Intrathecal Synthesis of Autoantibodies to Myelin Basic Protein in Multiple Sclerosis

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ABSTRACT: A solid phase radioimmunoassay was used to detect anti-myelin basic protein (MBP) antibodies in the CSF and serum of multiple sclerosis (MS) patients and controls. CSF and serum samples were assayed prior to acid hydrolysis in order to detect free anti-MBP as well as after acid hydrolysis to measure the total (free and bound) amount of antibody. An anti-MBP index controlling for serum levels as well as the degree of breakdown of the blood brain barrier was used to estimate intrathecal synthesis of anti-MBP. MS patients with acute exacerbations or chronically progressive disease have significantly elevated levels of both free and total CSF anti-MBP. The anti-MBP index is also significantly increased in MS patients with both forms of active disease. Anti-MBP antibodies are intrathecally produced in MS patients with active disease.

Multiple sclerosis (MS) is an acquired demyelinating disease of the central nervous system (CNS). The myelin sheath of the CNS is produced by oligodendrocytes. Myelin basic protein (MBP), a component of the myelin sheath, may be an important autoimmunogen in MS since it is capable of inducing experimental allergic encephalomyelitis, an animal model of MS.1 In MS patients with active disease, the cerebrospinal fluid (CSF) MBP is elevated, potentially serving as a chronic and recurrent autoimmunizing stimulus.2,3,4

Another feature of MS is increased IgG synthesis within the blood brain barrier (BBB).5-9 CSF IgG or IgG eluted from brain tissue of MS patients is characterized by oligoclonal bands superimposed on a background of polyclonal IgG.10-13 Oligoclonal banding is indicative of intrathecal IgG synthesis and it occurs in most MS patients. Whether they have active or inactive disease, the majority of clinically definite MS patients have increased intrathecal IgG synthesis.9 Theoretically, some of the IgG may be protective, while some may be involved in the pathogenesis of demyelination. The specificity of the majority of intrathecally produced IgG has not been elucidated.14,15

The recent observation of Dasgupta et al16 of MBP containing immune complexes in the blood of some MS patients led us to examine whether antibodies to MBP (anti-MBP) are intrathecally produced and whether their production is associated with disease activity.
METHODS

Anti-MBP levels were determined in CSF and serum before and after acid hydrolysis by a solid phase radioimmunoassay. The final IgG concentration in all CSF and serum samples was adjusted to 0.010 g/L. Immulon microtiter plates (96 wells/plate) were coated with human MBP (hMBP concentration = 1.0 ug/well). Staphylococcus A Protein (Staph A Prot) was iodinated (125I) by the method of Hunter and Greenwood. Aliquots of 100 ul CSF or serum (IgG concentration = 0.010 g/L) before and after acid hydrolysis were incubated in MBP coated wells for 2 hours at room temperature. After 5 washes, goat anti-human immunoglobulin was added and incubation continued for 1 hour at room temperature. After another 5 washes Staph A Prot (50,000 cpm/well) was added and the plates were incubated overnight at room temperature and finally, after 5 more washes the wells were individually counted. Results were expressed as percent (%) bound of total radioactivity (TC). Two positive (serum from a rabbit immunized with hMBP and a positive CSF pool) and two negative controls were used in each assay. When CSF samples with initially high IgG and anti-MBP values were serially diluted the anti-MBP levels (% bound) were parallel to the IgG concentrations. This assay was also validated by absorbing CSF anti-MBP with MBP prior to performing the assay. Absorption to MBP resulted in complete elimination of anti-MBP from samples that initially had high anti-MBP titers. Intraassay variability (CV) for 25 sets of duplicates was 3.86, while interassay variability (CV) for one set of quadruplicates over 10 different runs was 7.75. In order to determine nonspecific adherence of IgG to immulon plates, each sample was also assayed in wells that were not coated with hMBP. This nonspecific binding (<1% of TC) was subtracted from the matched counts of CSF and serum samples.

Anti-MBP levels were determined in 74 MS and 88 control patients. All MS patients had clinically definite disease. This group consisted of 15 patients in clinical remission with inactive disease (R) and 59 with active disease of which 31 had acute exacerbations (E) and 28 were experiencing chronically progressive MS (P). The control group consisted of 25 neurologically normal patients with psychoneurosis (N), 32 with degenerative disc disease (M) and 31 with various neurological diseases exclusive of MS.

Anti-MBP levels before and after acid hydrolysis in CSF and serum were expressed for each of the above clinical groups as % bound radioactivity ±2SD. Intergroup statistical differences were performed by Student’s t test. Intrathecal anti-MBP synthesis was calculated by the anti-MBP index:

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\text{Anti-MBP Index} = \frac{\text{CSF/Serum anti-MBP}}{1000 (\text{CSF/Serum Albumin})}
\]

This index was determined on the basis of post acid hydrolysis values. In this formula, the serum anti-MBP level controls for the amount of antibody present in the serum and the CSF/Serum albumin ratio controls for the degree of breakdown of the blood brain barrier which permits increased leakage into the CSF. Intergroup differences of anti-MBP indexes were again determined by Student’s t test.

All results are expressed as % bound radioactivity ±2SD. Nonspecific binding of IgG to immulon wells that were not coated with hMBP was subtracted from each sample value.

Results in 88 control patients were 2.3±0.5 in CSF with a corresponding serum value of 0.8±0.6. Patients with psychoneurosis and degenerative disc disease had similar low values in both CSF (1.5±1.7±0.4) and serum (0.7±0.5). This was due to nonspecific binding of immunoglobulins to MBP, and do not represent anti-MBP. These values were considered normal. The control neurological disease group had slightly higher CSF (3.8±0.6) and serum (1.1±0.6) results. While the majority of these patients with neurological diseases exclusive of MS had values similar to those of patients with psychoneurosis, a patient with subacute sclerosing panencephalitis (SSPE) and two of eight patients with post infectious encephalomyelitis (PIE) had higher results. The patient with SSPE had 16.0 in CSF and 3.9 in serum and the patients with PIE had 3.1 and 3.0 in CSF and 1.2 and 1.0 in serum. These CSF values were significantly increased and represent anti-MBP activity.

Anti-MBP was detectable in significantly higher amounts in CSF and serum of patients with active disease as opposed to those in remission or a group of controls (Table I, Figure 1). In all MS clinical subgroups except patients in remission, post acid hydrolysis levels were higher than prehydrolysis values. Data was analyzed on the basis of total anti-MBP levels obtained after acid hydrolysis.

[Figure I — Anti-MBP levels (mean ±2SD) in CSF (A) and serum (B) of multiple sclerosis (MS) and control (C) patients.]

PC: Student’s t test versus control patients
PR: Student’s t test versus MS patients in remission
Table 1: Anti-MBP Index and the effect of acid hydrolysis on anti-MBP levels in CSF and serum of multiple sclerosis (MS) patients and controls.

<table>
<thead>
<tr>
<th></th>
<th>n</th>
<th>CSF levels*</th>
<th>Serum levels*</th>
<th>Anti-MBP Index</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>-H*</td>
<td>+ H*</td>
<td>-H*</td>
<td>+ H*</td>
</tr>
<tr>
<td>Controls</td>
<td></td>
<td></td>
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<tr>
<td>N</td>
<td>25</td>
<td>0.4±0.2***</td>
<td>1.5±0.4</td>
<td>0.2±0.1</td>
</tr>
<tr>
<td>M</td>
<td>32</td>
<td>0.6±0.4</td>
<td>1.7±0.4</td>
<td>0.2±0.1</td>
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<tr>
<td>ND</td>
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<td>0.8±0.5</td>
<td>3.8±0.6</td>
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</tr>
<tr>
<td>Total</td>
<td>88</td>
<td></td>
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<td></td>
<td></td>
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<tr>
<td>R</td>
<td>15</td>
<td>1.6±0.5</td>
<td>1.6±0.4</td>
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</tr>
<tr>
<td>E</td>
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<td>18.8±7.8</td>
<td>3.4±3.0</td>
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<tr>
<td>P</td>
<td>28</td>
<td>24.4±5.7</td>
<td>53.6±15.6</td>
<td>2.5±0.8</td>
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</tbody>
</table>

* : Anti-MBP levels expressed as % bound radioactivity
** : CSF and serum samples assayed at 0.01 g/L IgG
*** : Results expressed as mean ±2SD

Anti-MBP Index = \( \frac{\text{CSF/serum anti-MBP}}{1000 \times \text{CSF/serum albumin}} \)

Patients subgroups:

CONTROLS:
- N = psychoneurosis
- M = degenerative disc disease
- ND = neurological diseases

MS:
- R = remission
- E = exacerbation
- P = progressive

Although MS patients in remission had increased levels of intrathecal IgG synthesis, their anti-MBP value was not detectable in either CSF (1.6±0.4) or serum (0.7±0.4). In contrast, MS patients with active disease had increased levels. Patients experiencing acute exacerbations had significantly elevated levels in both CSF (18.8±7.8) and serum (5.9±2.2). The highest CSF levels (53.6±15.6) were observed in MS patients with chronically progressive disease. Their serum level (3.6±1.0) was lower than patients with acute exacerbations. CSF and serum anti-MBP levels were significantly higher in MS patients with active disease versus MS patients in remission or control patients (Figure 1).

In the normal (N) control group, the anti-MBP index was 0.3±0.2 (Table 1, Figure 2). The index was not significantly elevated in MS patients in remission. Similar to anti-MBP levels, all MS patients with active disease had significantly higher anti-MBP indexes. The highest values were found in patients with chronic progressive disease (4.1±1.2). The index was 1.7±0.4 in patients with exacerbations.

DISCUSSION

Intrathecal IgG synthesis is a hallmark of MS. Although intrathecal IgG synthesis is increased in the majority of MS patients whether they have active or inactive disease, it was not the purpose of this study to determine the amount of total CSF IgG with specificity for MBP. It has been suggested that some of the intrathecal IgG is "nonsense antibody" without significance for MS. On the other hand, Panitch et al using a radioimmunoassay with guinea pig MBP, reported low anti-
MBP levels in MS patients with exacerbations. While several other authors have found antibodies against MBP in MS CSF, there have also been reported negative results. In this report we have demonstrated that autoantibodies to MBP are intrathecally produced in MS patients with active disease and that their presence correlates with disease activity. Conversely, anti-MBP was not present in MS patients in remission. Highest levels detected in patients with chronic progressive disease may be due to prolonged release of MBP into their CSF and to the spatial dissemination of the pathology within the CNS. Longitudinal case studies of patients with chronic progressive disease from our Multiple Sclerosis Patient Care and Research Clinic have shown CSF anti-MBP consistently present in high titers. Clinically active MS whether in the form of acute exacerbations or chronic progressive disease is characterized by the presence of both free circulating and bound anti-MBP antibodies.

Analysis of the CSF prior to acid hydrolysis detected free antibody in most patients with both forms of clinically active disease. However, the relative quantities of free and bound anti-MBP in different MS clinical subgroups has yet to be determined. Increased anti-MBP post hydrolysis values indicate that this antibody may be bound to proteins such as MBP or anti-idiotypes.

Intrathecally synthesized IgG which does not have specificity for MBP may be directed against other oligodendrocyte or myelin proteins or viral antigens, or they may have a protective role as anti-idiotypic antibodies or be involved in immunoregulation. Although anti-MBP antibodies may be increased in neurological diseases other than MS, such as SSPE and PIE, this does not negate the possibility that they may play a role in the pathogenesis of this disease. Whereas diseases other than MS with elevated anti-MBP may be associated with normal immunoregulation of the antibody, MS may be due to altered regulation of anti-MBP. Analyses of CSF from MS patients should include assays to detect MBP and anti-MBP as well as estimates of intrathecal IgG synthesis and oligoclonal banding.

ACKNOWLEDGEMENTS

This research was supported in part by the Multiple Sclerosis Society of Canada (Alberta Division), the Friends of the University of Alberta Multiple Sclerosis Research Clinic, the Tegler Foundation of Edmonton, Alberta, Canada, and a special contribution from the Medical Research Council. Ms. J. Christopherson, Ms. P. Shaw and Ms. B. L. Morris assisted with the clinical care of patients with progressive disease and Ms. V. Jeffrey provided excellent technical support. We are grateful for the support and encouragement of Dr. Harold Jacobs during the performance of this research.

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