodium chloride content. For example, Wilson (1966*a*) found that sheep grazing *Atriplex* spp. as a major food constituent had salt intakes as high as 4217 mmol/d.

Some effects of high NaCl intakes on food utilization have been determined. Croom *et al.* (1982) noted that the addition of 50 g NaCl/kg to high-grain diets increased the efficiency of organic matter (OM) utilization in steers. Hemsley *et al.* (1975) added 1366 mmol NaCl/d to a linseed-meal diet and 171 mmol/l to the drinking water of sheep and showed that this treatment reduced rumen OM digestion by 24% and rumen protein digestion by 10%. Further, Hemsley (1975) found that similar treatment produced a substantial increase in wool growth.

The recycling of plasma urea to the rumen has been extensively studied. It returns to the rumen directly across the rumen epithelium (Houpt & Houpt, 1968) and also via saliva (Somers, 1961). In animals given low-protein or high-energy diets, or both, returned urea helps maintain rumen ammonia above a concentration which limits microbial protein synthesis (Bondi, 1981).

The level of urea in the plasma is a major determinant of the amount of nitrogen recycled to the reticulo-rumen. The plasma level may be affected by NaCl intake, as sheep loaded with urea and infused intraruminally with NaCl showed an increased renal excretion and a consequent lowering of the plasma concentration of urea compared with animals not given NaCl (Godwin & Williams, 1984).

A decrease in urea recycling and a decrease in rumen turnover time should reduce the

digestion of low-protein roughages in the rumen. The present study examines the effects of high levels of NaCl on the digestibility of food and the dynamics of the rumen and kidneys of sheep given a low-protein roughage.

A summary of part of this work has been published (Godwin & Williams, 1983).

MATERIALS AND METHODS

Animals, method of feeding and diet

Twelve merino ewes, aged 2–4 years, and weighing 39.8 ± 1.0 kg body-weight were dosed with anthelmintic (Ranide; Merck, Sharpe & Dohme), fitted with a rumen cannula under surgical anaesthesia and individually housed in metabolism cages. They were given the experimental diet for at least 1 month before any experimental treatment was imposed.

The treatment group of eight animals was fed automatically (Nicol & Corbett, 1971) with one-twelfth of their daily ration at 2 h intervals.

The four control sheep (not infused with NaCl) were fed once daily at 12.00 hours to determine whether there was any consistent change in food intake or apparent digestibility during the experimental periods.

Food consisted of 96 g lucerne (*Medicago sativa*) chaff plus 704 g oaten chaff, with a moisture content of approximately 100 g/kg. The daily food supply had a gross energy value of approximately 16.9 MJ and contained approximately 6.8 g N, 127 mmol sodium, 112 mmol potassium, 86 mmol calcium, 55 mmol magnesium and 30 mmol phosphorus. All animals were dosed weekly with 7500 μ g vitamin A and 125 μ g vitamin D₃ (Vetemul; ICI).

Experimental procedures

Experimental group. The animals were infused intraruminally with salt solutions containing 0, 500, 750, 1000, 1250, 1500 or 2000 mmol NaCl/d in 436 ml water. Infusions were in this order for all sheep to reduce adaptation periods to the higher salt levels. Infused salt was considered as dietary for digestibility measurements.

Following a 9 d equilibration on each salt solution one jugular vein (1.5 mm OD), medical grade polyvinyl tubing; Dural Plastics), and the bladder (Folatex Balloon Paediatric Catheter 8 Ch; Euro-Medical Industries, Sussex) were catheterized. The jugular catheter was flushed daily with approximately 10 ml heparinized saline (9 g NaCl/l). The following morning a 3 d urine collection was commenced. At the end of this urine collection an intravenous infusion of 0.5 ml/min of a solution containing (g/l) 100 inulin, 50 *p*-amino hippurate (PAH) and 8 NaCl was commenced at 12.00 hours; blood samples were obtained from the contralateral jugular vein by needle puncture at 14.00, 16.00 and 18.00 hours and urine collected over the same period. Glomerular filtration rate (GFR) and effective renal plasma flow (ERPF) were calculated from inulin and PAH clearances respectively over this 4 h period. The correction of Rabinowitz *et al.* (1971) for plasma protein binding of PAH was used. The day following GFR and ERPF measurements, extracellular fluid volume (ECF) was estimated by injecting 20 ml inulin solution (200 g/l) intravenously and sampling blood from the contralateral vein at 3, 7, 20, 35, 50, 65, 80 and 100 min following injection. Calculation of ECF was according to the method of Poulsen *et al.* (1977).

Rumen fluid volume and outflow rate were measured on the same day as ECF by injecting 200 ml of a solution containing 1-108 g chromium as CrEDTA into the rumen and serially sampling rumen contents for 36 h. Calculations were by the formulae presented by Faichney (1975).

Following the previously described procedures, digestibility of the food was measured over a 10 d period. Daily faecal samples were dried at 95° for 4 d and pooled for each animal for later analysis.

Because substantial differences were noted in the digestibility of food and the rumen turnover rate when the animals were given the lowest and highest salt infusions, *ad lib*. food intake was measured at these two infusion levels. This study was made following the end of the collection period for the 2000 mmol NaCl/d infusion. Four of the animals were maintained on an infusion of 2000 mmol NaCl/d whilst the other four were infused with water only for 13 d. The diet, 120 g lucerne chaff plus 880 g oaten chaff/kg, was fed *ad lib*. during this period. Digestibility and food intake were determined over the following 10 d. The four animals that were infused with salt were then given water and vice versa. This salt infusion commenced at 500 mmol/d and increased to 2000 mmol/d over the next 10 d. These infusions were maintained over the next 20 d with measurements taken during the last 10 d. Samples of rumen fluid were collected at approximately 21.00 hours on the first and last day of these collection periods for determination of rumen indices.

Control group. During the time the experimental groups were given 800 g roughage/d, the control animals were given the same amount of food and any refusals were weighed and recorded. Digestibility of the ration was measured six times at about six-weekly intervals. Food was offered *ad lib.* during the same period as the experimental group. Intake and digestibility of food were also determined during this treatment. Samples of rumen fluid were taken at about 21.00 hours on the first and last day of each digestibility trial.

Analytical. Plasma and urine Mg, inorganic P (P_i), Ca, glucose, urea, uric acid + allantoin (URAL), creatinine and plasma protein were measured with a Cobas-Bio centrifugal analyser (Hoffman–La Roche, Switzerland). Pierce Kits (Pierce Chemical Co. Illinois) were used for Mg and P_i and Hoffman–La Roche kits for the remainder. Na and K were determined by flame photometry (EEL 227; Evans Electroselenium, Essex) and Cl was measured using a CMT 10 chloride titrator (Radiometer, Copenhagen). Inulin and PAH were determined by the methods of Bacon & Bell (1948) and Smith *et al.* (1945).

Faeces, food and urine were digested by the method of Cresser & Parsons (1979). Total N was estimated in the digest by steam distillation and titration and Na, K and Mg by atomic absorption spectrophotometry (SB 900; GBC Scientific Equipment, Melbourne). Ca and P_i were determined as for plasma.

 NH_3 concentration was measured in rumen fluid, after protein precipitation with ethanol, and also in urine, by distillation and titration. Urine urea and URAL values were corrected for NH_3 content from this value. Rumen total VFA was also determined by distillation of the supernatant fraction from centrifuged rumen contents and subsequent titration.

Osmolality of rumen fluid and urine was measured by freezing-point depression (Fiske Associates, Connecticut), Cr was determined in rumen fluid and in urine by atomic absorption spectrophotometry and packed cell volume (PCV) was estimated by a micromethod (Hawksley and Sons, Sussex). Organic matter of food and faeces was obtained by ashing for 2 h at 400° followed by 12 h at 600° and the energy contents of food and faeces were estimated by ballistic bomb calorimetry (Ser 330; Gallenkamp, Sussex). pH of urine and rumen fluid was measured with a glass electrode (Radiometer).

RESULTS

Control animals

Appetite. The animals when given 800 g/d showed no decline in food intake and refusals were rare throughout the experiment.

Apparent digestibility. There was little change in digestibility. The values for energy increased slightly after the first 6 weeks and then remained at this level (Table 1).

Ad lib. values. Ad lib. food intakes, digestibility of the various fractions measured and rumen fluid values are shown in Table 1. Ad lib. food intake by all sheep was greater than 800 g, the amount fed during the previous periods; the mean increase was 14%.

Feeding regimen							R	Restricted to 800 g/	to 800 g,	P/						15
Interval‡	1			3		~	-	4		5		9	-	1–6	intake	w. ike
	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM
Food intake (g/d)	800		800		800		800		800		800		800		912†	24.1
Apparent digestibility:																
Dry matter	0-62	0-012	0.62	0.014	0-64	0-018	0-61	0.011	0.60	0-016	0-63	0-012	0.62	0.005	0.58†	0.019
Organic matter	0.51	0.016	0.55	0-021	0.53	0.032	0.54	0.032	0-52	0-019	0.50	0.041	0.53	0.008	0.48+	0.041
Energy	0.52*	0.022	0.62	0.026	0-59	0-024	0-58	0-019	0-58	0.031	09.0	0.012	0·58	0.014	0.50	0.048
Nitrogen	0.61	0·008	0.64	0-041	0.62	0.034	0.63	0.016	0.64	0.021	0-59	0.013	0.62	0.008	0.55	0.039
Sodium	0·88	0.016	0·88	0.012	0-87	0-024	0-87	0.038	06-0	0.034	0.89	0.021	0.88	0-005	0·88	0-041
Potassium	0.88	0.032	0.88	0-031	0.86	0.029	0.90	0.034	0.85	0.048	7	Ģ	0.87	0.009	0.93^{+}	0-021
Calcium	0.16	0.031	0·13	0.031	0.16	0.046	Z	Ð	0-19	0-052	0.15	0.041	0.16	0.010	0.15	0.039
Magnesium	0.20	0.042	0.20	0.038	0.16	0.084	0.21	0-063	0.19	0-057	0.15	0.055	0·19	0.010	0.18	0.049
Phosphorus	0.22	0-054	0·15	0.083	0·25	0-076	0.19	060-0	0.26	0.068	0.19	0.072	0·21	0-017	0-05	0·043
Rumen indices:																
Ammonia (mmol/l)	5.4	0-84	6·1	0-73	4.9	0-61	5.4	0.83		0.70	5.2	0.98	5-4	0.16	5.4	0.48
VFA (mmol/l)	84	6.4	92	5.8	82	9.6	86	8:3	88	7-4	85	8·2	86	1:4	81†	3.6
Hd	6.34	0.291	09·9	0·312	6-52	0-411	6.31	0.210	4	Ð	6.40	0.263	6-43	0.055	6.32†	0.333

Table 1. The effect of ad lib. feeding on intake and apparent digestibility of food, and rumen indices of control sheep (Mean values with their standard errors for four sheen) I. R. GODWIN AND V. J. WILLIAMS

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ND, not determined; VFA, volatile fatty acids. * Significantly different from the mean 2-6 value (P < 0.01). † Significantly different from the mean 1–6 restricted value (P < 0.01). ‡ Intervals approximately 6 weeks.

The digestibility of all organic components and P was decreased with *ad lib*. feeding. However, values for Ca, Mg and Na were unaltered whilst the value for K was slightly increased. Rumen NH_3 was the same for both feeding levels but rumen VFA and pH were decreased with *ad lib*. feeding.

Experimental animals

Rumen fluid dynamics. Increasing salt infusions increased rumen fluid volume only slightly (from 5·1 to 5·5 litres) but rumen liquid outflow was markedly affected, rising from 3·42 to 8·12 litres/d with 0–2000 mmol NaCl infusion (Table 2). Analysis of urine for Cr indicated that a maximum of 3.6% was absorbed, which is considered of negligible importance.

Apparent digestibility. The digestibility of dry matter, OM, energy and N are shown in Table 3. No significant changes occurred in these values until 1000 mmol NaCl/d was infused. Each constituent then showed a linear decrement with additional salt. The apparent digestibility of the electrolytes measured are also shown in Table 3.

The digestibility of Na increased following the infusion of 500 mmol NaCl/d and remained at this level for the remaining infusates. K digestibility was significantly increased above the basal level with the infusion of 500 mmol NaCl/d, which was probably fortuitous as higher infusion rates had no effect. The digestibility of Ca increased with salt infusion until 1000 mmol/d were infused; thereafter it decreased slightly, but not to the pre-infusion value. Mg and P values were unchanged.

Rumen fluid constituents (Table 2). Rumen pH slightly increased from 6.35 to 6.83 with increasing salt infusion. No change occurred in rumen osmolality until 1000 mmol NaCl/d were infused; it then increased linearly with infusion rate. There was no effect on rumen NH₃ concentration until 1000 mmol NaCl/d were infused, then the values declined. Total VFA concentration followed a similar pattern to that of rumen NH₃, although the response was much more attenuated.

Plasma and renal N dynamics (Table 4). Urine flow rate increased linearly from 0.42 to 4.07 ml/min with the infusion of salt from 0 to 2000 mmol/d. GFR values were calculated from the renal clearances of both inulin and creatinine which correlated well $(y = 1.18x - 7.68, r^2 0.96, where x \text{ is creatinine and } y \text{ is inulin})$. The results presented are creatinine values. GFR showed no change until 1000 mmol NaCl/d were infused and it then increased in a curvilinear manner from 55.2 to 67.1 ml/min at 2000 mmol NaCl/d. ERPF increased linearly with the quantity of salt infused. Filtration fraction (FF), determined as the ratio, GFR :ERPF, was unaltered by the treatments. PCV remained relatively constant throughout the experiment.

Plasma urea level remained relatively constant at about 4 mmol/l until 1000 mmol NaCl/d were infused, when a large decrease to 1.7 mmol/l occurred. Increased salt infusions caused further slight decreases in the plasma level to 0.8 mmol/l with the maximum salt infusion.

Urea excretion was increased by the infusion of up to 750 mmol NaCl/d, but further salt input resulted in a decrease in excretion to below the pre-infusion value. Increasing salt infusion from 1250 to 2000 mmol/d caused no significant change in urea excretion. However, the fractional urea clearance (clearance of urea/GFR) increased with increasing salt infusion.

Plasma URAL showed no change until 1000 mmol NaCl/d were infused and thereafter it fell with increasing salt input. The plasma concentrations were all very low. The excretion of URAL showed a continual decline with increased salt input. However the fractional excretion of URAL increased with increasing salt intake after 750 mmol NaCl/d were infused. The values obtained with this level and subsequent levels of infusate were all greater than 100%, indicating substantial tubular secretion of URAL.

Volume (J) Liquid outflow (J/d) Ammonia (mmol/J) VFA	Mean 5 1 ^a 3 42 ^a 5 14 ^a 84 ^{ab} 284 ^a 6 35 ^a 1 09 ^a	SEM 0-29 0-511 0-826 4-2 5-0 5-0 0-094 0-314	Mean SEM M 5:2ab 0.48 5 5:93a 0.914 5 5:93a 0.914 5 88b 4.6 81 284a 4.2 283 6:35a 0.063 6 1.86b 0.124 2 VFA, volatile fatty acids. VFA, volatile fatty acids.	sem 0-48 0-643 0-914 4-6 4-2 0-063 0-124 0-124 with differ	Mean 5.2 ^{ab} 5.71 ^a 5.71 ^a 81 ^a 6.40 ^a 5.29 ^c 2.29 ^c cids.	sem 0.34 0.685 0.873 3.1 6.2 0.044 0.138	Mean 5-3 ^{ab} 4-88 ^{bc} 3-64 ^b 80 ^a 6-54 ^b 3-36 ^d	SEM 0-30 0-410 0-450 0-450 2-8 3-9 0-060 0-046 0-046	Mean sem Mean sem Mean sem Mean sem se	SEM 0.34 0.603 0.328 4.3 5.6 0.040 0.040	Mean 5-3 ^{ab} 6-90 ^{dl} 2-79 ^c 76 ^c 6-79 ^c 6-79 ^c 4-88 ^f	sem 0.42 0.728 0.647 2.0 4.2 0.074 0.074	Mean 5.5 ^b 8.12 ^e 2.21 ^e 75 ^c 312 ^e 6.83 ^e 6.36 ⁸	sem 0-23 0-887 0-964 3-4 3-4 3-8 0-121 0-121
Volume (1) Liquid outflow (1/d) Ammonia (mmol/1) VFA	5.1 ^a 3.42 ^a 5.14 ^a 84 ^{ab} 284 ^a 6.35 ^a 1.09 ^a	0-29 0-511 0-826 4-2 5-0 0-094 0-314	5.2 ^{ab} 3.81 ^{ab} 5.93 ^a 88 ^b 88 ^b 6.35 ^a 6.35 ^a 1.86 ^b	0.48 0.643 0.914 4.6 4.2 0.063 0.124 0.124 0.124 with diffe: with diffe:	5.2 ^{ab} 4.01 ^{ab} 5.71 ^a 81 ^a 6.40 ^a 6.40 ^a 2.29 ^c 2.29 ^c cids.	0.34 0.685 0.873 3.1 6.2 0.044 0.138	5.3 ^{8b} 4.88 ^{bc} 3.64 ^b 80 ^a 6.54 ^b 3.36 ^d	0-30 0-410 0-450 2-8 3-9 0-060 0-046 0-046	5.4 ^{ab} 5.61 ^c 2.71 ^e 79 ^{ac} 294 ^b 6.50 ^b 4.13 ^e 4.13 ^e	0:34 0:603 0:603 0:328 4:3 5:6 0:040 0:040	5.3 ^{ab} 6.90 ^{a1} 2.79 ^c 76 ^c 308 ^c 6.79 ^c 4.88 ^t	0.42 0.728 0.647 2.0 4.2 0.074 0.265	5.5 ^b 8.12 ^e 2.21 ^e 75 ^c 6.83 ^e 6.36 ^g	0.23 0.887 0.964 3.4 3.8 0.121 0.121
(l/d) Ammonia (mmol/l) VFA	5.14 ^a 84 ^{ab} 284 ^a 6.35 ^a 1.09 ^a	0.826 4.2 5.0 0.094 0.314	5.93 ^a 88 ^b 284 ^a 6.35 ^a 1.86 ^b 1.86 ^b	0-914 4-6 4-2 0-063 0-124 0-124 0:124 with diffe:	5.71 ^a 81 ^a 283 ^a 6.40 ^a 2.29 ^c cids.	0.873 3.1 6.2 0.044 0.138	3.64 ^b 80 ^a 6.54 ^b 3.36 ^d	0.450 2.8 3.9 0.060 0.046	2.71° 79ªc 294 ^b 6.50 ^b 4.13 ^e	0.328 4.3 5.6 0.040 0.264	2.79c 76° 308° 6.79c 4.88f	0.647 2.0 4.2 0.074 0.265	2.21° 75° 312° 6.36 [¢] 6.36 [¢]	0.964 3.4 3.8 0.121 0.308
(mmol/l) VFA (mmol/l)	84 ^{ab} 284 ^a 6.35 ^a 1.09 ^a	4-2 5-0 0-094 0-314	88 ^b 284 ^a 6·35 ^a 1·86 ^b	4·6 4·2 0·063 0·124 tile fatty a with diffe:	81ª 283ª 6.40ª 2.29 ^c cids.	3.1 6.2 0.044 0.138	80° 296 ^b 6·54 ^b 3·36 ^d	2-8 3-9 0-060 0-046 ificantly c	79ac 294 ^b 6-50 ^b 4-13 ^e	4·3 5·6 0·040 0·264 ○	76° 308° 6·79° 4·88 ^t	2.0 4.2 0.074 0.265	75° 312° 6.36 ⁸	3.4 3.8 0-121 0-308
	284ª 6.35ª 1.09ª	5-0 0-094 0-314	284 ^a 6-35 ^a 1-86 ^b VFA, volal	4.2 0.063 0.124 tile fatty a with diffe:	283 ^a 6.40 ^a 2.29 ^c cids.	6-2 0-044 0-138	296 ^b 6-54 ^b 3-36 ^d	3-9 0-060 0-046 ificantly c	294 ^b 6.50 ^b 4.13 ^e	5.6 0.040 0.264 ○ < 0.01).	308° 6.79° 4.88f	4·2 0·074 0·265	312° 6.83° 6.36 ⁸	3.8 0-121 0-308
Osmolality	6.35ª 1-09ª	0.094	6.35ª 1.86 ^b VFA, volaf	0-063 0-124 tile fatty a with diffei	6.40 ^a 2.29 ^c cids.	0-044 0-138	6.54 ^b 3.36 ^d	0-060 0-046	6·50 ^b 4·13 ^e	0-040 0-264 < 0-01).	6.79° 4.88 ^f	0-074 0-265	6.3° 6.36 ⁸	0-121 0-308
pH	1.09ª	0.314	I-86 ^b VFA, volat	0.124 tile fatty a with diffe	2.29° cids.	0.138	3.36 ^d	0-046 ifficantly ¢	4·13 ^e	0.264	4.88 ^f	0.265	6.368	0-308
water intake (1/d)			VFA, volat	tile fatty a with differ	cids.		-	ufficantly d	lifferent (F	< 0.01).				
Salt infused (mmol/d)		0	50	500	75	750	10	1000		1250		1500		2000
digestibility	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM
Dry matter Organic matter	0-65 ^a 0-54 ^a	0-012 0-024	0.65ª 0.56ª	0-013 0-021	0-63 ^a 0-54 ^a	0-011 0-021	0.50b 0.50b	0-010 0-019	0.59b 0.48bc		0.57 ^b 0.46 ^c	0-026 0-023	0-53° 0-45°	0-021 0-020
Energy	0.56^{a}	0-036	0.56ª	0-031	0.55ª	0-029	0.52 ^b	0.016	0.50°		0.49°	600·0	0.48°	0-012
Nitrogen	0.628	0-036	0.00b	0-041	0.05 ^a 0.01b	0-026	0.200 0.80b	0-026	0.20 ⁰	0-027	°10.0 db0.0	0-019 0-040	0.46 ⁴	0-030
Potassium	-co.0 868-0	0-014	0.95 ^b	0.012	-16-0	0-026	-60-0	0-028	0-89 ^a		0.88ª	0-036	0.85ª	0.042
Calcium	0.12ª	0.023	0·19 ^b	0-028	0.19^{b}	0.020	0.24°	0-029	0.21bc		0.20^{b}	0-017	0.18 ^b	0-036
Magnesium	0.10	0-042	0.18	0-038	0.16	0-051	0.22	0.036	0.21	0-051	0-18	0-046	0-16 0-18	0-043

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^{a-d} Means with different superscript letters were significantly different (P < 0.01).

Salt infused														
···(n/1011111)	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM
GFR (ml/min)	57.3ª	3·63	54-4 ^{a b}	6.40	48-3 ^b	7.19	55.2 ^{a b}	6.14	60.2 ^{a c}	4.21	64.9 ^{cd}	4-31	67.1 ^d	5.48
ERPF (ml/min)	413 ^a	5.1	424 ^b	4.6	436°	4·8	439 ^c	4.6	449d	5.3	458 ^e	6.2	471 ^f	0·L
FF Č	0·14	0.05	0.13	0.05	0.11	0·0	0.13	0-02	0.14	0.04	0·14	0-04	0.14	0-05
ECF (I)	7.8	1.21	6.7	96.0	8-0	0.94	8.3	0.75	8.3	0.68	6.9 6	0.70	8.4	1.13
UFR (ml/min)	0·42ª	0.22	0-95b	0.09	1.24°	0.12	1.99 ^d	0.32	2.52 ^e	0-25	3·11 ^f	0-38	4-07 ^g	0.62
Plasma urea	4·] ^a	0.94	3.6	0.84	4·0ª	0-92	l·7b	0-93	qI·I	0·84	1.0 ^b	0-51	0·8 ^b	0.63
(mmoi/l) Plasma TTRAL	0.178	0-03	0.14a	0-06	0.00a b	0.03	0-07b	0-07	0.05bc	0.07	0.030	0-02	0.030	0.03
(mmol/l)		>					>					5		5
Plasma protein	0.9	0-46	6.2	0.39	6.1	0-40	6.1	0.51	9-0	0-31	6·2	0-39	6·2	0-38
Urea excretion	14.9a	2.64	15.6ª	3.16	17-6ª	5-01	12.9ab	2.65	10.9bc	2.14	10.9bc	2.45	9.70	1.20
(µmol/min) Fractional urea	0.06^{a}	0.05	0.08^{a}	0.06	0.9ª	0.03	0-14b	0-03	0.16 ^b c	0.02	0.17°	0.03	0.18 ^c	0.04
excretion														
URAL excretion	6·2ª	0.42	6·2ª	0.43	6·0ª	0.40	6·0ª	0-36	5.0 ^b	0·38	4.6 ^{bc}	0·46	4·]°	0·38
Fractional URAL	0.90a	0.10	0.81 ^a	0.22	$0.14^{\rm b}$	0.12	0.15 ^b c	0-02	1.66°	0.15	2.38 ^d	0-35	2.05d	0.25
excretion	1	:										1		
NH ₃ excretion	2.5ª	0-41	2.3ª	0-56	2·4ª	0-65	2-6 ^a	0.48	2.6 ^a	0.56	3-0ª b	0-51	4.2 ^b	0-95
N excretion	2.2 ^a	60·0	2.2 ^a	60-0	2.3ª	0·11	2.0	0-29	1-9 ^b	0.05	2.0 ^{a b}	0-26	1.9 ^b	0.07
(g/d) PCV	35	3·1	35	2.6	35	2.0	35	3.5	36	2.6	35	2.4	35	1.9

t infinsed	C	_	500	0	75	750	10	1000	1	1250	15(1500	JUC	0000
mol/d)			2	\$		2		8	1	2	5	8	51	2
··· (n /mm	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM
odium	141	2.1	143		144		142	1	143	2.1	139	4.6	138	4.8
tassium	4.2	0·21	4-4	0.24	4.5	0.24	4.6	0-84	4.5	0.15	4:3 6	0.15	4.3	0.21
lcium	2.41	0.15	2-41		2.43		2.45	_	2.41	60·0	2.39	0.08	2.40	0.11
Magnesium	1.35 ^a	0.11	1.29ª b		1.29ª b		1.27ª b		$1.24^{\rm b}$	0.10	1.16°	0.05	1.16°	0·0
osphorus	1.69ª	0.02	1.72ª		1 · 74ª		1.82^{b}	-	1.91°	0.031	1-99 ^d	0-04	2.10^{e}	0·0
hloride	111	2.6	111	2.2	112		110		111	3·1	110	2.4	114	3.8
lucose	3.9	0.81	3.7	0·76	4·0		3.8	-	3.8	0.80	3.9	0.75	3.6	0·82

Table 5. The effects of sodium chloride infusion on plasma levels of electrolytes and glucose (mmol/l) in sheep

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Table 6. The effect of sodium chloride infusion on the renal excretion of electrolytes, glucose and urine pH and osmolality in sheep (Mean values with their standard errors for eight sheep)

387 Effects of salt on rumen and renal dynamics 0.00002 0.0043 0-045 0.005 0.001 0.25 12.19 0.05 0.01 4·87 0.62 0.200.03 SEM 10<u>·</u>0 31-3 15.8 2000 8-02^b 0.13^b 0.21^d 0.158 Mean 0-06d 0.13^b 0-00° 0-00° 61·1^d 2.4c 10·0 9.4^r 9.9d 1211·2⁸ 7738 11308 0-00020.005 900·0 0.0430.0040.0030·002 0·19 41-32 1-94 0.05 0·12 0.52 0.03 SEM 12.8 18.7 1500 0.001^b 0.23^{ed} Mean 600·0 0-01c q00-8 64-8^{cd} 0.12^b 2.2bc 0.03° 0·12f 0·1p 937-8^r 4.7^e 9.2° 855f 748f 0.0001 0·004 0·110 0.0040.0040.001 0-011 0-46 0-24 38.69 3.06 0.030.01 0·21 SEM 21·1 20·8 1250 0.003ª b 8-04^b Mean 0.12^{b} 0-03b 0.11e 0.007 0-09a 0.26° 0.03° 803·2^e 69-2° 4-4d 1 · 7ª 8.9° 702^e 806e 0.00020-0021 0·214 0.004 0.0140.002 0.005 31-62 SEM 3.01 0·02 0.36 0-62 0.030.24 28.6 26.0 1000 0-02^bc 0.27^bc 0-002^a 600·0 a.01b Mean 1.9a b 68.6^bc 0.03c 0.08ª 0·1p 0.1^{d} 665-2^d 7.1^b 3.7c 894ª 591d 0-0002 0-0016 0.00360.018 0.0030.004 0.10154-90 2·19 0.29 0·28 0.02 0.84 0.0325.2 SEM 31.6 750 0.001^a Mean 0.08ª 0-32^a 0-02^b 0.13^b 0.04^b 0·0 1.9ª b 8.26^a 68-4^в 535-9° 2.1^b **∂**.7.8b 0·01 1201° 469c 0.00010-003 0-045 0·003 0·168 41.36 0.63 0·021 4-65 0.020.25 0.02 0.26 SEM 0·01 41.6 81·6 500 0.29ab 0.001^a 70-0ab Mean 0.01a 600·0 0.07^a 0.06^b 8-32^a 0.05ª 387-5^b ₫1.7 q I ۰O 1 · 7ª i-5ª 384^b 1502^b 0.00010.002 0·004 0·152 0.001 0-46 28-61 0.020.02 0.25 0·01 0.01 0.29SEM 5.21 22-4 21-4 0 0.001^a 72.61^a Mean 0.01^a 0.03ª 0-01^a 0.07a 0-08a 0.02^a 8-35^a 1-4^a 6.3ª 0·01 79.4^a 1 · 6ª 143^a 1248^a Magnesium excre-tion (µmol/min) glucose excretion tion (µmol/min) tion (µmol/min) tion (µmol/min) Potassium excretion (µmol/min) phate excretion norganic phos-Calcium excre-Fractional Mg Chloride excre-Fractional Na Fractional Ca Glucose excre-(mosmol/kg) (µmol/min) Fractional Fractional Cl Va excretion Fractional K (mol/min) inorganic P Salt infused (mmol/d).. Osmolality excretion excretion excretion excretion excretion excretion Fractional Urine pH

^{a-g} Means with different superscript letters were significantly different (P < 0.01)

Table 7. The effects of sodium chloride infusion on ad lib. food intakes and digestibility in sheep

Salt intake (mmol/d)	0		200	00
	Mean	SEM	Mean	SEM
Food intake (g/d)	946	36	908	41
Water intake (1/d)	1.16	0.294	6.38	0.949
Apparent digestibility:				
Dry matter	0.61	0.016	0.54*	0.011
Organic matter	0.52	0.010	0.46*	0.009
Energy	0.54	0.008	0.48*	0.009
Nitrogen	0.61	0.024	0.49*	0.048
Sodium	0.86	0.043	0.91	0.038
Potassium	0.88	0.061	0.86	0.038
Calcium	0.10	0.024	0.18	0.062
Magnesium	0.20	0.082	0.21	0.084
Phosphorus	0.14	0.028	0.10	0.038
Rumen indices:				
Ammonia (mmol/l)	5.6	0.74	2.8*	0.29
VFA (mmol/l)	80	3.8	71*	4.2
Osmolality (mosmol/kg)	281	6.9	328*	12.1
pH	6.44	0.058	6·71*	0.124

(Mean values with their standard errors for eight sheep)

VFA, volatile fatty acids. * P < 0.01.

 NH_3 excretion in urine was not altered by salt intake, except for the highest level of salt infusion, and then it was significantly increased. This was caused by the high excretion of NH_3 by one sheep only (13.8 μ mol/min).

The concentration of plasma protein remained relatively constant throughout the experiment.

Total N excretion was constant until 1000 mmol NaCl/d were infused and thereafter it decreased with increasing salt intake.

Plasma and urine electrolytes and glucose. Plasma electrolytes and glucose concentrations are shown in Table 5 and their renal excretion in Table 6. Plasma Na and Cl levels were virtually unaffected by the treatments, although plasma Na tended to be lower with the higher infusates. As expected the excretion and fractional excretion of Na and Cl increased with increasing salt intake. Plasma K concentration was unaffected by NaCl infusions. However, the excretion and fractional excretion of K were both reduced by the higher salt infusions. Plasma Ca levels were not altered by the salt infusions, but the excretion and fractional excretion were increased continuously with increasing NaCl input. Plasma Mg was significantly lowered by the infusion of 1500 and 2000 mmol NaCl/d. This corresponded with an increased renal excretion of Mg. Fractional Mg excretion was somewhat variable and was lowest when no salt was infused. Plasma P_i increased with increasing NaCl input. Urinary P_i excretion and fractional excretion were both very low and below detection limits, particularly on the high-salt infusions. Plasma glucose was not affected by the treatments. Excretion of glucose was highest when 2000 mmol NaCl/d were infused and urine flow rate was highest (Table 4). Fractional excretion of glucose was not greatly affected. Urine pH decreased after 1000 mmol NaCl/d were infused and remained at this level

for the remaining infusion periods. Urine osmolality increased when 500 mmol NaCl/d was infused, but thereafter decreased with increasing infusion of salt.

Ad lib. *feeding*. Table 7 shows the values obtained during *ad lib*. feeding at the two infusion levels, 0 and 2000 mmol NaCl/d. Food intake was approximately 18 and 13.5% greater than the restricted level previously fed for the infusion levels 0 and 2000 mmol NaCl/d respectively, but the difference in effect between these two infusion levels was not significant.

The apparent digestibility coefficients were similar to the values obtained when these animals were given the limited intake of 800 g/d and infused with the same amount of NaCl; thus, compared with the values in Table 3, the apparent digestibilities were virtually unaffected by the greater food intake. This also applies to the rumen indices shown in Table 2.

DISCUSSION

The rate that OM is removed from the rumen is important in limiting the intake of some roughages (Balch & Campling, 1962). The expected effect of infusing high levels of NaCl into the rumen was to increase rumen turnover rate and subsequently to stimulate a greater *ad lib*. food intake. However, increasing osmotic pressure of the gastrointestinal contents has been shown to inhibit food intake in non-ruminants, at least temporarily (Kissileff & Van Itallie, 1982), and in ruminants (Ternouth, 1968). Kato *et al.* (1979) provided evidence that the effect is one of ionic composition (i.e. high Na⁺ or K⁺ levels, or both) rather than the actual colligative properties of the rumen ingesta. In the present experiments the increase in rumen osmolality following the administration of salt was substantial, yet *ad lib*. food intake was unaltered. NaCl did not decrease food intake and any effect when the sheep were offered food *ad lib*, was very small and not statistically significant (Table 7). Nevertheless, the osmolality of the rumen fluid may have negated the possible increase in food intake due to increased rumen digesta turnover. The slight increase in rumen volume with the additional salt may also have had some inhibiting effect (Balch & Campling, 1962).

The increase in rumen liquid outflow with increasing salt input was probably caused mainly by increased water intake (Table 4). Although it was only liquid outflow that was measured, the substantial changes in this index and the decrease in OM digestibility (Table 3) suggest that particulate outflow was also enhanced. As rumen volume was relatively unaltered the changes in concentrations of the microbial metabolites, NH_3 and VFA, probably largely reflect changes in their production rates.

The decreasing dry matter digestibility with increasing NaCl levels was probably a result of two factors. First, the increased rumen outflow decreases the time available for microbial digestion and, second, the higher osmotic pressure of rumen contents reduces the digestion of cellulose (Bergen, 1972; Bennink *et al.* 1978). The substantial decrease in the digestibility of energy also included a nitrogenous component. Jackson *et al.* (1971) found that increasing the Na content of rations from 5 to 25 g/kg resulted in a decrease in carcass energy content despite a similar food intake, but digestibility of the food was not measured.

The control animals in the present experiment were included for two reasons: (a) it had been observed in our laboratory on previous occasions that when animals were given low-quality roughages they occasionally had periods of poor appetite and (b) they were used to detect any gross changes in appetite or digestibility associated with environmental factors as the experiment was conducted over a 10-month period and the experimental area, although under cover, was not temperature controlled or free of draughts. Although the control group was fed once daily, whereas the experimental group was fed twelve times daily, Faichney (1968) showed that feeding sheep at three-hourly intervals or once daily caused no differences in the digestibility of the food.

In the event no major changes occurred in the indices measured in the control animals indicating that the effects noted in the experimental group were a direct reflection of the salt infusions rather than any concomitant changes in the environment. It has been shown that environmental factors may alter rumen metabolism (Yousri *et al.* 1977; Christopherson & Kennedy, 1983).

Ad lib. feeding of the control animals resulted in a 14% increase in food intake and a decrease in the apparent digestibility of the organic constituents of the food. This is a common observation (Agricultural Research Council, 1980) and is caused mostly by a difference in the rate of passage of digesta (Grovum & Williams, 1977).

Most studies on the effects of NaCl on rumen function have been made in animals given concentrate rations, i.e. food high in fermentable protein that would also be highly digestible in the small intestine (Hemsley *et al.* 1975; Croom *et al.* 1982). Many studies have also included a mixture of minerals as a supplement, such as artificial saliva by which an additional buffering effect in the rumen is achieved (Harrison *et al.* 1975; Thomson *et al.* 1978).

Rogers *et al.* (1979) gave steers a dominantly roughage ration and infused 26 mmol NaCl/kg body-weight, which was approximately equivalent to the 1000 mmol NaCl/d infused in the present study, and lowered rumen total VFA by 29%. In the present experiment total VFA concentration was lowered by only 11% at the highest level of salt infusion.

To maintain maximum microbial protein synthesis, rumen NH_3 -N concentrations must be maintained above 5 mmol/l (Bondi, 1981). The striking drop in rumen NH_3 with increasing salt input was probably caused by an increased proportion of dietary protein escaping degradation in the rumen. However, much of this protein was not available for absorption from the lower gut as shown by the lower digestibility of N. The hydrolytic activity of the rumen microflora on carbohydrate components can be reduced by a lower rumen NH_3 level (Wallace, 1979), which also helps to explain the lower energy digestibility values with the high levels of salt.

The return of plasma urea to the rumen is highly dependent on the plasma urea concentration and the presence of readily fermentable carbohydrate in the rumen (Kennedy & Milligan, 1980). There is also evidence that salivary flow rate is decreased when dietary salt is high (Tomas & Potter, 1975). The lowered salivary flow rate coupled with the reduced plasma urea level would reduce the return of urea to the rumen by this route.

The renal excretion of urea was lowered by the addition of NaCl. This reduced excretion of urea was the result of greatly decreased plasma concentrations and consequently of reduced filtered loads to the renal tubules, because the fractional excretion of urea actually increased and this may be attributed to the higher urine flow rates, the major determinant of urea clearance (Tang-Liu *et al.* 1983).

Dixon & Milligan (1983) measured urea transfer to the rumen, and excretion of urea in the urine, in steers given grass hay and fresh water or 10 g NaCl/l in the drinking water. They found no effects on urea metabolism. However the salt intake from this treatment was only about 20 mmol/kg body-weight per d. In the present study this level, equivalent to about 750 mmol/d, had little effect on the indices of N metabolism measured. Previous work from this laboratory has shown that relatively small intraruminal infusions of NaCl (500 mmol/d) will reduce the high plasma urea concentrations of urea-loaded sheep by increasing the renal excretion of urea (Godwin & Williams, 1984).

Ergene & Pickering (1978) gave sheep a low-protein diet and provided them with fresh or salt water. The animals given the salt water consumed 652 mmol NaCl/d and had an increased tubular reabsorption of urea. However, these studies were confounded by the concomitant intravenous infusion of antidiuretic hormone (ADH) to stabilize urine flow rate (Ergene, 1976).

Valtonen (1979) gave reindeer a low-protein ration and supplemented this with 100–170 mmol NaCl/d. This level of supplementation decreased the renal tubular load of urea as a result of lowered plasma levels and consequently decreased urea excretion. This effect was similar to that shown in the present study with sheep, but it occurred at a much lower level of salt supplementation, well below that likely to have any profound effect on rumen function. These results may be related to the fact that the reindeer kidney has a much greater proportion of cortical relative to medullary tissue than the sheep and is relatively resistant to ADH (Valtonen & Eriksson, 1977).

The level of URAL excretion in sheep urine is determined predominately by the amount of microbial protein synthesis in the rumen (Rys *et al.* 1975), although changes in the level of endogenous allantoin excretion may introduce substantial errors into this supposition (Antoniewicz *et al.* 1980). In the present study the infusion of NaCl caused a substantial drop in plasma and urinary excretion of these purine metabolites. The more rapid rumen turnover with salt infusion and the concomitantly lower rumen VFA and NH₃ as well as the decreased URAL excretion indicates a lower level of rumen microbial protein synthesis. The increased renal fractional excretion of URAL with salt infusion was greater than unity in most cases which confirms the result of Chesley *et al.* (1978) that sheep secrete urate by a tubular mechanism. Increased salt intake does not affect uric acid excretion in man (Breslau & Pak, 1983).

The excretion of NH_3 in the urine was low and unaltered in all but one animal by saline infusion. The alkaline pH of the urine prevents the excretion of appreciable quantities of NH_3 , as its main function is to buffer secreted acid.

Total N excretion in the urine followed much the same pattern as urea, as urea was a substantial component. Brook *et al.* (1964) have suggested that catheterization of the bladder increases protein excretion in the form of mucus secreted from goblet cells along the urethra; therefore the values for total N excretion may be artificially high.

The lack of change in plasma protein and PCV suggests that blood volume was not altered appreciably, although ECF volume showed a trend toward greater values with increased salt input. This is a typical effect of increased NaCl intake (Dirks *et al.* 1976; Godwin & Williams, 1984). Potter (1961) found that sheep given 13 g NaCl/l as drinking water, through which they voluntarily consumed 1000 mmol NaCl/d for prolonged periods, have an increased GFR and a reduced ERPF resulting in a greater renal filtration fraction. In the present study, with this level of salt intake there was no change in GFR, ERPF or filtration fraction. However, Potter's (1961) animals were drenched with 1 litre of their drinking water just before the functional measurements.

Chronic salt loading results in an increased ERPF and ECF and, due to a reduced sensitivity of the tubulo-glomerular feedback mechanism in the kidney, increased GFR (Haberle & Davis, 1982). The enhanced ERPF may be due to an increase in systemic blood pressure, although Potter (1972) reports that the blood pressure of sheep is somewhat refractory to dietary NaCl.

The osmolality of urine samples showed a typical response to osmotic diuresis, i.e. a decrease with increasing salt intake.

The striking increase in Ca excretion with NaCl infusions is of interest as the digestibility of Ca did not follow the renal excretory pattern. The renal fractional excretion of Ca also increased more than five-fold. ECF volume expansion enhances Ca excretion (Massry *et al.* 1967). Recently Goulding & Campbell (1983) have shown that dietary NaCl enhances bone loss and Ca excretion in rats, thus it is possible that in sheep high NaCl intake causes bone Ca release. The renal clearances of Na and diffusible Ca are closely related (Massry *et al.* 1968) and Na is required for active Ca reabsorption (Borle, 1982). Na-induced hypercalciuria is accompanied by increased 1,25-dihydroxyvitamin D₃ synthesis and enhanced Ca absorption from the intestine of man (Breslau *et al.* 1982). In cattle, increasing

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the Na concentration of the rumen, omasum and, abomasum increases the absorption of Ca from these compartments (Timet *et al.* 1978). In the present study there was an increase in Ca digestibility until 1000 mmol NaCl/d were infused and thereafter it decreased slightly. This suggests that the Na-effect described by Timet *et al.* (1978) may have occurred up to this level and at higher Na levels the changes in the rate of passage modify the overall effect. Ca is mostly absorbed from the proximal small intestine (Phillipson & Storry, 1965), where Na is required for its optimal absorption (Martin & De Luca, 1969).

Mg absorption showed a similar pattern to that of Ca absorption, even though Mg is absorbed mostly from the rumen (Tomas & Potter, 1976). Increasing the Na⁺:K⁺ value of rumen contents increases the net absorption of Mg from an isolated sheep rumen (Care *et al.* 1981). Again this effect may be limited by an increased rate of passage of digesta at higher NaCl infusion rates. The increased excretion of Mg with the high levels of NaCl was probably the cause of the lowered plasma Mg levels. Increasing the water intake of sheep by intraruminal infusion causes increased urinary losses of both Mg and Ca (Suttle & Field, 1967).

The decrease found in P_i excretion may well be an analytical artefact as many of the urine samples, particularly the more dilute samples from salt-infused animals, were below detection limits for P_i . The urine of sheep given roughage rations normally contains very little P_i (Tomas, 1974; Godwin & Williams, 1982) with the major route of excretion being via saliva and hence in the faeces (Tomas, 1974). The increase in the plasma level of P_i , although substantial, was still below the threshold for P_i excretion (Watson, 1933) and hence the very low excretion rates. The increase in plasma P_i may be related to a reduced salivary flow, caused by the salt infusion (Tomas & Potter, 1975), or possibly by parathyroid hormone stimulation of osteoclastic activity in response to salt loading (Breslau *et al.* 1982; Goulding & Campbell, 1983). Parathyroid hormone is thought not to affect P_i excretion in sheep when plasma levels are below the threshold described by Watson (1933) (Clark *et al.* 1975).

Glucose excretion by ruminants is considerably higher than that of most other species (Macfarlane, 1976). The high salt infusion levels increased this excretion, probably simply as a result of diuretic wash out, although glucose excretion has been negatively related to plasma pH and urine pH (I. R. Godwin and G. A. Chaffey, unpublished results). In the present study, urine pH was decreased slightly by the salt infusion. ECF volume expansion also results in a shunting of blood from the medullary to the cortical areas of the kidney (Knox & Haas, 1982) which leads to an increased production of glucose by gluconeogenesis (McIntosh *et al.* 1973) which may possibly diffuse into the tubular fluid.

Under natural conditions salt may enter the animal in either the food or in drinking water. Addition of salt via these two sources may well have different effects, depending on the concentrations of salt and the frequency of feeding and watering (Wilson, 1966b). Continuous infusion of salt through a rumen fistula and free access to fresh water as used in the present study bypasses any effects on palatability and also reduces the likelihood of a rapid and large increase in rumen osmolality. The turnover of rumen contents occurs continuously and the animal can exercise no control over the amount and timing of its salt intake with continuous infusion. Drenching lambs with salt (34 mmol/kg body-weight per d) once daily causes little effect on growth rate (Hamilton *et al.* 1983). This appears to be due to a rapid wash out of rumen contents shortly after drenching, and a quick return to normal rumen function and eating behaviour (J. A. Hamilton, personal communication). Much of the literature on salt effects in sheep are confounded by differences in presentation of the salt and hence probable differences in gut function as a result. Further studies comparing the effects of these different modes of presentation are warranted.

In conclusion, 750 mmol NaCl/d may be infused intraruminally into sheep given

low-protein roughages with little effect except for increased water intake and excretion. Higher salt inputs cause an increased rumen turnover rate and a consequent lowering of dry matter digestibility and particularly N digestibility. This results in decreased rumen NH_3 and plasma urea levels and a decreased urea excretion rate. The kidneys also show an enhanced excretion of filtered urea and Ca. The previously described effects would almost certainly be detrimental to the production of animal protein.

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