Effect of pomegranate seed oil on hyperlipidaemic subjects: a double-blind placebo-controlled clinical trial

Parvin Mirmiran1,2, Mohammad Reza Fazeli3, Golaleh Asghari1, Abbas Shafiee4 and Fereidoun Azizi5*

1Obesity Research Center, Research Institute for Endocrine Sciences, Shahid Beheshti University of Medical Sciences, Tehran, Iran
2Faculty of Nutrition and Food Technology, National Nutrition and Food Technology Research Institute, Shahid Beheshti University of Medical Sciences, Tehran, Iran
3Department of Drug and Food Control, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran 14174, Iran
4Pharmacutical Sciences Research Center, Tehran University of Medicinal Sciences, Tehran 14174, Iran
5Endocrine Research Center, Research Institute for Endocrine Sciences, Shahid Beheshti University of Medical Sciences, PO Box 19395-4763, Tehran, Iran

(Received 3 September 2009 – Revised 26 January 2010 – Accepted 27 January 2010 – First published online 25 March 2010)

In vivo and in vitro studies have shown that punicic acid, a type of conjugated fatty acid and the main constituent of pomegranate seed oil (PSO), has anti-atherogenic effects. The present study aimed at determining the effect of PSO treatment on serum lipid profiles. This double-blind placebo-controlled randomised clinical trial included fifty-one hyperlipidaemic subjects, diagnosed according to National Cholesterol Education Program definition, and randomly assigned to the PSO and the control groups. The PSO and placebo groups received 400 mg PSO and placebo twice daily, respectively and were followed up for 4 weeks. Serum concentrations of lipids and lipoproteins were measured before and 4 weeks after intervention. Mean concentration of TAG and the TAG:HDL cholesterol (HDL-C) ratio were significantly decreased after 4 weeks in the PSO group as compared with baseline values (2·75 (SD 1·40) v. 3·45 (SD 1·56) mmol/l, P=0·009 and 5·7 (SD 4·6) v. 7·5 (SD 5·0), P=0·031, respectively). The treatment effect was statistically significant in the PSO group as compared with controls in diminution of cholesterol:HDL-C ratio (5·4 (SD 1·5) v. 5·9 (SD 1·4), P<0·05) adjusted for baseline values. We found a mean difference for PSO v. placebo in HDL-C concentration (0·13 v. −0·02 mmol/l) and cholesterol:HDL-C ratio (−0·42 v. 0·01, P<0·05). Serum cholesterol, LDL cholesterol and glucose concentrations and body composition variables remained unchanged. It is concluded that administration of PSO for 4 weeks in hyperlipidaemic subjects had favourable effects on lipid profiles including TAG and TAG:HDL-C ratio.

Pomegranate seed oil: Hyperlipidaemia: Randomised clinical trials

Normalisation of dyslipidaemia, a modifiable risk factor, can prevent or reduce the risk of CVD, which is a leading cause of death worldwide2,3.

In order to treat dyslipidaemia, different pharmacologic and non-pharmacologic approaches have been used. Statins are the first-line therapy for management of increased LDL cholesterol (LDL-C), despite adverse effects such as gastrointestinal symptoms and muscle aches3. Although fibrates may decrease serum TAG and increase serum HDL cholesterol (HDL-C), more powerful drugs for normalisation of these lipids need to be investigated.

Conjugated fatty acids are well-known hypolipidaemic agents4,5 documented for their effect on lipid metabolism6. PSO (Punica granatum) consists of about 80 % conjugated octadecatrienoic fatty acids, with a high content of 9-cis, 11-trans, 13-cis acid or punicic acid (PA), one of the isomers of conjugated linolenic acid (CLN)7, animal studies report controversial results for the hypolipidaemic role of PSO8,9.

Although Arao et al.8 showed that PSO supplementation in obese hyperlipidaemic Otsuka—Long–Evans–Tokushima fatty rats reduces TAG accumulation, Yang et al.9 demonstrated that PSO does not alter serum cholesterol concentration.

Considering the lack of data on the effect of PSO on lipid profiles in human subjects, the present study investigated the effect of PSO treatment on lipid profiles of hyperlipidaemic subjects.

Subjects and methods

Subjects and study design

This is a parallel, randomised, double-blind and placebo-controlled study. Subjects were recruited from an endocrine clinic in sequential fashion. Inclusion criteria were age over 20 years, not pregnant or lactating, having no diagnosed
Pomegranate seed oil and lipid profile

Pomegranate seed oil (PSO) capsules (Pometane) were obtained from Vitane Pharma GmbH, Inc. (Wolfratshausen, Germany), and the raw material for PSO was obtained from Sarouneh Company (Urmia, Iran). Chemical analysis of PSO showed the major forms of fatty acids to be: PA (18:3) 72.0 %; oil acid (18:2) 10.7 %; linoleic acid (18:2) 8.4 %; palmitic acid (16:0) 4.0 %; stearic acid (18:0) 2.6 %; trace amounts (<1%) of other fatty acids, with total amount of 7.3% of SFA, 11.7% of MUFA and 81% of PUFA; crude PSO contained the antioxidant vitamin E of 36.90 mg/100 g and was kept at cold temperature to avoid oxidation until formulated in capsules.

Biochemical analysis

Fasting blood samples were taken after 12:00–14:00 hours overnight fasting at baseline and 4 weeks after intervention, and they were centrifuged to separate serum, for biochemical analysis, conducted at the endocrine research laboratory on the day of blood collection. Fasting plasma glucose was measured by the enzymatic colorimetric method using a glucose oxidation kit. Serum total cholesterol and TAG levels were assayed using enzymatic colorimetric tests with cholesterol esterase and cholesterol oxidase and glycerophosphate oxidase, respectively; HDL-C was measured by the immunoturbidimetry method after precipitation of apo B containing lipoproteins with phosphotungstic acid (Pars Azmoun Company, Tehran, Iran). LDL-C was calculated from serum total cholesterol and TAG and HDL-C, according to Friedewald's equation (10); it was not calculated when serum TG > 4.52 mmol/l. Insulin and oxidised LDL were measured by

Table 1. Anthropometric, body composition, blood pressure and biochemical variables at baseline and after 4 weeks in the pomegranate seed oil (PSO) and placebo groups†

<table>
<thead>
<tr>
<th></th>
<th>PSO (n 23)</th>
<th>Placebo (n 22)</th>
<th>P for treatment effect‡</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Baseline</td>
<td>4 weeks</td>
<td>Baseline</td>
</tr>
<tr>
<td>Mean</td>
<td>SD</td>
<td>Mean</td>
<td>SD</td>
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<tr>
<td>----------------------</td>
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</tr>
<tr>
<td>Cholesterol (mmol/l)</td>
<td>6.79±0.81</td>
<td>6.92±1.22</td>
<td>6.94±0.86</td>
</tr>
<tr>
<td>TAG (mmol/l)</td>
<td>3.45±1.56</td>
<td>2.75±1.40</td>
<td>3.63±1.79</td>
</tr>
<tr>
<td>LDL-C (mmol/l)</td>
<td>3.90±1.09</td>
<td>4.39±1.48</td>
<td>3.95±1.14</td>
</tr>
<tr>
<td>HDL-C (mmol/l)</td>
<td>1.25±0.39</td>
<td>1.38±0.44</td>
<td>1.27±0.23</td>
</tr>
<tr>
<td>Ox-LDL (mU/l)</td>
<td>167±43</td>
<td>170±40</td>
<td>173±39</td>
</tr>
<tr>
<td>Cholesterol:HDL-C</td>
<td>3.37±1.24</td>
<td>3.41±1.18</td>
<td>3.19±1.12</td>
</tr>
<tr>
<td>Cholesterol:HDL-C</td>
<td>5.87±1.67</td>
<td>5.45±1.51</td>
<td>5.58±1.21</td>
</tr>
<tr>
<td>TAG:HDL-C ratio</td>
<td>7.49±4.95</td>
<td>5.73±4.55</td>
<td>6.92±3.75</td>
</tr>
<tr>
<td>Insulin (mU/l)</td>
<td>7.57±3.18</td>
<td>7.9±3.36</td>
<td>7.06±2.87</td>
</tr>
<tr>
<td>Insulin:glucose ratio</td>
<td>0.08±0.04</td>
<td>0.09±0.04</td>
<td>0.08±0.04</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>1.68±0.69</td>
<td>1.80±0.84</td>
<td>1.55±0.65</td>
</tr>
<tr>
<td>Body fat (%)</td>
<td>36.6±6.8</td>
<td>36.1±10.0</td>
<td>41.4±6.4</td>
</tr>
<tr>
<td>Fat mass (kg)</td>
<td>27.2±6.6</td>
<td>26.8±9.1</td>
<td>31.7±8.6</td>
</tr>
<tr>
<td>Lean mass (kg)</td>
<td>47.0±7.9</td>
<td>43.4±11.2</td>
<td>44.0±6.1</td>
</tr>
<tr>
<td>Waist:hip ratio</td>
<td>0.83±0.07</td>
<td>0.81±0.19</td>
<td>0.87±0.09</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>27.2±3.1</td>
<td>27.0±3.2</td>
<td>28.3±3.7</td>
</tr>
</tbody>
</table>

**L**D-C, **LDL** cholesterol; **HDL-C, HDL** cholesterol; **Ox-LDL**, oxidised LDL; **HOMA-IR**, homeostasis model for insulin resistance.

Mean values were significantly different within the groups using paired t test: *P<0.009, **P<0.031, ***P<0.001.

† All PSO variables were not significant at baseline, in comparison to placebo.

‡ Analysis of covariance with 4-week values as dependent variables, baseline values as covariates and treatment group as a fixed factor.
ELISA kits (Merckodia AB Company, Uppsala, Sweden); insulin resistance was calculated by the homeostasis model for insulin resistance (HOMA-IR) model, using the formula:

\[
\text{HOMA-IR} = \frac{\text{glucose (mmol/l)} \times \text{insulin (mU/l)}}{22.5}.
\]

**Statistical method**

To have a clinically significant effect on lowering serum cholesterol\(^{(11)}\), we based our sample calculation to detect a difference of 200 mg/l and an SD of 200 mg/l in endpoint cholesterol concentrations between the PSO and placebo groups, and we found that a sample size of 21 was sufficient with 95 % CI and 90 % power in each group. Fifty-one hyperlipidaemic subjects were enrolled to compensate for eventual study dropouts.

SPSS (version 16.0; SPSS, Inc., Chicago, IL, USA) was used for all statistical analyses. Differences between the two groups at baseline were tested with Student’s t test and the Mann–Whitney test. Paired Student’s t test and Wilcoxon ranked test were used to compare the baseline and 4-week values in each group. To distinguish treatment effect between the groups, following adjustment of their baseline values, analysis of covariance test was used.

**Results**

Forty-five of fifty-one patients enrolled in our study were eligible for final analysis, as six withdrew and did not provide 4-week blood samples. Mean values for baseline body weight and age were 74·2 (SD 10·0) and 75·7 (SD 12·2) kg, and 51 (SD 9) and 55 (SD 9) years for the PSO and placebo groups, respectively. No significant differences between the groups were seen for age, sex, weight, height, consumption of lipid lowering drugs or n-3 supplements and smoking. All the subjects maintained good health throughout the study, without any major adverse events.

A comparison of dietary intake and physical activity variables at baseline and after 4 weeks between the PSO and placebo groups revealed no significant differences. Energy intake and dietary cholesterol decreased in the placebo group during the treatment period (\(P < 0.05\)).

The effects of PSO on biochemical, anthropometric, body composition and blood pressure variables are given in Table 1; PSO administration resulted in a decrease in serum TAG concentration (2·75 (SD 1·40) v. 3·45 (SD 1·56) mmol/l, \(P < 0.01\)) and a non-significant increase in HDL-C concentration (1·38 (SD 0·44) v. 1·25 (SD 0·39) mmol/l, \(P = 0·081\)) compared with baseline values; TAG:HDL-C ratio decreased significantly within the PSO group (5·73 (SD 4·55) v. 7·49 (SD 4·95), \(P < 0·03\)). Comparison of treatment effects between the groups with baseline values as covariates revealed lower values for cholesterol:HDL-C (5·45 (SD 1·51) v. 5·89 (SD 1·43), \(P < 0·05\)) and higher, but non-significant HDL-C concentrations (1·38 (SD 0·44) v. 1·25 (SD 0·26) mmol/l, \(P = 0·059\)) in the PSO group as compared with the placebo group, respectively. Other lipid profile variables including cholesterol, LDL-C, oxidised LDL and LDL-C:HDL-C ratio revealed no significant differences within or between the groups. Fat mass, BMI and waist:hip ratio decreased in the PSO group, but not significantly. PSO had no effect on systolic and diastolic blood pressure within or between the groups (data not shown). Means for differences were significant in HDL-C concentration (0·13 v. −0·02 mmol/l) and cholesterol:HDL-C ratio (−0·42 v. 0·01) when the PSO group was compared with the placebo group (\(P < 0·05\) for both) (Fig. 1).

**Discussion**

In the present study, 4 weeks of PSO treatment decreased serum TAG concentration, TAG:HDL-C ratio and cholesterol:HDL-C ratio in the PSO group, as compared with the placebo group. Insignificant differences in physical activity levels and daily diets of the subjects throughout the study duration indicate that any changes in variables were probably due to PSO treatment. Since a target of 2·11 mmol/l or less for serum TAG in elderly women has been proposed\(^{(12)}\), the decrease in serum TAG from 3·45 (SD 1·56) to 2·75 (SD 1·40) mmol/l may be clinically valuable. Data indicate that each mg/dl increase in HDL-C corresponds to a 2–3 % risk reduction of CHD\(^{(13)}\); hence our HDL-C increase of 50 mg/l in the present study reflects a 10–15 % risk reduction of CHD. Besides the lipid profile, we investigated TAG:HDL-C, cholesterol:HDL-C and LDL-C:HDL-C as lipid-related indices. The TAG:HDL-C ratio predicts CVD and metabolic syndrome\(^{(14)}\), explaining its crucial role in treatment guidelines; therefore, 1·8- fold reduction of the TAG:HDL-C ratio in the present study is another significant finding. High cholesterol:HDL-C ratios (≥4) are also associated with coronary artery disease\(^{(15)}\), and we have

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**Fig. 1.** Mean differences compared with baseline in the pomegranate seed oil (PSO; ■) and placebo groups (□). Ox-LDL-C, oxidised LDL; TC, total cholesterol; HDL-C, HDL cholesterol; LDL-C, LDL cholesterol; HOMA-IR, homeostasis model for insulin resistance. *\(P < 0·05\).
found a significant difference in cholesterol: HDL-C ratio after adjusting for baseline values, in PSO group v. placebo group.

A recent study in healthy human subjects reported Trichosanthes kirilowii seeds, naturally occurring sources of PA, had no significant effect on serum lipid levels contrary to our finding which may be attributable to the pre-existence of hyperlipidaemia in the present study. Current data on the effect of PSO on lipid concentration and metabolism are controversial\(^8\)\(^,\)\(^9\)\(^,\)\(^17\). Similar to our findings, the liver accumulation of TAG was reduced by PSO administration\(^8\)\(^,\)\(^18\); a plausible mechanism, demonstrated in an in vitro study, that 9-cis, 11-trans, 13-cis CLN suppressed cellular synthesis of TAG in HepG2 cells\(^19\).

One study suggests a metabolic pathway for 9-cis, 11-trans, 13-cis CLN is its conversion to 9-cis, 11-trans conjugated linoleic acid (CLA) in a rat model; this conversion coefficient is about 12% for PSO\(^20\). Also two recently published articles have demonstrated that PA from *T. kirilowii* seed oil incorporated into different tissues and serum metabolites to CLA in rats and human subjects\(^21\)\(^,\)\(^22\).

Hence, the physiological activities of PSO may be attributable to 9-cis, 11-trans CLA derived endogenously or to PA itself. Scientific evidence reveals the inconsistent effects of 9-cis, 11-trans CLA on lipid profiles\(^23\)\(^–\)\(^26\), alterations that may occur mostly through the carnitine palmitoyltransferase activity-enzyme involved in fatty acid β-oxidation, as mitochondrial and peroxisomal carnitine palmitoyltransferase activity is reported to increase with PSO\(^18\) and CLN\(^27\); therefore, there is no conclusive evidence to confirm the effect of CLN and CLA on lipid profiles, but it does seem that the isomer type and amount of CLN and CLA could be determining factors.

In the present study, no significant changes were observed in body composition (fat or lean mass), BMI or waist:hip ratio following treatment. Since energy intake, a main contributor to weight gain, did not change during the 4 weeks, decrease in body composition indices is attributable either to PSO or to 9-cis, 11-trans CLA. *In vivo* studies on weight management and body composition found a reduction on perirenal, epididymal and omental adipose tissues following PA supplementation\(^8\)\(^,\)\(^18\).

In the present study, we found no beneficial relationship between insulin profiles and PSO supplementation. A recent animal study showed that PSO reduced the risk of diabetes by lowering glucose and insulin concentrations and insulin sensitivity\(^28\), a result also consistent for specific isomers of CLA, as shown by the Risérus *et al.*\(^24\) study which reported that administration of 3 g of 9-cis, 11-trans CLA over 12 weeks yielded a 15% decrease in insulin sensitivity.

The present study has a few limitations. Serum PSO and CLA concentrations were not measured, which may further have facilitated assessment of compliance of the subjects by measuring PSO metabolites. Our sample size was small, and patients with mixed hyperlipidaemia were enrolled; study duration was 4 weeks and only one blood sample was obtained following intervention. Another limitation might be that we powered our study based on a 200 mg/l difference in total cholesterol between the PSO and placebo groups; this, however, was not achieved, and despite changes that were observed, one may speculate that the present study is underpowered.

To conclude, the present study found that PSO consumption in hyperlipidaemic subjects did not alter cholesterol and LDL-C but did reduce TAG and TAG: HDL-C ratio during 4-week study period. Further studies of larger samples with longer durations need to confirm the effects of PSO and related mechanisms.

Acknowledgements

The authors would like to thank the subjects who participated in the present study. The present study was supported by a financial grant (no. 234) from the Research Institute of Endocrine Sciences, Shahid Beheshti University Medical Sciences and Pharmacy and Pharmaceutical Sciences Research Center, Tehran University of Medical Sciences, Iran. None of the authors had any personal or financial conflicts of interest. P. M. was involved in design, implementation and analysis; M. R. F. was involved in preparation of PSO capsules and implementation; G. A. was involved in review of the literature, implementation and analysis; A. S. supervised of technical issues in the preparation of PSO and design; F. A. supervised overall project, design, implementation and analysis.

References


