The Genetical Relationship of Impulsiveness and Sensation Seeking to Eysenck's Personality Dimensions

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The genetical analysis of covariance structures is used to explore the genetical and environmental intercorrelations of impulsiveness and sensation seeking factors and their conformity to Eysenck's principal personality dimensions. The independent dimensions of psychoticism, extraversion, neuroticism, and lie scale are not found to give a very satisfactory account of the genetical factor structure. In particular, it is clear that impulsiveness and sensation seeking are not simple reflections of extraversion.

Key words: Genetic analysis, Covariance structures, Impulsiveness, Sensation seeking, Extraversion, Eysenck Personality Questionnaire, Twin study

INTRODUCTION

For many years Eysenck [8] has argued that the main features of individual differences in personality can be explained with reference to three independent high-order factors: psychoticism (P), extraversion (E), and neuroticism (N). These three factors are always extracted from administrations of the Eysenck Personality Questionnaire (EPQ) (and similar ones are extracted from other personality scales), along with a further factor -- the lie scale (L), which seems to be a measure of dissimulation or social desirability but whose significance is less clear than the three major factors. Some authors have considered that this scheme is too coarse-grained and ignores many facets of personality that really provide the interesting contrasts between people. Thus Guildford [13] concluded that E was a kind of "shotgun wedding" between rhathymia (akin to impulsiveness) and sociability. Eaves and Eysenck [3] tested 837 twin pairs with scales of sociability and impulsiveness and showed, indeed, that the subjects × scales interaction had a significant genetical component suggesting "some justification for regarding Sociability and Impulsiveness as distinguishable genetically". However, they estimated the genetical corre-
lation between the two as 0.42 and the environmental correlation to be 0.66, supporting Eaves' earlier view [2] that the unitary nature of extraversion is due more to environmental than to genetical influences.

Various authors have suggested that impulsiveness itself is a combination of subfactors that would more usefully be considered separately. Eysenck and Eysenck [9] have factor analysed a set of items related to impulsiveness (in the broad sense) into four primary factors, which they call impulsiveness in the narrow sense (IMPN), risk taking (RISK), non-planning (NONP), and liveliness (LIVE), and have related these to P, E, N, and L. Eaves et al [7] estimated the common factor and specific components of the variation in these four traits and showed that the same factor structure was operating for both genetical and environmental sources of variation.

Zuckerman [24] has demonstrated that sensation seeking is an important aspect of personality through which the individual regulates his degree of arousal. Consequently one would expect it to be related to extraversion, which is thought to vary with the same physiological function [8]. However, Eysenck and Zuckerman [10] have shown that the four subscales of sensation seeking have different phenotypic correlations with P, E, N, and L. These four subfactors are Disinhibition (DIS), thrill and adventure seeking (TAS), experience seeking (ES), and boredom susceptibility (BS), and the genetical and environmental covariance among these four factors has been explored by Fulker et al [12].

The present paper takes advantage of the fact that scores for the four impulsiveness factors, the four sensation seeking factors and P, E, N, and L were obtained for samples from the Maudsley Twin Register in 1975. This provides the opportunity to examine the genetical and environmental causes of covariation among the 12 variables using the genetical analysis of covariance structures approach of Martin and Eaves [19].

The genetical analysis of covariance structures, based on the work of Jöreskog [15], allows the research worker to test models of covariation incorporating various genetical and environmental sources of covariation and different factor structures through which these sources influence the traits in question. In our case, with 12 variables, there are many models one might fit to the data, including a great variety of empirical factor structures. This approach risks the accusation of "looking for a model that fits", so we shall restrict our hypotheses to those that attempt to relate the covariation among the eight factors of impulsiveness and sensation seeking to Eysenck's four principal dimensions of personality, P, E, N, and L.

THE DATA

The three self-report questionnaires were sent by post to twins from the Maudsley Twin Register during 1975, the impulsiveness questionnaire and EPQ together, and the sensation seeking questionnaire (SSQ) on a separate occasion. Details of the impulsiveness and EPQ scales are given by Eysenck and Eysenck [9] and Eaves et al [7]; those of the sensation seeking questionnaire, by Fulker et al [12]. The number of items in each is: IMPN (12), RISK (10), NONP (12), LIVE (6), P (25), E (21), N (23), L (21), DIS (10), TAS (10), ES (10), and BS (10). Whereas 588 pairs replied to the first two questionnaires, only 441 pairs replied to the SSQ, leaving an intersect of 438 pairs whose sex and zygosity distribution is shown in Table 1. Zygosity determination in the Maudsley Twin Register is discussed by Kasriel and Eaves [17]. The age range of respondents was 16 to 73 years, with a mean of 31 years.

An angular transformation was applied to the raw scores for each factor to improve the additive properties of the scales. Between- and within-mean products matrices were calculated for all five twin groups, providing ten matrices in all. The between-mean products matrices were corrected for linear regression in age, so reducing their degrees of freedom by one. The within-mean products matrix for DZ opposite-sex pairs was corrected for the mean difference between males and females, thus reducing its degrees of freedom by one. The ten 12 x 12 matrices, corrected for age and sex, are reproduced in the Appendix.
THE METHOD

The genetical analysis of covariance structures was adapted from the work of Jöreskog [eg, 15] on confirmatory factor analysis. Its development is discussed by Martin and Eaves [19] and further illustrated by Eaves et al [7] and Fulker [11]. Briefly, it allows one to test hypotheses about the genetical and environmental sources of variation simultaneously with psychological hypotheses about the contribution of these sources to the structure of covariation between variables and the residual variation specific to particular variables.

A simple model for the sources of variation in our data is that only individual environmental experiences \( E_i \) and additive gene action \( D_R \) need be invoked to explain variation in personality factor scores. In the univariate case these sources contribute to the mean squares between (B) and within (W) MZ and DZ pairs with the following coefficients:

\[
\begin{align*}
MS_{BMZ} &= E_i + D_R \\
MS_{WMZ} &= E_i \\
MS_{BDZ} &= E_i + \frac{1}{4} D_R \\
MS_{WDZ} &= E_i + \frac{1}{4} D_R
\end{align*}
\]

This simple model has been found adequate to explain individual differences in extraversion, neuroticism, and psychoticism [3, 4, 5, 6, 20].

We fitted this \( E_i D_R \) model to data for females and males separately for each of the 12 variables in this study. It gave an adequate explanation in all cases except ES in males \( (\chi^2 = 6.5) \), and only in that case and L for both sexes did a model incorporating \( E_i \) and a between-families environmental component \( E_2 \) give a marginally better fit.

It seems, therefore, that the multivariate extension of this simple \( E_i D_R \) model will be a good initial hypothesis for the sources of covariation, and this may be written:

\[
\begin{align*}
\Sigma_{BMZ} &= H' + E^2 + \Delta' + D^2 \\
\Sigma_{WMZ} &= H' + E^2 \\
\Sigma_{BDZ} &= H' + E^2 + \frac{1}{4} (\Delta' + D^2) \\
\Sigma_{WDZ} &= H' + E^2 + \frac{1}{4} (\Delta' + D^2)
\end{align*}
\]

where \( \Sigma_i \) is the \( i \)-th expected mean products matrix. Here \( H \) and \( \Delta \) are matrices of \( E_i \) and \( D_R \) factor loadings, respectively, and \( E^2 \) and \( D^2 \) are the corresponding diagonal matrices of specific variance components for those two sources.

In more general terms, we may write the expectation for a mean-products matrix:

\[
\Sigma_i = \sum_{j=1}^{p} c_{ij} \left[ B_j (\Lambda_j, \Phi_j, \Lambda'_j) B'_j + \Theta_j^2 \right]
\]

### TABLE 1. Sex and Zygosity Composition of the Twin Sample

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<thead>
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<th>Opposite sex</th>
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where there are $p$ sources of variation and $c_{ij}$ is the coefficient from the univariate model for the $i$-th mean square and $j$-th source. For the $j$-th source $\Delta_j$ is the matrix of factor loadings and $\Theta_j \mathbf{I}$ the diagonal matrix of specific variance components, as above. Note, however, that we may complicate the model by introducing correlations between the factors in $\Phi_j$, or relate the factor structures of different sources by a simple scalar held in the diagonal matrix $B$. We shall not employ these facilities much in this example.

Having specified the sources of variation and the factor structures of our model, how do we go about testing it? The approach is described fully by Martin and Eaves [19]. Generally, our data will consist of $k$ matrices of mean products. We may write $S_i$ for the $i$-th matrix, having $N_i$ degrees of freedom. Given some model for the $S_i$, we may compute the expected values $\Sigma_i$, being positive definite, for particular values of the parameters of the model. When the observations are multivariate normal, we may write the log likelihood of obtaining the $k$ observed independent $S_i$ as:

$$
\log L = -\frac{1}{2} \sum_{i=1}^{k} N_i \left\{ \log |\Sigma_i| + \text{tr} (S_i \Sigma_i^{-1}) \right\}
$$

(omitting the constant term).

For a given model we require the parameter estimates that maximise $\log L$. Given maximum-likelihood estimates of our parameters, we may test the hypothesis that a less restricting model (ie, one involving more parameters) does not significantly improve the fit by computing:

$$
\chi^2 = 2(L_0 - L_1),
$$

where $L_1$ is the log likelihood obtained under the restricted hypothesis ($H_1$) and $L_0$ is the log likelihood obtained under the less demanding hypothesis ($H_0$). The $H_0$ we shall adopt in practice is that which assumes that as many parameters are required to explain the data as there are independent mean squares and mean products in the first place; ie, $\Sigma_i = S_i$ for every $i$. In this case we have simply:

$$
L_0 = -\frac{1}{2} \sum_{i=1}^{k} N_i [\log |S_i| + p]
$$

When we have $k$ matrices the $\chi^2$ has $\frac{1}{2}kp (p + 1) - m$ df, where $m$ denotes the number of parameters estimated under $H_0$ and $p$ is the number of variables.

The likelihood is maximised by attempting to minimise $-\log L$ for a given model. There are many numerical methods for doing this. A variety of these methods has been implemented by the Numerical Algorithms Group (1974), and we employed the most flexible of their FORTRAN routines, E04HAF, for constrained minimisation. The routine has the advantage of allowing the user certain flexibility in the choice of method. In particular, minimisation can be based on evaluation of the function values alone (in this case the values of $-\log L$ and any functions used in specifying constraints on parameter values), or minimisation can be assisted by computation of first derivatives or of first and second derivatives of the function. Furthermore, differentiation can proceed numerically or can be programmed precisely by the user. For our problem the Powell 64 method was used, which relies only on the evaluation of the functions themselves, since coding the first derivatives was tedious, and their approximate routine was used because of the need to ensure the $\hat{\Sigma}_i$ are all positive definite. For our simple example these constraints should be automatically satisfied, providing we estimate $D$ and $E$ rather than $D^2$ and $E^2$. The problem thus reduces to an unconstrained problem in our case. However, in problems that are factorially more complex, further
General Structure of Personality Dimensions

TABLE 2. Observed Phenotypic Correlations Between Characters*

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*Females, upper triangle (620 df); males, lower triangle (214 df). Decimal points omitted.

Constraints may be required to ensure that the correlation matrix of the factor loadings is positive definite. This may be done numerically in several ways — eg, by constraining the eigenvalues of the leading minor determinants to be positive. The E04HAF routine uses a penalty function technique due to Lootsma [18] for constraining the estimates in the required region.

RESULTS

Before starting to fit models of covariation, it is interesting to inspect the observed correlations (age corrected) for males and females shown in Table 2. With some exceptions, the correlations are remarkably consistent between the sexes, and so we might expect to be able to fit the same factor structure in males and females. The intercorrelations of P, E, N, and L are low, except for the consistent negative correlation between P and L, which has been noted in other studies [eg, 20].

The first model we may wish to test postulates that the covariation can be explained by a single environmental factor and a single genetical factor, each loading on all 12 variables and with corresponding E! and D! specific components. There are thus 48 free parameters to be estimated and, when this model is fitted, it yields a log likelihood Lt = 9081.3. The log likelihood is based on the null hypothesis that there are as many free parameters as unique statistics (ie, a perfect fit), Lo = 9745.3 so x² = 2(9745.3 -9081.3) = 1328. Our data contain 780 unique statistics, and we have estimated 48 free parameters, so the chi-square has 732 degrees of freedom.

This model fits very badly indeed, but its failure is a useful benchmark against which to judge a more elaborate psychological model. A further clue to this bad failure may be found in the univariate analyses and in the original analyses of the four impulsiveness variables [7] and sensation seeking [12]. These all suggest that, for some of the characters under study, the genetical control of variation differs in males and females. Investigation of this sex limitation in the impulsiveness variable revealed that, whereas the genetical covariance could be explained by the same common factor in both sexes, the specific genetic variation was controlled by different genes in males and females. Fulker et al [12] found a similar phenomenon in their analysis of the sensation seeking data, where subscale profiles showed a different pattern of inheritance in males and females.
Clearly, we must take account of this sex limitation in our model, and we do it by estimating separate $d_j$'s for males and females. The model is that described above, except that we have slightly different expectations for the opposite-sex (OS) pairs, as follows:

$$
\Sigma_{\text{BOS}} = \frac{1}{4} \Delta \Delta' + \frac{1}{4} (D_{m}^2 + D_{f}^2) + HH' + E^2
$$

$$
\Sigma_{\text{WOS}} = \frac{1}{4} \Delta \Delta' + \frac{1}{4} (D_{m}^2 + D_{f}^2) + HH' + E^2
$$

$D_{m}^2$ and $D_{f}^2$ denote the specific additive genetical variances for males and females, respectively. In the expectations for like-sex pairs we merely substitute $D_{m}^2$ for $D^2$ in the males and $D_{f}^2$ for $D^2$ in the female pairs.

The psychological model we wish to test is that the genetical and environmental covariation between the 12 personality variables can be adequately explained within the framework of Eysenck's four principal dimensions. We therefore define four factors, one each for P, E, N, and L, and allow the eight impulsiveness and sensation seeking variables to load on them. Although experience and the data suggest that we should allow the P and L factors to be correlated, for the sake of simplicity we shall make the four factors orthogonal (i.e., we fix $\Phi$ as an identity matrix). We shall expect the four genetical factors to account for all the genetical variation and covariation of P, E, N, and L, respectively, and so we shall not allow any specific genetic variance components for these four pivotal variables. We expect, however, specific environmental variation for all variables, if only because of measurement error.

Our model thus consists of four orthogonal $E_j$ factors, each loading on nine variables (four impulsiveness, four sensation seeking, plus the "superfactor"), four $D_R$ factors in the same structure, 12 $E_j$ specific standard deviations ($\Theta_j$), and eight $D_R$ specific standard deviations (the $\Theta_j$ for P, E, N, and L are fixed to zero) each for males and females. Thus, there is a total of 100 ($36 + 36 + 12 + 8 + 8$) free parameters to be estimated.

Not surprisingly, this maximisation consumes a lot of computer time, but the log likelihood finally converged on is $L_1 = 9263.8$. The likelihood ratio test for goodness-of-fit of the model gives $\chi^2_{680} = 963$, still a very poor fit but a great improvement ($\chi^2_{52} = 365$) on the first model.

The maximum-likelihood estimates of the parameters are shown in Table 3. Ideally, we should like to attach standard errors to these estimates and perhaps discard the nonsignificant ones before refitting the model. Martin and Eaves [19] show how the covariance matrix of the estimates may be evaluated, but in this example with 100 parameters the evaluation took too long, even with the extensive computer resources at our disposal.

If the stringent application of Eysenck's superfactor model produces such a poor fit, we may ask whether a better fit can be obtained by relaxing some of the constraints of the model. If we allow P, E, N, and L to have specific genetical components (different for males and females) so that the superfactors are not required to account for all the variation and covariation in these four variables, the addition of these eight free parameters gives a fit of $\chi^2_{672} = 878$ or an improvement of $\chi^2 = 85$ over the more stringent model.

If the factor structures for environmental and genetical sources of variation are very similar, it may be possible to improve the fit of the model by constraining the $E_j$ and $D_R$ factor loadings to be related by a single constant, $b$ (which will be related to the heritability of the common factors). This was done successfully for the impulsiveness data alone by Eaves et al [7]. This modification was made to the less stringent model resulting in a saving of 35 parameters ($-36 + 1$), but this 73-parameter model gave an even worse fit, $\chi^2_{707} = 1057$, or a deterioration of $\chi^2_{55} = 179$.

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### TABLE 3. Maximum-Likelihood Estimates for Parameters of the Superfactor Model Allowing for the
Apparent Effects of Sex Limitation on Specific Genetical Variation*

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*Decimal points omitted
It is apparent that other factor structures may give a better account of the data, but none of the models tried went anywhere near an acceptable level of significance. This seems to be a problem frequently encountered but little discussed by those who work with maximum likelihood factor analysis and analysis of covariance structures. The trouble seems to be that one obtains a very sensitive test of the model, which may fail from all sorts of trivial departures not related to the hypothesis under test. Of course, the significance level is directly related to sample size, but it does seem to be common experience among workers in this area that it is often impossible to get any but the most overspecified model to fit [1, 14].

How, then, is one to judge when to stop fitting extra parameters when all such models fail on the likelihood ratio criterion? Jöreskog [16] advocates use of Tucker and Lewis' reliability coefficient [23] as a guide to the proportion of total dispersion accounted for by the model.

For our case with several matrices we may modify their argument and the computation of the reliability coefficient, $\rho$, as follows. For a given model, $\chi^2_n/n$ gives the likelihood ratio criterion for goodness-of-fit. This $\chi^2_n/n$ is $F_n/\infty$, and if the model fits, the expected value of $F$ is 1, so for a particular case $F-1$ is a measure of goodness-of-fit of the model. If we now compute $F_0$ for a "stupid" model, $H_0$ (eg, no factors, no genetic variation), and $F_1$ under a particular "sensible" model, $H_1$, then $F_0-F_1$ is a measure of "improvement" due to $H_1$. The "total improvement possible" is $F_0-1$, so the reliability coefficient

$$
\rho = \frac{F_0-F_1}{F_0-1}
$$

is the proportional improvement achieved by our model $H_1$. This is the same as Tucker and Lewis' coefficient, except that they apply a correction to improve the approximation to $\chi^2$.

The most inappropriate model imaginable for our data might consist of only 12 E1 specific standard deviations. No covariation is allowed for between variables, nor is any additive genetic variation specified to account for different MZ and DZ intrapair correlations. When this 12-parameter model is fitted it yields a $\chi^2_{768} = 3312$. Judged against this, the 48-parameter model first tested has a reliability coefficient of 0.75, the 100-parameter "superfactor" model has $\rho = 0.87$, and the less stringent 108-parameter model has $\rho = 0.91$. A coefficient of 0.95 has been suggested as an acceptable reliability for a model, and by this criterion the 100-parameter "superfactor" is inadequate.

Another criterion that may assist decision-making in this type of model fitting is Akaike's Information Criterion [see, eg, 22] $AIC = 2L_1 + 2m (m$ free parameters), which balances an increase in likelihood with the loss of parsimony involved in fitting the extra parameters to achieve this. Akaike suggests that the model with minimum AIC is the most desirable, and by this criterion we should still choose the relaxed superfactor model of 108 parameters. However, the object of this paper is to examine the conformity of the pattern of environmental and genetic covariation between the impulsiveness and sensation seeking variables with Eysenck's scheme of the principal personality dimensions, and we shall restrict our discussion to this model.

**DISCUSSION**

Several criteria suggest that the "superfactor" model for genetical and environmental covariance between the 12 personality variables shown in Table 3 is wanting. However, let us judge the seriousness of this inadequacy by closer inspection of the results.

In Table 4 we have converted the parameter estimates of Table 3 to show the percentage contribution of each of the factors and specific components to the total expected variance for each character in females. These percentages are much the same in males except in the...
TABLE 4. Percentage Contributions of Individual Environmental (E) and Additive Genetic (DG) Factor and Specific Components to Total Variation for Each Character in Females

<table>
<thead>
<tr>
<th>Character</th>
<th>IMPN</th>
<th>RISK</th>
<th>NONP</th>
<th>LIVE</th>
<th>P</th>
<th>E</th>
<th>N</th>
<th>DIS</th>
<th>TAS</th>
<th>BS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total</td>
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<td>0.0884</td>
<td>0.0504</td>
<td>0.1218</td>
<td>0.0274</td>
<td>0.0775</td>
<td>0.0715</td>
<td>0.0458</td>
<td>0.0727</td>
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</table>

**Additive genes (DG)**

<table>
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<tr>
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<th>NONP</th>
<th>LIVE</th>
<th>P</th>
<th>E</th>
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<th>DIS</th>
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<tbody>
<tr>
<td>Total</td>
<td>0.0775</td>
<td>0.0715</td>
<td>0.0727</td>
<td>0.0094</td>
<td>0.0274</td>
<td>0.0775</td>
<td>0.0715</td>
<td>0.0458</td>
<td>0.0727</td>
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</table>

**Specific (Error)**

<table>
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<tr>
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<th>LIVE</th>
<th>P</th>
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<th>BS</th>
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<td>0.0094</td>
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<td>0.0775</td>
<td>0.0715</td>
<td>0.0458</td>
<td>0.0727</td>
<td>0.0094</td>
</tr>
</tbody>
</table>

**Expected measurement error is calculated as e**
specific genetical contributions for risk (0%), nonplanning (2%), liveliness (24%), and disinhibition (18%). One possible explanation for different specific genetic contributions to variance in males and females is the very different degree of selection for the male and the female twins (83 vs 286 same-sexed pairs), and one wonders if some of the apparent male–female differences might stem from this source.

It is clear that most of the E1 variance is specific to each variable and that the “factors” are largely artificial, most loading substantially on only one variable. Thus, factor II takes out all the E1 variance for E, III for IMPN, and I for RISK. The E1 variance for DIS is divided equally between factors I and IV, and it is only factor I that has anything like the appearance of a genuine E1 common factor. Since E1 variance is, of its nature, specific, it is not surprising to find that this is an unimportant source of covariation.

Such covariation as there is appears more likely to be explained by the four genetical superfactors that have been forced to assume the dimensions of P, E, N, and L. The sum of components due to genetic factors and specific genes for any one variable is the heritability for that variable and it can be seen that most of these are rather low. Nevertheless, for all variables except RISK, over half the variation is due to common factors.

The genetical P factor accounts for much of the covariation with non-planning, whereas the E factor loads heavily on disinhibition and boredom susceptibility. The N factor appears to account for little of the genetical covariation between variables, but L, or social desirability, appears to be genetically related (inversely) to thrill and adventure seeking and experience seeking.

Because the raw data were subjected to an angular transformation each variable has an expected measurement error variance equal to \( \frac{\pi}{4} \) (where the scale has \( n \) items), provided the items are all of equal difficulty. If the items of a scale are not equally difficult, then this estimate of error will be larger than the true value. These expected measurement errors, expressed as a percentage of the expected total variation for each variable, are shown in Table 4 under the column marked “Error”. It can be seen that in many cases error accounts for a substantial part of the specific (or quasi-specific) E1 variation. Correction of “heritabilities” for this source of measurement error will increase the heritability of the reliable variance, substantially in some cases.

Although we are not particularly satisfied with the model we have set out to test, another way of viewing its adequacy is to inspect the residuals left after subtracting the predicted correlations for those observed. To do this for all ten 12 × 12 matrices is more than the mind can absorb, but inspection of the single phenotypic correlation matrix will give us some guide. The residual phenotypic correlation matrices for males and females are shown in Table 5, and most of the deviations are quite respectably small. Of the larger deviations, many if not most can be seen to involve the L factor, which suggests that we have not satisfactorily incorporated this variable in our factor structure. On the other hand, a possible clue to this anomaly may lie in the biggest deviation of Table 5, which is for the P–L correlation. Our model failed to take account of this well-known correlation, and forcing these two factors to be orthogonal may have been detrimental to the model.

However, the fundamental hypothesis we set out to test, that covariation between impulsiveness and sensation seeking variables would be largely explicable in terms of the E factor, has received only limited support. The P and L factors seem equally important, but it is also apparent that the attempt to fit these variables within the straightjacket of P, E, N, and L, is far from satisfactory.

Whatever the substantive findings of this paper, the work illustrates the value of the method in allowing us to combine the testing of structural psychological hypotheses with
### Table 5. Residual Phenotypic Correlations (Observed–Expected) *

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*Females, Upper Triangle; Males, Lower Triangle. Decimal points omitted.

A variety of genetical and environmental models of covariation. The scientific advantage of this approach over the all too common practice of “look and see” cannot be overemphasised.

**Acknowledgments.** We should like to thank Professor H. J. Eysenck for his comments on the manuscript and Professor M. Zuckerman and Dr. S. B. G. Eysenck for permission to use their data collected for the Maudsley Twin Register by Mrs. J. Kasriel. The extensive computations were carried out on the CDC 7600 at University of Manchester Regional Computing Centre. N. G. M. and L. J. E. were supported by an M. R. C. programme grant in psychogenetics.

**REFERENCES**

208 Martin, Eaves, and Fulker


Correspondence: Dr. N. G. Martin, Department of Population Biology, Australian National University, Canberra 2601, Australia.

APPENDIX

\[
\begin{align*}
\text{2A female between pairs matrix}\ 172 df. \\
\text{IB} & = 0.5717397 & 0.0387668 & 0.0140114 & 0.0247526 \text{ subject to the Cambridge Core terms of use,} \\
\text{IB} & = 0.0387668 & 0.5717397 & 0.0387668 & 0.0140114 \\
\text{IB} & = 0.0140114 & 0.0387668 & 0.5717397 & 0.0247526 \\
\text{IB} & = 0.0247526 & 0.0140114 & 0.0387668 & 0.5717397 \\
\end{align*}
\]

\[
\begin{align*}
\text{2A female within pairs matrix}\ 172 df. \\
\text{IB} & = 0.0358619 & 0.0218966 & 0.0140114 & 0.0247526 \text{ subject to the Cambridge Core terms of use,} \\
\text{IB} & = 0.0218966 & 0.0358619 & 0.0218966 & 0.0140114 \\
\text{IB} & = 0.0140114 & 0.0218966 & 0.0358619 & 0.0247526 \\
\text{IB} & = 0.0247526 & 0.0140114 & 0.0218966 & 0.0358619 \\
\end{align*}
\]

\[
\begin{align*}
\text{1B female between pairs matrix}\ 172 df. \\
\text{IB} & = 0.0434547 & 0.0191720 & 0.0102280 & 0.0208290 \text{ subject to the Cambridge Core terms of use,} \\
\text{IB} & = 0.0191720 & 0.0434547 & 0.0191720 & 0.0102280 \\
\text{IB} & = 0.0102280 & 0.0191720 & 0.0434547 & 0.0102280 \\
\text{IB} & = 0.0208290 & 0.0102280 & 0.0191720 & 0.0434547 \\
\end{align*}
\]