Folate has recently been proposed as a new antioxidant. Folate supplementation may have a protective effect in counteracting oxidant-induced apoptotic damage. The present studies were undertaken to examine whether there is a direct link between folate levels, antioxidant capability and reduced apoptotic damage. Using an in vitro cellular model of 7-ketocholesterol (KC)-induced apoptosis, U937 cells were pre-cultured with a folate-deficient medium supplemented with various levels of folate (2–1500 μmol/l) before treatment with 7-KC. Apoptotic markers, mitochondria-associated death signals and levels of reactive oxygen species were assayed. After treatment with 7-KC for 30 h, low and high levels of folate supplementation significantly (P<0.05) reduced nuclear DNA loss. Only high levels of folate supplementation (>1000 μmol/l) were effective in counteracting 7-KC-promoted apoptotic membrane phosphatidylserine exposure and DNA laddering. The attenuation of 7-KC-induced apoptotic damage by high-dose folate supplementation coincided with a partial normalization of mitochondria membrane potential dissipation, a suppression of cytochrome c release and an inhibition of procaspase 3 activation. The prevention of mitochondrial dysfunctions and apoptotic processes was associated with antioxidant actions of high-dose folate supplementation alleviates oxidative stress, mitochondria-associated death signalling and apoptosis induced by 7-KC. However, the in vivo relevance is not clear and requires further study.

Folate supplementation: Antioxidant: Apoptosis: Mitochondria-associated death signalling: 7-Ketocholesterol

It has recently been proposed that folate scavenges peroxyl radicals, azide radicals and hydroxyl radicals in an in vitro radical reaction model system (Joshi et al. 2001). However, the antioxidant action of folate against cellular damage is not fully understood. Huang et al. (2001) reported that folate depletion accompanied by elevated homocysteine concentrations was correlated with increased lipid peroxidation in rat livers. Numerous investigations have shown that folate deficiency promoted H₂O₂ generation and lipid peroxidation in HepG2 cells (Chen et al. 2001; Chern et al. 2001). In human subjects, increased folate intake was found to improve endothelial function in patients with coronary artery disease (Doshi et al. 2001). Reduction of intracellular endothelial superoxide may have contributed to the effect. These studies raise the possibility that folate supplementation may protect cells against oxidative damage, yet a direct link has not been established.

Generation of reactive oxygen species can activate apoptosis, a programmed cell death (Palmer & Paulson, 1997; Chandra et al. 2000). Apoptotic damage in cells is characterized by chromatin condensation, nucleosomal DNA fragmentation and membrane phosphatidylserine translocation (Kroemer et al. 1995; Granville et al. 1998). Although the mechanisms by which reactive oxygen species induce apoptosis are not fully defined, it has been suggested that mitochondria play a central role (Kroemer et al. 1997; Bernardi et al. 2001). In the oxidant-promoted apoptosis, the depolarization of mitochondrial membrane potential has been attributed to the mitochondrial release of pro-apoptotic proteins such as cytochrome c, which activates the apoptotic executor of caspase 3 to initiate apoptotic DNA damage or membrane disorder (Li et al. 1997; Zamzami & Kroemer, 2001; Lim et al. 2002). These oxidative apoptotic insults may play a pivotal role in atherosclerosis, because appreciable amounts of apoptotic cells are evident in human atheromatous lesions (Han et al. 1995; Isner et al. 1995). 7-Ketocholesterol (KC), one of the prominently oxidized cholesterols and the most abundantly formed oxysterol in human atherosclerotic plaque (Brown et al. 1997; Garcia-Cruet et al. 1999), has been shown to induce apoptosis in endothelial cells (Lizard et al. 1997, 1999), smooth muscle cells (Nishio et al. 1996; Nishio & Watanabe, 1996) and monocytic cells (Aupeix et al. 1995; O’Callaghan et al. 1999). The loss of mitochondrial transmembrane potential and the cytosolic release of cytochrome c occurred during 7-KC-induced apoptosis (Lizard et al. 2000). Antioxidants such as glutathione and vitamin E can prevent 7-KC-induced apoptosis.

Abbreviations: FD, folate-deficient; KC, ketocholesterol; PI, propidium iodide.

* Corresponding author: Dr R.-F. S. Huang, fax +86 2 29021215, email rweifen@mails.fju.edu.tw
by the disruption of mitochondrial transmembrane potential and cytochrome c release (Lizard et al. 2000).

Folate deficiency was shown previously to induce apoptotic damage in several cell lines (James et al. 1994; Koury et al. 1997; Huang et al. 1999) and in animals (James et al. 1997; Craciunescu et al. 2004). The underlying mechanisms relating to the antioxidant capability of folate and its role in the signalling of mitochondria-associated apoptotic death are largely unknown. 7-KC-induced apoptosis provides us with a unique model to investigate whether there is a direct link between folate levels, antioxidant capability, mitochondria-associated death signalling and reduced apoptotic damage. U937 cells were chosen as an in vitro cellular model, because it is one of the most commonly used cell lines to study the molecular mechanisms of oxysterol-induced apoptosis (Lizard et al. 1998, 2000). U937 cells are also frequently used as macrophage reference models that are sensitive to oxysterols toxicity in the same range of concentrations observed in other vascular cells (Lizard et al. 1996, 1997, 1998). We added various levels of folate to folate-deficient (FD) medium and measured apoptotic variables, mitochondrial transmembrane potential, cytochrome c release, procaspase 3 activation and intracellular reactive oxygen species levels in U937 cells in the presence or absence of 7-KC.

Materials and methods

Materials

7-KC, folate (pteroylmonoglutamic acid), rodamine 123, propidium iodide (PI), 2',7'-dichlorofluorescin diacetate and hydroethidine were purchased from Sigma Chemical Co. (St Louis, MO, USA). Fetal bovine serum was purchased from HyClone Laboratories (Logan, UT, USA). Complete FD Roswell Park Memorial Institute 1640 medium and antibiotics were purchased from Gibco Laboratories (Grand Island, NY, USA). The rabbit antihuman CPP32 and cytochrome c antibody were from BD Biosciences (San Jose, CA, USA) and R&D Systems (Minneapolis, MN, USA). Multimers of 100 bp DNA were from Pharmacia Biotech (Piscataway, NJ, USA).

Cell culture

U937 cells, a human promonocytic leukaemia cell line, were purchased from the cell bank of NRIH (Taipei, Taiwan; CCRC 60027). U937 cells were regularly maintained in complete Roswell Park Memorial Institute 1640 medium supplemented with heat-inactivated fetal bovine serum (100 ml/l) and antibiotics (200 U penicillin/l and 20 mg streptomycin/l). Cells were maintained in suspension cultures in a humidified 5% CO₂ atmosphere at 37°C. Cellular viability was determined by Trypan Blue exclusion.

Experimental treatments

For all experiments, a 7-KC stock solution was prepared according to the methods of Lizard et al. (2000). Briefly, 800 μg 7-KC was dissolved in 50 μl absolute ethanol and 950 μl FD medium was added. The solution was then sonicated. The ethanol level in culture medium of final concentration 40 μg 7-KC/ml had no effect on cell viability when it was measured with Trypan Blue. A stock solution of folate was prepared at 10 mmol/l by dissolving 44 mg folate in 10 ml NaHCO₃ (10 g/l) solution. To make up media with various levels of folate, folate stock was added to FD medium and the pH was adjusted to 7.4 before sterilization by filtration. All stock solutions were freshly prepared before experiments.

The doubling time for U937 cells is 26 ± 2 h. Viability and growth rate of cells were frequently checked using Trypan Blue exclusion. Cells in the log growth phase were sub-cultured into various sets of plates for conducting each experiment. U937 cells were pre-incubated in FD medium with various levels of folate for 6 h and then treated with 7-KC to induce apoptosis. Cultures without 7-KC treatment at the designated experimental time points were used as additional controls. After the treatments were terminated, cells were harvested for the assay.

Analysis of apoptotic cells with membrane phosphatidylserine exposure

The Annexin-V-Fluos kit (Roche Diagnostics, Mannheim, Germany) was used to measure the apoptotic cells with membrane phosphatidylserine translocation. Cells were suspended in incubation buffer that contained 1 mg PI/l and a 1:50 dilution of Annexin-V-Fluos labelling solution. The mixed solution was incubated for 15 min on ice, and green Annexin-V and red PI fluorescence intensities were analysed on a Coulter EPICS XL-MCL flow cytometer (Coulter Corporation, Miami, FL, USA). Cells with Annexin-V-positive and PI-negative fluorescence were defined as apoptotic cells. Cells with Annexin-V-negative and PI-positive fluorescence (PI permeable) were defined as necrotic cells.

Analysis of apoptotic cells with hypodiploid DNA contents

Cells were fixed in ice-cold ethanol. RNase A (500 mg/l) and Triton (5 g/l) were added to samples at 37°C for further incubation for 60 min. Cells were then incubated with PI (50 mg/l) for 20 min at 37°C. After centrifugation, cellular DNA (red fluorescence) in 10 000 cells was analysed on a flow cytometer with the System II software program (Coulter Corporation). The percentage of cells in different cell cycle phases was estimated from PI histograms using WinMDI 2.8 software (Coulter Corporation). Hypodiploid cells, i.e. those with lower G0/G1 DNA contents, were defined as apoptotic cells, as described by Endresen et al. (1994).

Agarose electrophoresis of DNA fragmentation

As described by Huang et al. (2002), 3 × 10⁶ cells at each indicated time point were harvested and suspended in ice-cold lysis buffer (100 mmol NaCl/l, 10 mmol Tris-HCl/l, 25 mmol EDTA/l, 5 g SDS/l and 0.3 g proteinase K/l) for 15 h in a water-bath at 50°C. The standard phenol–chloroform–isoamyl alcohol (25:24:1, by vol.) method was used to remove protein and extract DNA. RNA was digested with 1 mg ribonuclease A/l for 1 h at 37°C. DNA fragments

References

The references are not included in the natural text representation.
were electrophoresed on an agarose (20 g/l) mini-gel at 100 V for 40 min and visualized with ethidium bromide staining under UV illumination.

**Quantification of depolarized cells with mitochondrial membrane potential dissipation**

Rodamine 123 has been widely used as a flow cytometric probe for mitochondrial membrane potential (Juan et al. 1994). Rodamine 123 was added directly to the culture medium (5 mg/l) and incubated for 30 min at 37°C. After centrifugation, cells were analysed with a flow cytometer (excitation 488 nm, emission 625 nm). Dead cells and debris were excluded from the analysis by electronic gating of forward- and side-scatter measurement.

**Determination of intracellular reactive oxygen species levels**

Intracellular reactive oxygen species were assayed using the fluorescent dyes 2',7'-dichlorofluorescin diacetate and hydroethidine (Robinson et al. 1994). Intracellular H$_2$O$_2$ or NO oxidizes 2',7'-dichlorofluorescin to the highly fluorescent compound. Hydroethidine oxidation is particularly sensitive to superoxide anions. Thirty minutes before 7-KC treatment was terminated, 2',7'-dichlorofluorescin diacetate (5 µmol/l) or hydroethidine (5 µmol/l) was added into cultured cells. At each indicated time point, cells were harvested and the fluorescence intensity of intracellular dichlorofluorescin (emission 530 nm) or hydroethidine (emission 585 nm) was monitored on a flow cytometer (Coulter Corporation).

**Subcellular fractionation and Western blot analysis**

Cytochrome c release into the cytosol and degradation of procaspase 3 were investigated by Western blot analysis. The analysis of cytochrome c release to cytosol was performed according to the method of Lizard et al. (2000). In brief, cells were suspended in lysis buffer (20 mmol HEPES–KOH/l (pH 7.5), 10 mmol KCl/l, 1.5 mmol MgCl$_2$/l, 1 mmol EDTA/l, 1 mmol sodium ethylene glycol tetraacetic acid/l, 1 mmol dithiothreitol/l, 250 mmol sucrose/l and a mixture of protease inhibitors) and were homogenized by successive passages through a 27-gauge fine needle. By centrifuging the homogenate at 1000 g for 10 min, the resulting supernatant fraction was subjected to 15 500 g centrifugation at 4°C for 30 min to generate cytosol. For analysis of procaspase 3 (CPP32), cells were lysed in lysis buffer (50 mmol Tris–HCl/l (pH 8.0), 150 mmol NaCl/l, 1 g SDS/l, 1 mmol sodium ethylene glycol tetraacetic acid/l, 1 mmol dithiothreitol/l, 250 mmol sucrose/l and a mixture of protease inhibitors) for 30 min on ice and then centrifuged at 4°C (14 000 g for 15 min). The protein concentration was measured by Bio-Rad assay (Bio-Rad Laboratories, Hercules, CA, USA) with bovine serum albumin as the standard. Proteins (50 μg) were incubated in loading buffer, separated by SDS–PAGE, and electroblotted to a polyvinylidene difluoride membrane. Proteins were probed with human monoclonal antibody directed against cytochrome c and caspase 3 (Santa Cruz, CA, USA), followed by peroxidase-labelled anti-mouse antibodies and visualized using an enhanced chemiluminescence detection kit (Amersham International Amersham, Bucks., UK) by autoradiography.

**Statistical analysis**

All results were presented as mean values and standard deviations. One-way ANOVA and Duncan’s test were used for comparisons among folate-supplemented groups. Statistical significance was set at P<0.05.

**Results**

Dose effects of folate supplementation on 7-ketocholesterol-induced apoptosis

Previous studies have demonstrated that treatment with 40 μg 7-KC/ml can induce apoptosis in the U937 cellular model (Lizard et al. 1998, 2000). We measured the dose effects of folate supplementation against apoptotic damage as a function of time after treatment with 40 μg 7-KC/ml. The results are shown in Table 1. In the absence of 7-KC treatment, cells cultured in FD medium did not display apoptosis within the experimental time periods. After 24 h of treatment with 7-KC, the apoptotic cells with hypodiploid DNA content were increased to 18%. Folate supplementation at 100–1500 µmol/l significantly reduced the hypodiploid cellular fractions, with the greatest magnitude of reduction with high-folate supplementation. After 30 h of treatment with 7-KC, pre-incubation of 7-KC-treated cells with media that contained a normal folate level (2 µmol/l) and high folate levels (1000 and 1500 µmol/l) significantly reduced (P<0.05) the hypodiploid cellular fractions (%). Folate at 1500 µmol/l markedly diminished 7-KC-promoted hypodiploid cellular fractions by half. The fraction of apoptotic cells with membrane phosphatidylserine translocation (Annexin-V-positive and PI-negative) was 25% after 24 h of 7-KC treatment, and this increased to 49% after 30 h of treatment with 7-KC. Folate supplementation (2–1500 µmol/l) did not affect 7-KC-induced apoptotic membrane disorder at 24 h, whereas supplementation with 1500 µmol folate/l significantly reduced (P<0.05) Annexin-V-positive cells (by half) after 30 h of 7-KC treatment.

The protective effects of folate supplements against apoptotic DNA damage were further confirmed by the results of electrophoretic analysis (Fig. 1). After 30 h of 7-KC treatment, apoptotic DNA laddering was seen (lane 3). Folate supplements at 1000–2000 µmol/l (lanes 6–8), but not at 2–100 µmol/l (lanes 4 and 5), substantially decreased the 7-KC-induced DNA fragmentation. In the absence of 7-KC treatment, no DNA fragmentation was detected in cells cultured with complete or FD medium (lanes 1 and 2).

Effects of folate supplementation on 7-ketocholesterol-induced mitochondria membrane potential dissipation

To understand whether the impairment of 7-KC-induced apoptosis by folate might be associated with changes in
Effects of folate supplementation on the percentages of apoptotic cells in the presence or absence of 7-ketocholesterol treatment*

Table 1. (Mean values and standard deviations of three independent experiments in triplicate)

<table>
<thead>
<tr>
<th>Culture time (h)</th>
<th>Cells with hypodiploid DNA contents (%)</th>
<th>Annexin-V-positive (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>FD</td>
<td>FD + 7-KC</td>
</tr>
<tr>
<td></td>
<td>Mean ± SD</td>
<td>Mean ± SD</td>
</tr>
<tr>
<td>24</td>
<td>0.8 ± 0.1</td>
<td>18 ± 3</td>
</tr>
<tr>
<td>30</td>
<td>0.9 ± 0.1</td>
<td>19 ± 1</td>
</tr>
<tr>
<td>24</td>
<td>7 ± 0.1</td>
<td>25 ± 6</td>
</tr>
<tr>
<td>30</td>
<td>0.6 ± 0.1</td>
<td>49 ± 3</td>
</tr>
<tr>
<td>24</td>
<td>2 ± 100 µmol/l</td>
<td>15 ± 3</td>
</tr>
<tr>
<td>30</td>
<td>1500 µmol/l</td>
<td>14 ± 1</td>
</tr>
<tr>
<td>24</td>
<td>1000 µmol/l</td>
<td>11 ± 1</td>
</tr>
<tr>
<td>30</td>
<td>1500 µmol/l</td>
<td>12 ± 1</td>
</tr>
<tr>
<td>24</td>
<td>2000 µmol/l</td>
<td>12 ± 1</td>
</tr>
</tbody>
</table>

FD, folate deficient; KC, ketocholesterol.

Effects of folate supplementation on mitochondria-associated signalling

The apoptotic signalling proteins of cytochrome c release from mitochondria into the cytosol and the cleavage of the inactive precursor (CPP32) were assessed by Western blot analysis. The results are presented in Figs. 3 and 4. After treatment with 7-KC for 24 h, a release of cytochrome c to cytosol was apparent (Fig. 3(B), lane 2), whereas no CPP32 cleavage was observed (Fig. 3(A), lane 2). Folate supplementation at 1500 µmol/l inhibited the 7-KC-promoted cytochrome c release (Fig. 3(B), lane 5). This cytochrome c release in 7-KC-treated cells was followed by the activation of CPP32 after 30 h of treatment with 7-KC (Fig. 3(A), lane 6). Folate supplementation at 1000 and 1500 µmol/l partially suppressed CPP32 cleavage (Fig. 3(A), lanes 7 and 8). The protective effects of high-dose folate against cytochrome c release and caspase 3 activation persisted to 36 h of 7-KC treatment, as shown in Fig. 4 ((A) and (B), lanes 4 and 5).

Effects of folate supplementation on 7-ketocholesterol-induced reactive oxygen species production

Table 2 presents results on superoxide anion scavenging by folate supplementation. Compared with 7-KC-untreated cells cultured in FD medium, folate supplementation at 2, 100, 1000 and 1500 µmol/l significantly reduced intracellular superoxide levels to 81, 75, 56 and 37 %. After 7-KC treatment for 12, 24 and 30 h, the intracellular superoxide levels of FD cells increased to 2-fold of those in 7-KC-untreated cells. Folate supplementation at 1500 µmol/l had a significantly greater (P<0.05) capability to scavenge 7-KC-elicited superoxide generation (a 42 % reduction).
compared with folate supplementation at 2 μmol/l (an 18% reduction) after 30 h of treatment with 7-KC. 7-KC treatment and folate supplementation did not affect intracellular H₂O₂ generation as measured by dichlorofluorescin fluorescence (results not shown).

Discussion

Our present results confirm previous findings that 7-KC promotes apoptosis in U937 cells (Lizard et al. 1998, 2000; O’Callaghan et al. 2001). Treatment with 7-KC caused a concomitant loss in mitochondria membrane potential, release of cytochrome c, and subsequently, the activation of apoptotic executor caspase 3, which was in agreement with the proposed mechanisms in the transmission of death signalling (Lizard et al. 2000). In the absence of 7-KC treatment, however, no apoptotic damage was observed in cells cultured in FD medium during the experimental periods. This is in contrast to previous evidence, which linked folate deficiency to apoptotic promotion in Chinese hamster ovary cells (James et al. 1994), HepG2 cells (Huang et al. 1999) and erythroblasts (Koury et al. 1997). Different cell types and the duration of folate deficiency may be attributable to these observed discrepancies. A longer period of folate deficiency (days or weeks) was frequently used to induce apoptosis, whereas our experimental time period for U937 cells was about 12–36 h.

The present results demonstrate that in vitro folate supplementation protects against 7-KC-triggered apoptotic damage. Supplemented folate at a culture medium dose (2 μmol/l) was sufficient to diminish hypodiploid cellular fractions on 7-KC treatment, whereas only high-dose supplementation was able to ameliorate the apoptotic membrane disorder and DNA fragmentation through suppressing mitochondria-associated death signalling. These results suggest that folate supplementation counteracted the 7-KC-induced apoptotic damage at various intracellular targets and molecular levels. In early literature, the proposed links for folate and apoptosis included decreased thymidylate synthesis (Koury et al. 1997), deoxynucleotide pool imbalance (James et al. 1994, 1997), DNA mismatch repair (Gu et al. 2002), elevated homocysteine levels (Duan et al. 2002) and cell cycle-dependent processes (Huang et al. 1999). We found that folate may act as a modulator of mitochondria-associated death signalling to alleviate apoptotic damage, which, to our knowledge, has not been demonstrated previously.

The molecular mechanisms by which high-dose folate could suppress mitochondria-associated death signalling are largely unknown. One of the potential protective mechanisms could be radical scavenging. Reactive oxygen species, such as superoxide and H₂O₂, have been reported to be important regulators of mitochondria membrane potential and cytochrome c release in several kinds of
oxidative stress-promoted apoptosis (Cai & Jones, 1998; Stridh et al. 1998; Tada-Oikawa et al. 1999). By scavenging superoxide, antioxidants such as vitamin E and N-acetylcysteine could reverse mitochondria membrane depolarization and cytochrome c release to the cytosol in 7-KC-treated U937 cells (Lizard et al. 2000). Indeed, we found that high-dose folate supplementation significantly reduced intracellular superoxide levels during the early phase of 7-KC treatment, which coincided with the concomitant reverse of mitochondria dysfunction and apoptotic damage. These results suggest that folate antioxidant capability, particularly for superoxide scavenging, as has been proposed by previous investigators (Doshi et al. 2001; Huang et al. 2002), can partially account for the impairment in mitochondria-associated death signals.

The anti-apoptotic function by folate may not be limited to its antioxidant property. High-dose folate supplementation may enrich the mitochondria folate pool to strengthen respiratory function, the impairment of which leads to superoxide production and abnormal mitochondrial permeability transition during apoptosis (Cai & Jones, 1998; Crompton, 1999). This postulation is partially supported by the finding that a lipophilic folate antagonist, at high concentrations, can be accumulated in mitochondria to act as a respiration inhibitor (Sprecher et al. 1995). Alternatively, the suppression of mitochondrial cytochrome c release by folate may be attributable to a downstream effect of folate metabolites through mitochondrial C1 metabolism. 10-Formyl-tetrahydrofolate, a major mitochondria folate coenzyme (Horne

![Fig. 3. Modulation of cytochrome c release and procaspase 3 activation by folate supplementation. The cleavage of caspase 3 precursor (CPP32, (A)), the release of cytochrome c into the cytosol (B) and β-actin (C) were investigated by Western blot analysis. U937 cells were pre-incubated in folate-deficient medium supplemented with various levels of folate for 6 h before treatment with 7-KC. For details of procedures, see p. 889. Lane 1: cells not treated with 7-ketocholesterol (KC). Lanes 2–5: cells treated with 7-KC for 24 h, and supplemented with folate at concentrations of 0 (lane 2), 2 (lane 3), 1000 (lane 4), and 1500 (lane 5) μmol/l. Lanes 6–7: cells treated with 7-KC for 30 h, and supplemented with folate at concentrations of 0 (lane 6), 1000 (lane 7), and 1500 (lane 8) μmol/l.](image)

![Fig. 4. Effects of folate supplementation on procaspase 3 activation (A), cytochrome c release (B) and β-actin (C) after 36 h of treatment with 7-ketocholesterol (KC). Folate pre-incubation is the same as described in Fig. 3. For further details of procedures, see p. 889. Lane 1: cells not treated with 7-KC. Lanes 2–5: cells treated with 7-KC and supplemented with folate at concentrations of 0 (lane 2), 2 (lane 3), 1500 (lane 4) and 2000 (lane 5) μmol/l.](image)

Table 2. Effects of folate supplementation on superoxide production in the presence or absence of 7-ketocholesterol treatment* (Values are means and standard deviations for three independent experiments in triplicate)

<table>
<thead>
<tr>
<th>Culture time (h)</th>
<th>Treatment</th>
<th>FD</th>
<th>FD + folate supplementation (μmol/l)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>2</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td></td>
<td>1000</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>1500</td>
<td></td>
</tr>
<tr>
<td>HE fluorescent intensity</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>12</td>
<td>− 7-KC</td>
<td>16a</td>
<td>2 100</td>
</tr>
<tr>
<td></td>
<td>+7-KC</td>
<td>33a</td>
<td>6 100</td>
</tr>
<tr>
<td></td>
<td>+7-KC</td>
<td>34a</td>
<td>3 100</td>
</tr>
<tr>
<td></td>
<td>+7-KC</td>
<td>39a</td>
<td>1 100</td>
</tr>
<tr>
<td>24</td>
<td>− 7-KC</td>
<td>13b</td>
<td>1 81</td>
</tr>
<tr>
<td></td>
<td>+7-KC</td>
<td>35b</td>
<td>7 106</td>
</tr>
<tr>
<td></td>
<td>+7-KC</td>
<td>27b</td>
<td>5 79</td>
</tr>
<tr>
<td></td>
<td>+7-KC</td>
<td>32b</td>
<td>5 82</td>
</tr>
<tr>
<td>30</td>
<td>− 7-KC</td>
<td>9c</td>
<td>1 56</td>
</tr>
<tr>
<td></td>
<td>+7-KC</td>
<td>26c</td>
<td>3 78</td>
</tr>
<tr>
<td></td>
<td>+7-KC</td>
<td>26c</td>
<td>3 73</td>
</tr>
<tr>
<td></td>
<td>+7-KC</td>
<td>26c</td>
<td>1 66</td>
</tr>
<tr>
<td>a,b,c,d Mean values within a row with unlike superscript letters were significantly different (P&lt;0.05).</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>* U937 cells were preincubated with FD-medium supplemented with 2, 100, 1000 and 1500 μmol/l folate/l for 6 h before treatment with 7-KC (40 μg/l). At the indicated time points, cells were labelled by HE fluorescent dye for 30 min, harvested, and analysed by flow cytometer. For details of procedures, see p. 889.</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>† Percentage of FD value.</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Folate and 7-ketocholesterol-induced apoptosis

et al. 1989), has been shown to support the respiration of rat mitochondria by reducing cytochrome c at the site of respiratory complex IV enzymes (Brookes & Baggott, 2002). The altered redox state of cytochrome c could modulate its release to the cytosol, and its activation to trigger programmed cell death (Calver et al. 1997; Hancock et al. 2001; Lim et al. 2002). More studies are warranted to elucidate the regulatory role of folate on mitochondria-associated death signalling.

In the current investigation, the effective dose of folate to counteract mitochondria dysfunction was found at mM levels, which are much greater than those encountered in the human physiological state. It is open to debate whether in vitro experiments are meaningful in the context of the whole organism. However, it is also generally accepted that metabolic observations in cultured cell lines may not be directly applicable to the multicellular environment. Supraphysiological levels of treatment proved to be necessary to achieve the potential effects on cells in vitro over relatively short experimental time intervals (hours in the present study). We cannot exclude the possibility that, over a longer time course (weeks or months), high dietary folate supplementation could be sufficient to protect against mitochondria dysfunction in vivo. This possibility is supported by the observation that rats fed a high-folate diet had significantly lower percentages of depolarized cells and mitochondrial protein oxidative damage in their livers compared with rats fed a FD diet (Chang, 2004). In human subjects, folic acid supplements at a high dose of 5 mg/d (more than 10-fold the daily recommended value) for 6 weeks or 4 months significantly improved endothelial dysfunction in patients with coronary artery disease (Title et al. 2000; Doshi et al. 2001). Clinical intervention trials on folate and cancer have further revealed that supplementation with a pharmacological dose of folate (5–20 mg) may have a chemoprotective role (Kim, 1999). Although the mechanisms of long-duration high dietary folate supplementation to protect against human disease are far from understood, our present observations from an in vitro cellular model may provide potential insights into the possible role of high-dose folate in reducing oxidative stress and mitochondria dysfunction.

Given the assumptions and limitations of in vitro studies, the results in the present report suggest that folate supplementation exerts differentially protective effects against 7-KC-induced damage. High-dose supplementation alleviates oxidative stress, mitochondria-associated death signalling and apoptosis induced by 7-KC. However, the in vivo relevance is not clear and requires further study.

Acknowledgements

The authors acknowledge Dr Y.-A. Lee, Department of Life Science, Fu-Jen University, for providing UV illumination and Dr N. S. Shaw, Department of Biotechnology, National Taiwan University, for technical assistance in Western blotting. This study was funded by a grant from the National Science Council, Taiwan, Republic of China (NSC 92-2320-B-030-002).

References


Chang CM (2004), Effects of folate supplementation on mitochondrial DNA deletion, function and oxidative damage in rat liver, MSc Thesis, Fu-Jen University, Taiwan.


Hancock JT, Desikan R & Neill SJ (2001) Does the redox


