Fibre-mediated physiological effects of raw and processed carrots in humans*

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Fibre-mediated physiological effects of raw and processed carrots were investigated in twenty-four young women under strict dietary control in two randomized crossover studies. For 3 weeks between 405 and 688 g of either raw frozen, blanched or canned carrots (first study), or raw or raw frozen carrots (second study) were consumed in addition to a low-fibre basal diet. Carrots provided 15 g dietary fibre (DF)/d. Total DF intake was 16.0 to 19.0 g (control periods) and 31 to 34 g (experimental periods). Faecal bulking effects of raw and processed carrots were similar (between 2.4 and 3.7 g additional stool/g carrot fibre in the diet). Faecal excretion of dry matter, fibre, and protein also increased significantly during carrot consumption. Fermentability of carrot fibre constituents was high (91–94%) and independent of processing, in spite of differences in the distribution of soluble and insoluble fibre and in the texture of raw and processed carrots. There was no effect of either type of carrot on serum total and high-density-lipoprotein-cholesterol or on faecal bile acid excretion.

Carrots: Fibre: Humans

Much of the plant food we consume is processed. Freezing, blanching, cooking and canning are procedures which are often applied to vegetables. Cooking especially is usually accompanied by tissue softening and changes in textural characteristics. Heating can affect the fibre content or modify the fibre distribution between water-soluble and insoluble fractions in several ways (Anderson & Clydesdale, 1980; Nyman et al. 1987; Lintas & Cappeloni, 1988). Losses of soluble components such as sugars and minerals into the cooking water result in an apparent increase in fibre content. Solubilization of insoluble fibre increases the portion of soluble fibre. Degradation of polysaccharides into low-molecular-weight fragments can have the same effect, but it can also decrease total fibre content, if oligosaccharides are formed which escape analysis by the normal methods for dietary fibre.

The chemical structure and the physicochemical properties of dietary fibre are both thought to determine their physiological effects. Therefore, it is often stated that modification of these properties may affect the action of fibre. Fibre sources containing high proportions of insoluble components like cereal brans and whole grain cereals have good faecal bulking capacities (Cummings, 1986), however, cooking of bran diminishes this

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effect (Wyman et al. 1976). More soluble types of fibre like those contained in fruits and vegetables or purified fibres like pectin are less effective in this respect. On the other hand, these fibres have a greater effect on blood lipids and on carbohydrate metabolism when they are consumed in sufficient amounts (Berger & Venhaus, 1992; Truswell & Beynen, 1992). Many actions of fibre are influenced by its fermentability in the large intestine. The effect on stool weight seems to be inversely related to the bacterial degradation of fibre (Stephen & Cummings, 1980a). Soluble fibres are more easily fermented than insoluble fibres (Cummings, 1984). Heat treatment of cereals at low moisture, like popping and extrusion cooking, has been shown to increase fibre fermentability in the rat due to an increase of soluble components (Björck et al. 1984; Nyman et al. 1987a).

However, few human studies have actually addressed the effects of processing at high moisture levels such as those found in vegetables. The purpose of the present investigation was therefore to provide information on whether or not processing of a commonly eaten vegetable, i.e. carrots, affected the composition and solubility of fibre and thereby its physiological action in humans. Our main focus points in this study were colonic fermentation, bulking capacity of carrot fibre, and faecal excretion of fat, protein and bile acids. In addition, we hoped to resolve the question of whether or not carrots and carrot fibres exert cholesterol-lowering effects (Jenkins et al. 1979; Robertson et al. 1979).

EXPERIMENTAL

Subjects

A total of twenty-four healthy, free-living female subjects (22–29 years) took part in a series of balance experiments organized in two distinct studies. Informed written consent was obtained from all volunteers. The studies were approved by the Ethical Committee of the Medical Faculty of the University of Kiel.

Study design

All food consumed during the experiments was prepared in the institute kitchen and was weighed to the nearest gram. Extra food and energy-containing beverages were not permitted. The subjects had lunch together in the institute; foods for all other meals were prepacked for home consumption.

First study. The study comprised four experimental periods of 21 d each. The periods were separated from each other by at least 3 weeks to ensure that one dietary period had no residual effect on the next. During all periods the subjects consumed the same strictly controlled basal diet. This diet was supplemented either with a sugar jelly (low-fibre control diet), raw frozen, blanched frozen or canned carrots. The twelve subjects who took part in this study consumed each of the four diets in a randomized Latin square design.

Second study. Twelve subjects participated in the second study. The study comprised two experimental periods of 36 d each; the periods were separated from each other by 3 weeks. During the first 15 d of each period, all subjects consumed a low-fibre basal diet supplemented with a sugar jelly (low-fibre control diet). During the following 21 d the jelly was replaced by either raw or raw frozen carrots, subjects being assigned to these two treatments in a randomized crossover fashion.

Diets

The subjects had a controlled food intake that maintained their body weight in a range of ±1 kg of their starting weight. Each subject’s normal energy intake was calculated from a 7 d diet record by use of a food table (Deutsche Forschungsanstalt für Lebensmittelchemie,
Table 1. *Foods consumed daily during the low-fibre basal diet*.

<table>
<thead>
<tr>
<th>Food</th>
<th>Intake (g/d)</th>
<th>Food</th>
<th>Intake (g/d)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rye mixed bread</td>
<td>200</td>
<td>Yoghurt</td>
<td>150</td>
</tr>
<tr>
<td>Potatoes</td>
<td>100</td>
<td>Cheese</td>
<td>75</td>
</tr>
<tr>
<td>Cucumber or salad†</td>
<td>150</td>
<td>Soft cheese</td>
<td>40</td>
</tr>
<tr>
<td>Strawberries or orange†</td>
<td>150</td>
<td>Sausage</td>
<td>40</td>
</tr>
<tr>
<td>Meat balls</td>
<td>75</td>
<td>Honey/marmalade</td>
<td>40</td>
</tr>
<tr>
<td>Pudding‡</td>
<td>150</td>
<td>Margarine</td>
<td>20</td>
</tr>
</tbody>
</table>

* Each subject consumed the same amount of these foods daily during each experimental period of the studies. Additional amounts of pudding, soft cheese, sausage, margarine and honey were eaten to adjust food intake to individual energy requirement.
† Foods were consumed in rotation.
‡ Pudding was prepared from milk, sugar, starch, eggs and vanilla.

1986). The low-fibre basal diet consisted of two 1 d menus of similar composition, which were fed in rotation throughout both studies. All subjects consumed the same amount of fibre-containing foods, whereas fibre-free foods corresponded to individual energy requirements. The foods consumed during the basal diet are given in Table 1.

**Carrots**

Carrots (*Daucus carota* sp. sativus) harvested in 1988 and in 1990 were used in the studies.

During the first study, raw carrots (harvested in 1988) from the same batch were washed, steam-peeled, trimmed, cut into pieces (10 x 10 x 10 mm) and rinsed. The prepared vegetables were either frozen and packaged (raw frozen carrots), or water blanched at 98–100° for 2 min, cooled after blanching at 10° for 2 min, frozen and packaged (blanched frozen carrots), or canned at 120° for 20 min (canned carrots). Carrots were prepared in an amount sufficient for the consumption during all study periods. Thus it was ensured that each type of carrots had a constant composition throughout the study.

During the second study, raw and raw frozen carrots were obtained from one batch of carrots harvested in 1990. Raw frozen carrots were prepared as described above. Raw carrots were stored at 2° until they were consumed. They were prepared by washing, peeling by hand and coarse scraping.

Daily intakes of raw and processed carrots were calculated to provide 15 g total dietary fibre, as analysed by the method of Prosky et al. (1985). Table 2 shows the intake of carrots, of total dietary fibre and sugar derived from carrots and the intake of sugar from jelly. Table 3 gives the intake of energy and nutrients during the experimental periods.

**Blood collection**

During the first study, at days 2 and 21, in ten subjects venous blood samples were obtained from a superficial arm vein by venepuncture after 12 h fasting. During the second study, blood was drawn in eleven subjects at days 2, 15 and 36.

**Balance technique**

In the first study, balances were performed during the last week of each experimental period. During the second study, balances were performed from days 9 to 15 (low-fibre control diet) and from days 30 to 36 of each experimental period. Duplicates of all food...
Table 2. Daily intake (g/d) of carrots, carrot fibre and sugar from carrots and jelly*

<table>
<thead>
<tr>
<th></th>
<th>Carrots</th>
<th>Sugar from:</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Amount</td>
<td>Dietary fibre</td>
</tr>
<tr>
<td>Carrots harvested in 1988</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(first study)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Raw frozen</td>
<td>575</td>
<td>15·0</td>
</tr>
<tr>
<td>Blanched frozen</td>
<td>508</td>
<td>15·0</td>
</tr>
<tr>
<td>Canned</td>
<td>688</td>
<td>15·0</td>
</tr>
<tr>
<td>Carrots harvested in 1990</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(second study)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Raw</td>
<td>405</td>
<td>15·0</td>
</tr>
<tr>
<td>Raw frozen</td>
<td>469</td>
<td>15·0</td>
</tr>
</tbody>
</table>

* Each subject consumed the same amount of carrots.
† Sum of sucrose, glucose and fructose.
‡ Sucrose only.

consumed were weighed, homogenized, and frozen at \(-25^\circ\) until freeze-dried. Faeces were collected quantitatively and frozen at \(-25^\circ\). Acid brilliant green (E 142, provided by H. Schulz, Dragoco, Holzminden, Germany) was given as a faecal marker at the beginning and the end of each collection period. After each balance period faeces were thawed, pooled together, and homogenized before freeze-drying a portion. Faeces collected during the low-fibre control diets (days 9 to 15) in the two experimental periods of the second study were pooled together and one sample for each subject was freeze-dried.

**Analytical methods**

Freeze-dried samples of food and faeces were milled through a 0·5 mm mesh screen. Total dietary fibre in the diet was determined gravimetrically by the method of Prosky et al. (1985) using the buffers as described by Prosky et al. (1988). Insoluble dietary fibre was measured according to Prosky et al. (1988). Neutral non-starch polysaccharides (NSP) in foods and faeces were determined by GLC as alditol acetates by the method C of Theander & Westerlund (1986) using 1-methylimidazole as a catalyst for the derivatization of NSP monomers. Corrections for hydrolytic losses and detector response were made by performing the analyses with known sugar standards. Uronic acids were measured in the acidic hydrolysate according to the method of Englyst & Cummings (1984). Total NSP was calculated as the sum of neutral sugars and uronic acids and expressed as polysaccharides (weight of monomers \(\times 0.9\)). N was analysed by a microKjeldahl method; protein was calculated as N \(\times 6.25\). Crude fat was determined after acid hydrolysis by extraction with petroleum ether (40–60°). Ash was determined in a muffle furnace at 550°. Dry matter of diets and faeces was obtained by drying the freeze-dried samples to constant weight at 105° (Arbeitsgemeinschaft Getreideforschung, 1978). Starch was measured as described by Bach Knudsen et al. (1987). Sugars were analysed after water extraction using enzyme test kits (Boehringer Mannheim GmbH, Mannheim, Germany). Serum total cholesterol was determined enzymically by the CHOD-Iodide method (Merckotest 14350; Merck, Darmstadt, Germany). High-density-lipoprotein (HDL)-cholesterol was measured by precipitating all other cholesterol fractions with heparin and Mg (Fällungsreagenz 15007; Merck) (Eckel et al. 1977) leaving a supernatant that was assayed for cholesterol by use of
Table 3. Intake of energy (MJ/d) and nutrients (g/d) by young women during two studies of the physiological effects of raw and processed carrot consumption*  
(Mean values and standard deviations for twelve subjects during the balance periods)

<table>
<thead>
<tr>
<th></th>
<th>First study</th>
<th>Second study</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Diets containing carrots harvested in 1988</td>
<td>Diets containing carrots harvested in 1990</td>
</tr>
<tr>
<td>Low-fibre</td>
<td>Low-fibre control diet</td>
<td>Low-fibre control diet</td>
</tr>
<tr>
<td>control diet</td>
<td>Raw frozen</td>
<td>Raw</td>
</tr>
<tr>
<td></td>
<td>Mean  SD</td>
<td>Mean  SD</td>
</tr>
<tr>
<td>Energy</td>
<td>7.0  0.8</td>
<td>7.2  0.9</td>
</tr>
<tr>
<td>Protein</td>
<td>82.3  9.2</td>
<td>83.5  9.3</td>
</tr>
<tr>
<td>Fat</td>
<td>61.1  16.9</td>
<td>61.9  17.9</td>
</tr>
<tr>
<td>Starch</td>
<td>103.3  2.3</td>
<td>103.5  2.5</td>
</tr>
<tr>
<td>Glucose</td>
<td>13.1  3.9</td>
<td>18.6  3.8</td>
</tr>
<tr>
<td>Fructose</td>
<td>13.7  3.7</td>
<td>20.9  3.6</td>
</tr>
<tr>
<td>Sucrose</td>
<td>46.7  2.4</td>
<td>32.4  2.7</td>
</tr>
<tr>
<td>Lactose</td>
<td>20.8  3.6</td>
<td>21.2  3.3</td>
</tr>
<tr>
<td>Galactose</td>
<td>16  0.1</td>
<td>1.6  0.2</td>
</tr>
<tr>
<td>Total dietary</td>
<td>19.0  0.3</td>
<td>33.9  0.3</td>
</tr>
<tr>
<td>fibre</td>
<td>Non-starch polysaccharides</td>
<td>Non-starch polysaccharides</td>
</tr>
<tr>
<td></td>
<td>Mean  SD</td>
<td>Mean  SD</td>
</tr>
<tr>
<td></td>
<td>17.4  0.7</td>
<td>32.2  0.8</td>
</tr>
</tbody>
</table>

* For details of subjects and diets, see Tables 1 and 2, and pp. 580–581.
a test kit (Merckotest 14350). Accuracy of total cholesterol and HDL-cholesterol determinations were ensured through measurement of a standard serum (Kontrollogen LP; Behringwerke, Marburg, Germany). Faecal bile acids were measured by GLC after extraction from powdered dry faeces, saponification, separation from neutral steroids and derivatization as described by Schweizer et al. (1983).

**Calculations**

Soluble fibre was calculated as the difference between total and insoluble fibre. Fermentability of NSP constituents of the diets was calculated as the difference between dietary intake and faecal excretion, expressed as a percentage of intake.

During the second study, intake, faecal excretion and fermentation of NSP derived from carrots were also calculated. Intake and faecal excretion of carrot NSP were calculated as the difference between carrot-containing diets and the corresponding low-fibre control diets. Fermentation of carrot NSP was calculated as the difference between estimated dietary intake and faecal excretion, expressed as a percentage of intake. Faecal output due to carrots during the second study was calculated as the difference between carrot-containing diets and the corresponding low-fibre control diets.

**Statistical analysis**

The effects of processing on the composition of the carrots consumed during the first and second studies respectively, and the effects of the harvest year on the composition of raw frozen carrots were analysed by one-way analysis of variance. Linear contrasts were used to compare treatment means.

The results of the studies were treated statistically by analysis of variance in which subject, period and diet effects were evaluated. Results obtained with diets containing raw frozen carrots harvested in 1988 and in 1990 were compared by one-way analysis of variance in which the effect of the harvest year was determined. Where significant differences between diets were established, differences were tested by linear contrasts. Linear contrasts were established by Student’s *t*-test with comparison-wise error rates (General linear models procedure (GLM), SAS Institute, Heidelberg, Germany). All tests were considered significant at the 5% level (*P* < 0.05).

**RESULTS**

**Composition of carrots**

The total dietary fibre and NSP content of the carrots, the percentage of soluble and insoluble fibre and the NSP monomers are given in Table 4.

Heat treatment, i.e. blanching and canning, increased the fibre content in carrot dry matter compared with raw frozen carrots due to losses in non-fibre material, mainly sugars. On a fresh weight basis the fibre values were higher in blanched and lower in canned than in raw frozen carrots. Blanching and canning also increased the percentage of soluble fibre compared with the raw frozen carrots. The raw frozen carrots from the 1990 harvest contained more dietary fibre than those harvested in 1988, calculated on a fresh weight basis, whereas the values in the dry materials were the same. Freezing reduced the dietary fibre content compared with the raw carrots when the values were calculated on a fresh weight basis, but in the dry material there was no difference. The percentage of soluble fibre was the same for both raw and raw frozen carrots and was higher compared with the value of the raw frozen carrots from the 1988 harvest. The main dietary fibre constituents in
Table 4. Content of total dietary fibre and non-starch polysaccharides (NSP) in carrots, and percentage of soluble and insoluble dietary fibre and of NSP monomers

(Values are means with their standard errors of four analyses)

<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Raw (Mean)</td>
<td>Blanched (Mean)</td>
<td>Canned (Mean)</td>
</tr>
<tr>
<td>Total dietary fibre (g/kg FW)</td>
<td>26.1</td>
<td>29.5</td>
</tr>
<tr>
<td>Total dietary fibre (g/kg DM)</td>
<td>268</td>
<td>333</td>
</tr>
<tr>
<td>Soluble (% TDF)†</td>
<td>26</td>
<td>43</td>
</tr>
<tr>
<td>Insoluble (% TDF)</td>
<td>74</td>
<td>57</td>
</tr>
<tr>
<td>Total NSP (g/kg DM)</td>
<td>280</td>
<td>342</td>
</tr>
<tr>
<td>NSP-monomers (% NSP)</td>
<td>Rhamnose</td>
<td>3.6</td>
</tr>
<tr>
<td>Arabinose</td>
<td>7.9</td>
<td>7.9</td>
</tr>
<tr>
<td>Xylose</td>
<td>18</td>
<td>2.6</td>
</tr>
<tr>
<td>Mannose</td>
<td>3.2</td>
<td>3.2</td>
</tr>
<tr>
<td>Galactose</td>
<td>11.1</td>
<td>11.4</td>
</tr>
<tr>
<td>Glucose</td>
<td>33.3</td>
<td>34.2</td>
</tr>
<tr>
<td>Uronic acids</td>
<td>39.1</td>
<td>36.8</td>
</tr>
</tbody>
</table>

FW, fresh weight; DM, dry matter; TDF, total dietary fibre; NS, not significant.
* P < 0.05; ** P < 0.01; *** P < 0.001.
† Values obtained by calculation, see p. 584.
carrots were uronic acids and glucose, followed by galactose and arabinose. In comparison to the raw frozen carrots, blanching had no effect on the percentage of glucose, but decreased uronic acids, whereas canning caused a decrease in uronic acids and an increase in glucose. Compared with the raw carrots, freezing increased arabinose and galactose and decreased uronic acids.

Effect of carrots on faecal excretion

The effect of the consumption of carrots harvested in 1988 on important faecal variables is shown in Table 5. The consumption of carrots caused an increase in faecal wet and dry weight compared with the values on the control diets. For each gram of additional fibre, wet weight increased between 2.4 and 3.7 g, mean dry weight by 0.4 g. The fibre from the various carrots led also to significantly elevated losses of water, NSP, N and ash, whereas fat excretion was not affected. An influence of processing of carrots could only be observed in the case of stool water which was higher during the consumption of raw frozen and blanched frozen carrots compared with canned carrots. The mean daily faecal bile acid excretion and the high extent of conversion of primary to secondary bile acids were similar in all periods of the first study.

The results of the second study (carrots harvested in 1990) are shown in Table 6. Faecal output increased due to carrot consumption, but no differences in the effects of raw and raw frozen carrots could be observed.

Fermentation of carrot NSP

Table 7 shows the intake, faecal excretion and fermentation of NSP monomers during the first study. Although the diets containing raw frozen, blanched or canned carrots from the 1988 harvest provided higher amounts of NSP monomers than the control diet there was no, or only a small, increase in faecal excretion of the monosaccharides, even when the differences from the control diet were significant. The additional fibre constituents were extensively degraded, leading to higher values for the fermentation of NSP monomers during the carrot-containing diets than during the control diet. Compared with the control, total NSP intake increased by 15.8, 15.5 and 15.5 g/d when the raw frozen, blanched, and canned carrots respectively, were consumed. The corresponding NSP losses increased on average by 1.3, 1.4 and 0.9 g/d respectively. From these values it was estimated that a mean of 92, 91 and 94% of fibre in the raw frozen, blanched, and canned carrots from the 1988 harvest respectively, was broken down. No significant effect of processing of carrots on the fermentation of NSP could be observed.

Table 8 gives the intake, faecal excretion and fermentation of NSP during the second study. There were no differences in faecal losses and digestibilities of NSP monomers between the two carrot-containing diets with the exception of galactose fermentation. The additional fibre constituents provided by raw or raw frozen carrots from the 1990 harvest were extensively fermented, independent of whether the fibre originated from raw or raw frozen carrots. The most resistant NSP constituents were glucose-containing polymers.

Table 9 shows a comparison of the effects of the diets containing raw frozen carrots from the two harvest years. There were few differences between the effects of the two diets.

Effects on serum cholesterol

Table 10 gives the concentrations of serum total cholesterol and of HDL-cholesterol during the first study. Both serum total cholesterol and HDL-cholesterol decreased within each experimental period, independent of the diet. However, there were no differences in the
Table 5. Study 1. Effects of diets containing different processed carrots (harvested in 1988) on faecal output in young women†
(Values are means with their pooled standard errors for twelve subjects)

<table>
<thead>
<tr>
<th>Faecal output</th>
<th>Low-fibre control diet (Mean)</th>
<th>Diets containing processed carrots</th>
<th>Statistical significance of diet effects</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Wet wt (g/d)</td>
<td>93.2</td>
<td>149.3</td>
</tr>
<tr>
<td></td>
<td>Dry wt (g/d)</td>
<td>21.2</td>
<td>27.6</td>
</tr>
<tr>
<td></td>
<td>Water (%)</td>
<td>74.7</td>
<td>79.3</td>
</tr>
<tr>
<td></td>
<td>NSP (g/d)</td>
<td>3.4</td>
<td>4.7</td>
</tr>
<tr>
<td></td>
<td>Nitrogen (g/d)</td>
<td>1.3</td>
<td>1.6</td>
</tr>
</tbody>
</table>
|                       | Fat (g/d)                     | 2.0                               | 2.2                                  | 2.2                                  | 2.2                                  | 0.09                                   | NSP, non-starch polysaccharides; NS, not significant.* P < 0.05; *** P < 0.001.† For details of diets and procedures, see Table 1 and pp. 580–584.
|                       | Ash (g/d)                     | 3.5                               | 4.6                                  | 4.6                                  | 4.5                                  | 0.15                                   | ***                                  | NS                  | NS                  | NS                  |
|                       | Cholic acid (mg/d)            | 3                                 | 7                                    | 5                                    | 4                                    | 1.98                                   | NSP                                  | NS                  | NS                  | NS                  |
|                       | Chenodeoxycholic acid (mg/d)  | 2                                 | 3                                    | 2                                    | 2                                    | 1.14                                   | NSP                                  | NS                  | NS                  | NS                  |
|                       | Deoxycholic acid (mg/d)       | 69                                | 76                                   | 85                                   | 69                                   | 8.18                                   | NSP                                  | NS                  | NS                  | NS                  |
|                       | Lithocholic acid (mg/d)       | 42                                | 40                                   | 41                                   | 39                                   | 4.13                                   | NSP                                  | NS                  | NS                  | NS                  |
|                       | Total bile acids (mg/d)       | 116                               | 126                                  | 134                                  | 115                                  | 12.76                                  | NSP                                  | NS                  | NS                  | NS                  |

† For details of diets and procedures, see Table 1 and pp. 580–584.
Table 6. Study 2. Effect of diets containing raw and raw frozen carrots (harvested in 1990) on faecal output in young women†
(Values are means with their pooled standard errors for twelve subjects)

<table>
<thead>
<tr>
<th>Faecal output</th>
<th>Low-fibre control diet‡</th>
<th>Diets containing carrots</th>
<th></th>
<th>Increase in faecal output due to carrots§</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean (SE)</td>
<td>Raw (Mean)</td>
<td>Raw frozen (Mean)</td>
<td>SEM</td>
</tr>
<tr>
<td>Wet wt (g/d)</td>
<td>83.7 (9.56)</td>
<td>132.6 (9.71)</td>
<td>124.4 (9.71)</td>
<td>NS</td>
</tr>
<tr>
<td>Dry wt (g/d)</td>
<td>22.5 (1.34)</td>
<td>28.0 (1.68)</td>
<td>29.4 (1.68)</td>
<td>NS</td>
</tr>
<tr>
<td>Water (%)</td>
<td>71.0 (1.89)</td>
<td>76.5 (1.38)</td>
<td>73.6 (1.38)</td>
<td>NS</td>
</tr>
<tr>
<td>NSP (g/d)</td>
<td>3.4 (0.31)</td>
<td>4.3 (0.28)</td>
<td>4.3 (0.28)</td>
<td>NS</td>
</tr>
<tr>
<td>Nitrogen (g/d)</td>
<td>1.3 (0.09)</td>
<td>1.6 (0.09)</td>
<td>1.7 (0.09)</td>
<td>NS</td>
</tr>
<tr>
<td>Fat (g/d)</td>
<td>2.7 (0.17)</td>
<td>3.1 (0.22)</td>
<td>3.4 (0.22)</td>
<td>NS</td>
</tr>
<tr>
<td>Ash (g/d)</td>
<td>4.0 (0.21)</td>
<td>5.1 (0.32)</td>
<td>5.5 (0.32)</td>
<td>NS</td>
</tr>
</tbody>
</table>

NSP, non-starch polysaccharides; NS, not significant.
† For details of diets and procedures see Tables 1–3 and pp. 580–584.
‡ Values were used for the calculation of the effects of carrots.
§ For the calculation of faecal output due to the intake of carrots see p. 584.
Table 7. Study 1. Intake, faecal excretion and apparent digestibility of non-starch polysaccharide (NSP) monomers by young women consuming a control diet or diets containing different processed carrots (harvested in 1988)†

(Values are means with their pooled standard errors for twelve subjects)

<table>
<thead>
<tr>
<th></th>
<th>Low-fibre control diet (Mean)</th>
<th>Diets containing processed carrots</th>
<th>Statistical significance of diet effects</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Raw frozen (Mean)</td>
<td>Blanched frozen (Mean)</td>
</tr>
<tr>
<td>Rhamnose</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intake (g/d)</td>
<td>0.1</td>
<td>0.6</td>
<td>0.6</td>
</tr>
<tr>
<td>Faecal excretion (g/d)</td>
<td>0.1</td>
<td>0.2</td>
<td>0.2</td>
</tr>
<tr>
<td>Apparent digestibility (%)</td>
<td>NA</td>
<td>63.9</td>
<td>63.9</td>
</tr>
<tr>
<td>Arabinose</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intake (g/d)</td>
<td>2.2</td>
<td>3.6</td>
<td>3.6</td>
</tr>
<tr>
<td>Faecal excretion (g/d)</td>
<td>0.6</td>
<td>0.7</td>
<td>0.7</td>
</tr>
<tr>
<td>Apparent digestibility (%)</td>
<td>73.9</td>
<td>80.2</td>
<td>81.5</td>
</tr>
<tr>
<td>Xylose</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intake (g/d)</td>
<td>3.8</td>
<td>4.1</td>
<td>4.2</td>
</tr>
<tr>
<td>Faecal excretion (g/d)</td>
<td>0.7</td>
<td>0.8</td>
<td>0.8</td>
</tr>
<tr>
<td>Apparent digestibility (%)</td>
<td>82.5</td>
<td>80.7</td>
<td>81.0</td>
</tr>
<tr>
<td>Mannose</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intake (g/d)</td>
<td>1.2</td>
<td>1.7</td>
<td>1.7</td>
</tr>
<tr>
<td>Faecal excretion (g/d)</td>
<td>0.1</td>
<td>0.2</td>
<td>0.2</td>
</tr>
<tr>
<td>Apparent digestibility (%)</td>
<td>91.6</td>
<td>90.2</td>
<td>90.2</td>
</tr>
<tr>
<td>Galactose</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intake (g/d)</td>
<td>2.0</td>
<td>3.7</td>
<td>3.8</td>
</tr>
<tr>
<td>Faecal excretion (g/d)</td>
<td>0.3</td>
<td>0.4</td>
<td>0.4</td>
</tr>
<tr>
<td>Apparent digestibility (%)</td>
<td>84.0</td>
<td>89.0</td>
<td>88.7</td>
</tr>
<tr>
<td>Glucose</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intake (g/d)</td>
<td>6.0</td>
<td>11.1</td>
<td>11.3</td>
</tr>
<tr>
<td>Faecal excretion (g/d)</td>
<td>1.3</td>
<td>1.8</td>
<td>2.1</td>
</tr>
<tr>
<td>Apparent digestibility (%)</td>
<td>78.0</td>
<td>83.7</td>
<td>81.6</td>
</tr>
<tr>
<td>Uronic acids</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intake (g/d)</td>
<td>2.1</td>
<td>8.3</td>
<td>7.8</td>
</tr>
<tr>
<td>Faecal excretion (g/d)</td>
<td>0.3</td>
<td>0.6</td>
<td>0.5</td>
</tr>
<tr>
<td>Apparent digestibility (%)</td>
<td>88.0</td>
<td>93.2</td>
<td>93.5</td>
</tr>
<tr>
<td>Total NSP</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intake (g/d)</td>
<td>17.4</td>
<td>33.2</td>
<td>32.9</td>
</tr>
<tr>
<td>Faecal excretion (g/d)</td>
<td>3.4</td>
<td>4.7</td>
<td>4.9</td>
</tr>
<tr>
<td>Apparent digestibility (%)</td>
<td>80.6</td>
<td>85.9</td>
<td>85.3</td>
</tr>
</tbody>
</table>

NA, not applicable; NS, not significant.

* P < 0.05; ** P < 0.01; *** P < 0.001.

† For details of diets and procedures, see Tables 1–3 and pp. 580–584.
Table 8. Study 2. Intake, faecal excretion and apparent digestibility of non-starch polysaccharide (NSP) monomers by young women consuming a control diet, or diets containing raw or raw frozen carrots (harvested in 1990), together with intake, faecal excretion and apparent digestibility of carrot NSP†

(Values are means and pooled standard errors for twelve subjects)

<table>
<thead>
<tr>
<th>NSP derived from carrots§</th>
<th>Diets containing carrots</th>
<th>Low-fibre control diet‡</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Raw (Mean)</td>
<td>Raw frozen (Mean)</td>
</tr>
<tr>
<td></td>
<td>SEM</td>
<td>SEM</td>
</tr>
<tr>
<td></td>
<td>Statistical significance</td>
<td>Statistical significance</td>
</tr>
<tr>
<td></td>
<td>of diet effects</td>
<td>of diet effects</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Rhamnose</th>
<th>Intake (g/d)</th>
<th>Faecal excretion (g/d)</th>
<th>Apparent digestibility (%)</th>
<th>Intake (g/d)</th>
<th>Faecal excretion (g/d)</th>
<th>Apparent digestibility (%)</th>
<th>Intake (g/d)</th>
<th>Faecal excretion (g/d)</th>
<th>Apparent digestibility (%)</th>
<th>Intake (g/d)</th>
<th>Faecal excretion (g/d)</th>
<th>Apparent digestibility (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean</td>
<td>SE</td>
<td>(Mean)</td>
<td>Mean</td>
<td>SE</td>
<td>(Mean)</td>
<td>Mean</td>
<td>SE</td>
<td>(Mean)</td>
<td>Mean</td>
<td>SE</td>
<td>(Mean)</td>
</tr>
<tr>
<td>Rhamnose</td>
<td>0.1</td>
<td>0.01</td>
<td>0.6</td>
<td>0.6</td>
<td>0.01</td>
<td>NS</td>
<td>0.5</td>
<td>0.5</td>
<td>0.01</td>
<td>NS</td>
<td>NA</td>
<td>NS</td>
</tr>
<tr>
<td>Arabinose</td>
<td>2.5</td>
<td>0.03</td>
<td>3.5</td>
<td>3.9</td>
<td>0.02</td>
<td>***</td>
<td>76.9</td>
<td>74.8</td>
<td>4.17</td>
<td>NS</td>
<td>NA</td>
<td>NS</td>
</tr>
<tr>
<td>Xylose</td>
<td>3.6</td>
<td>0.03</td>
<td>4.0</td>
<td>4.0</td>
<td>0.02</td>
<td>NS</td>
<td>100.3</td>
<td>99.1</td>
<td>3.14</td>
<td>NS</td>
<td>NA</td>
<td>NS</td>
</tr>
<tr>
<td>Mannose</td>
<td>0.6</td>
<td>0.03</td>
<td>1.0</td>
<td>1.0</td>
<td>0.01</td>
<td>NS</td>
<td>0.4</td>
<td>0.4</td>
<td>0.01</td>
<td>NS</td>
<td>NA</td>
<td>NS</td>
</tr>
<tr>
<td>Galactose</td>
<td>82.6</td>
<td>2.50</td>
<td>86.3</td>
<td>85.5</td>
<td>1.17</td>
<td>NS</td>
<td>91.6</td>
<td>89.6</td>
<td>3.04</td>
<td>NS</td>
<td>NA</td>
<td>NS</td>
</tr>
<tr>
<td>Glucose</td>
<td>74.5</td>
<td>1.34</td>
<td>86.7</td>
<td>89.2</td>
<td>0.60</td>
<td>*</td>
<td>95.9</td>
<td>96.5</td>
<td>1.09</td>
<td>NS</td>
<td>NA</td>
<td>NS</td>
</tr>
<tr>
<td>Uronic acids</td>
<td>75.6</td>
<td>3.86</td>
<td>80.7</td>
<td>81.6</td>
<td>2.22</td>
<td>NS</td>
<td>86.4</td>
<td>88.5</td>
<td>4.37</td>
<td>NS</td>
<td>NA</td>
<td>NS</td>
</tr>
<tr>
<td>Total NSP</td>
<td>15.5</td>
<td>0.12</td>
<td>29.6</td>
<td>30.3</td>
<td>0.07</td>
<td>***</td>
<td>14.1</td>
<td>14.8</td>
<td>0.06</td>
<td>NS</td>
<td>NA</td>
<td>NS</td>
</tr>
</tbody>
</table>

NA, not applicable; NS, not significant.

* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.

† For details of diets and procedures, see Tables 1–3 and pp. 580–584.

‡ Values were used for the calculation of the apparent digestibility of carrot NSP, see p. 584.

§ For calculation of intake, faecal excretion and apparent digestibility of carrot NSP monomers, see p. 584.
Table 9. Comparison of the effects of diets containing raw frozen carrots harvested in 1988 and in 1990 on faecal output and on apparent digestibility of non-starch polysaccharide (NSP) monomers by young women†

(Values are standard errors for twelve subjects)‡

<table>
<thead>
<tr>
<th>Faecal excretion</th>
<th>SEM</th>
<th>Statistical significance of differences</th>
<th>Faecal excretion</th>
<th>SEM</th>
<th>Statistical significance of differences</th>
<th>Apparent digestibility</th>
<th>SEM</th>
<th>Statistical significance of differences</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fresh wt</td>
<td>19.19</td>
<td>NS</td>
<td>Rhamnose</td>
<td>0.02</td>
<td>NS</td>
<td>Rhamnose</td>
<td>3.65</td>
<td>NS</td>
</tr>
<tr>
<td>Dry wt</td>
<td>1.51</td>
<td>NS</td>
<td>Arabinose</td>
<td>0.03</td>
<td>NS</td>
<td>Arabinose</td>
<td>0.75</td>
<td>***</td>
</tr>
<tr>
<td>Water</td>
<td>1.82</td>
<td>*</td>
<td>Xylose</td>
<td>0.04</td>
<td>***</td>
<td>Xylose</td>
<td>0.90</td>
<td>NS</td>
</tr>
<tr>
<td>Nitrogen</td>
<td>0.10</td>
<td>NS</td>
<td>Mannose</td>
<td>0.02</td>
<td>NS</td>
<td>Mannose</td>
<td>1.67</td>
<td>NS</td>
</tr>
<tr>
<td>Fat</td>
<td>0.23</td>
<td>*</td>
<td>Galactose</td>
<td>0.03</td>
<td>NS</td>
<td>Galactose</td>
<td>0.68</td>
<td>NS</td>
</tr>
<tr>
<td>Ash</td>
<td>0.29</td>
<td>*</td>
<td>Glucose</td>
<td>0.29</td>
<td>NS</td>
<td>Glucose</td>
<td>2.70</td>
<td>NS</td>
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<tr>
<td>—</td>
<td>—</td>
<td>—</td>
<td>Uronic acids</td>
<td>0.08</td>
<td>NS</td>
<td>Uronic acids</td>
<td>1.04</td>
<td>NS</td>
</tr>
<tr>
<td>—</td>
<td>—</td>
<td>—</td>
<td>Total NSP</td>
<td>0.41</td>
<td>NS</td>
<td>Total NSP</td>
<td>1.29</td>
<td>NS</td>
</tr>
</tbody>
</table>

NS, not significant.
* P < 0.05; ** P < 0.01; *** P < 0.001.
† For details of diets and procedures, see Tables 1–3, and pp. 580–584.
‡ For mean values, see Tables 5–8.
Table 10. Study 1. Total serum cholesterol and high-density-lipoprotein (HDL)-cholesterol concentrations of young women consuming a control diet, or diets containing various processed carrots (harvested in 1988)†

(Values are means and pooled standard errors for ten subjects)

<table>
<thead>
<tr>
<th></th>
<th>Control (Mean)</th>
<th>Raw frozen (Mean)</th>
<th>Blanched frozen (Mean)</th>
<th>Canned (Mean)</th>
<th>SEM</th>
<th>Control v. raw frozen carrots</th>
<th>Raw frozen v. canned carrots</th>
<th>Raw frozen v. blanched frozen carrots</th>
<th>Blanched frozen v. canned carrots</th>
</tr>
</thead>
<tbody>
<tr>
<td>Low-fibre control diet</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Initial</td>
<td>4.8</td>
<td>4.9</td>
<td>4.8</td>
<td>4.7</td>
<td>0.14</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
</tr>
<tr>
<td>Final</td>
<td>4.2</td>
<td>4.2</td>
<td>3.9</td>
<td>4.2</td>
<td>0.11</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
</tr>
<tr>
<td>Difference</td>
<td>0.6</td>
<td>0.7</td>
<td>0.9</td>
<td>0.5</td>
<td>0.15</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
</tr>
<tr>
<td>HDL-cholesterol (mmol/l)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Initial</td>
<td>1.5</td>
<td>1.6</td>
<td>1.5</td>
<td>1.6</td>
<td>0.03</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
</tr>
<tr>
<td>Final</td>
<td>1.4</td>
<td>1.3</td>
<td>1.3</td>
<td>1.3</td>
<td>0.04</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
</tr>
<tr>
<td>Difference</td>
<td>0.1</td>
<td>0.3</td>
<td>0.2</td>
<td>0.3</td>
<td>0.06</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
</tr>
</tbody>
</table>

NS, not significant.

**P < 0.01.

† For details of diets and procedures, see Tables 1–3 and pp. 580–584.
Table 1. Study 2. Total serum cholesterol and high-density-lipoprotein (HDL)-cholesterol concentrations of young women consuming a control diet, or diets containing raw or raw frozen carrots (harvested in 1990)†
(Values are means and pooled standard errors for eleven subjects)

<table>
<thead>
<tr>
<th></th>
<th>Low-fibre control diets</th>
<th>Statistical significance of diet effects</th>
<th>Diets containing carrots</th>
<th>Statistical significance of diet effects</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>RC† (Mean)</td>
<td>RFC§ (Mean)</td>
<td>SEM</td>
<td>Raw (Mean)</td>
</tr>
<tr>
<td>Total serum cholesterol (mmol/l)</td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Initial</td>
<td>4.8</td>
<td>4.9</td>
<td>0.14</td>
<td>NS</td>
</tr>
<tr>
<td>Final</td>
<td>4.8</td>
<td>4.9</td>
<td>0.15</td>
<td>NS</td>
</tr>
<tr>
<td>Difference</td>
<td>0.0</td>
<td>0.0</td>
<td>0.24</td>
<td>NS</td>
</tr>
<tr>
<td>HDL-cholesterol (mmol/l)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Initial</td>
<td>1.7</td>
<td>1.7</td>
<td>0.03</td>
<td>NS</td>
</tr>
<tr>
<td>Final</td>
<td>1.6</td>
<td>1.7</td>
<td>0.04</td>
<td>NS</td>
</tr>
<tr>
<td>Difference</td>
<td>0.0</td>
<td>0.0</td>
<td>0.04</td>
<td>NS</td>
</tr>
</tbody>
</table>

NS, not significant.
† For details of diets and procedures, see Tables 1–3 and pp. 580–584.
‡ Low-fibre control diet preceding diet containing raw carrots.
§ Low-fibre control preceding diet containing raw frozen carrots.
decreases between the different experimental diets. Table 11 shows the cholesterol values measured during the second study. There were no changes in serum total cholesterol and in HDL-cholesterol during the low-fibre control diets or during the carrot-containing diets.

**DISCUSSION**

In this work we investigated whether or not processing of carrots had an influence on various fibre-mediated physiological effects of carrots. For practical reasons (length and number of study periods, keeping quality of raw carrots) the raw carrots in the first study were frozen. The second study was conducted mainly in order to see whether such raw frozen carrots would indeed closely match raw carrots in their physiological effects. We also hoped to reveal any differences in raw frozen carrots which could be attributed to different harvest years.

**Dietary fibre content in carrots and structure of cell walls**

Because processing changed the total dietary fibre content in carrots, different amounts of carrots had to be consumed to achieve equal intakes of fibre. Similarly, as shown in other studies (Anderson & Clydesdale, 1980; Nyman et al. 1987b; Lintas & Cappeloni, 1988), heat treatment affected the chemical composition and the physical structure of the carrot fibre. Both blanching and canning increased the percentage of soluble fibre, probably due to cleavage of glycosidic linkages in polysaccharide chains producing smaller compounds. Freezing had no effect in this respect. These findings corroborate the microscopic examination of the carrot cell walls (C. Schlienger, personal communication). The cell walls of the raw carrots were dense with a very clear middle lamella and cellulose microfibrils adhered to each other. Only very few lesions could be seen. The structure of the raw frozen carrots was similar to that of the raw carrots with some more lesions but no definite breaks in the cell walls. Blanching caused the carrot cell wall to swell. Probably due to the solubilization of pectin, walls split at the level of the middle lamella and intercellular spaces. A dissociation of the walls could be observed, but they did not break up completely. The canned carrots suffered the most important cell lesions. Not only the pectin but also the hemicellulose bridges had been denaturated. The cellulose microfibril group dissociated and swelled. Due to the swelling of the matrix the tissues lost their rigidity. Thus, the canned carrots were much softer compared with blanched and frozen carrots, and especially with raw carrots which were very hard.

**Effect of processing of carrot fibre on fermentation and faecal output**

Several physiological effects of dietary fibre are related to its degradation in the large intestine. The fermentative breakdown of dietary fibre polysaccharides may be of importance because of its impact on energy supply not only to the colonic mucosa (Roediger, 1980) but also for the body as a whole (McNeill, 1984). There are also hypotheses that short-chain fatty acids resulting from the fermentation may affect liver metabolism (Chen et al. 1984) and may play a role in the protection against colonic carcinogenesis due to their contribution to lowering colonic pH (Jacobs, 1990). Fibre fermentation was found to be inversely related to its stool bulking capacity (Stephen & Cummings, 1980a).

In our measurements of faecal saccharides we could not distinguish between unfermented dietary saccharides and those potentially derived from bacterial or endogenous origin. It is difficult to study the basal saccharide excretion on a fibre-free diet in humans as can be done in rats (Nyman et al. 1991). However, studies in man and rats provide evidence that
Fibre in Raw and Processed Carrots

Saccharides of mucins and mucopolysaccharides produced by the host are utilized by the gut microflora (Salyers & McCarthy, 1989). Animal experiments indicate that the contribution of bacteria to faecal NSP may be only small (Nyman & Asp, 1985). In an investigation of the carbohydrate content of faecal bacteria of pigs it was found that NSP accounted for only 4.3% of bacterial dry matter. Glucose (2.36%), galactose (0.96%), rhamnose (0.71%) and uronic acids (0.3%) were the main components (Longland & Low, 1990). In human faeces, up to 55% of faecal dry matter can consist of bacteria (Stephen & Cummings, 1980b). If these bacteria contained similar amounts of NSP to those found in the microflora of pigs (Longland & Low, 1990), in our study they could have contributed a maximum of 0.3, 0.1, 0.1 and 0.05 g of glucose, galactose, rhamnose, and uronic acids respectively to the daily faecal losses of these saccharides, with only negligible differences between the single experimental periods. Therefore we can safely assume that measurements of apparent digestibility of fibre as performed in this study provide relevant information about true fibre fermentation.

The potential susceptibility of dietary fibre to fermentation by bacterial enzymes seems to be dependent on its solubility. Soluble components are more easily degraded than insoluble ones (Cummings, 1984). Provided that the additional consumption of carrot fibre had no effect on the digestibility of fibre in the basal diet (Key & Mathers, 1990), carrot-fibre fermentation may be estimated from differences in the amount of fibre intake and excretion between the carrot-containing diets and the corresponding control diets. According to this calculation, fibres from the different carrots were fermented equally and rather completely (91–94%). The higher proportion of insoluble fibre components in the raw and raw frozen carrots compared with the blanched and canned carrots, and also the more intact cell walls which could be observed microscopically, did not protect them from extensive degradation. The most resistant saccharides were glucose, representing mainly cellulose, and rhamnose, representing the resistant core regions of the carrot pectins.

When similarly processed carrots as used in our studies were fed to rats, fibre fermentability differed from one harvest to the other (Nyman et al. 1991). During one year, blanched carrots were fermented only to 53% compared with raw frozen and cooked carrots which were degraded to 74 and 84% respectively. However, carrots from another year were fermented to a higher degree (83–91%) and were unaffected by processing, as was the case in the present study. In contrast to the studies in rats, no influence of the harvest year on the fermentation of carrot fibre could be observed in our human study involving different harvest years. At such a high fermentation efficiency subtle cell wall differences due to harvest year or processing are indeed not likely to be important. However, when the overall fermentation rates measured in humans are compared with those in rats, it appears that fermentation was slightly more complete in humans.

Faecal bulking capacity of carrot fibre was comparable with the effects of other vegetables (Wisker & Feldheim, 1990) and of finely ground cereal brans (Wisker et al. 1986, 1992), although the carrot fibre was fermented to a higher degree. The increase in stool weight was due to an elevated output both of dry matter and of water. The percentage of faecal water increased with the raw frozen and blanched carrots from the 1988 harvest and with the raw carrots harvested in 1990. This was probably caused by reduced transit times, because under these conditions there might have been less time for water reabsorption (Cummings, 1986). However, transit times were not measured in this study. The higher excretion of faecal N during the consumption of the different carrots may reflect an increase in microbial mass due to the fermentation of the carrot fibre (Stephen & Cummings, 1980b).

There were no differences in the faecal bulking effects between various processed carrots of the same harvest and also no differences between carrots from different harvests,
although there were changes in the chemical composition and in the solubility characteristics of the carrot fibre. This is consistent with the equal fermentability of all carrots found in this human study. In the study with rats (Nyman et al. 1991) the blanched carrots with the lowest fermentability had a greater effect on faecal wet and dry weight compared with raw frozen and cooked carrots. However, this effect could not be repeated when carrots from a different harvest year were studied. Faecal bulking and fermentability were therefore closely related in both humans and in the rat model.

**Serum cholesterol and faecal bile acids**

Highly fermentable isolated fibre sources like pectin have been shown to reduce serum cholesterol levels (Stasse-Wolthuis et al. 1980). A few studies have also reported such an effect after consumption of different vegetables (Gormley et al. 1977, 1979) and carrots (Robertson et al. 1979). The mechanism by which dietary fibre affects the metabolism of serum lipids is not fully understood. A decreased absorption of bile acids because of binding to dietary fibre in the intestinal lumen and the inhibition of hepatic cholesterol synthesis by the products of fermentation, the short-chain fatty acids, are discussed as possible mechanisms (Kritchevsky & Story, 1986).

During the first study the consumption of the experimental diets resulted in a reduction in the concentration of both serum total and HDL-cholesterol. However, because an equal cholesterol lowering could also be observed during the control diet, it could not be attributed to fibre present in the different carrots. Rather the decrease in cholesterol may have been due to differences in the fat content of these subjects' normal diets and the experimental diets consumed in the study. The experimental diets contained less energy in the form of fat (32%) compared with average German diets (42%; Deutsche Gesellschaft für Ernährung, 1988). Changes in the amount of dietary fat can affect serum total cholesterol (Stasse-Wolthuis et al. 1979). This interpretation is supported by the results of the second study, when neither the control diet nor the diets containing raw or raw frozen carrots had any effect on serum total and HDL-cholesterol. However, in this second study the low-fibre basal diet, which preceded the carrot-containing diets in order to avoid differences other than carrots between pre-study diets and experimental diets, did not exert any influence on serum cholesterol.

As our subjects were young women, cholesterol values may have been influenced by the menstrual cycle. However, the fluctuations in HDL- and total cholesterol during a normal menstrual cycle do not apparently show a clear pattern. No changes and a cyclical decrease in cholesterol values have both been reported (Adlercreutz & Tallqvist, 1959; Demacker et al. 1982). In users of oral contraceptives a cyclical decrease in HDL- and total cholesterol was observed (Demacker et al. 1982). Thus, the time of blood sampling during the cycle may be important. During our studies blood samples were taken independently of the menstrual cycle of the subjects. However, it is unlikely that there was a systematic effect of menstrual cycle, because the experimental periods lasted 3 weeks each and were separated from each other by 3 weeks. A decrease of HDL-cholesterol together with total serum cholesterol was also reported when beans (Anderson et al. 1990) or various dietary fibre sources (Stasse-Wolthuis et al. 1979; Kesäniemi et al. 1990) were consumed by male subjects (Anderson et al. 1990; Kesäniemi et al. 1990) or by subjects of both sexes (Stasse-Wolthuis et al. 1979).

The decrease in serum cholesterol during the first study was not connected with increases in faecal bile acid excretion. Faecal bile acids were more dilute after carrots than after the control diet, but the interindividual variation was considerable. Compared with other studies (Schweizer et al. 1983) bile acid losses were rather small, probably due to the low consumption of fat with these diets.
Isolated pectin has been shown to decrease serum cholesterol in several studies. The smallest amount showing a significant effect was a dose of 9 g pectin/d (Stasse-Wolthuis et al. 1980). Pectic substances, measured as uronic acids, contribute about one third to carrot fibre. The carrots consumed in our studies provided daily between 5 and 6 g uronic acids, an amount which was probably too small for a cholesterol-lowering effect. In addition, native pectic substances present in cell walls differ from isolated pectin in respect to molecular weight and number of side chains and therefore may exert different physiological effects than isolated pectin.

Thus, our results differ from those of Robertson et al. (1979) who reported a significant lowering of slightly elevated serum total cholesterol when 200 g raw carrots corresponding to 6 g carrot fibre per day were consumed over 3 weeks in addition to a freely chosen diet. This cholesterol-lowering effect was also accompanied by an increase in faecal bile acid excretion. More in agreement with our findings, the addition of isolated fibres from carrots, cabbage or apples to a controlled diet had no effect on normal serum total cholesterol levels, but the carrot fibre caused a decrease of HDL-cholesterol, which could not be explained (Jenkins et al. 1979).

Conclusions

Our investigations in humans have shown that dietary fibres in carrots are highly fermentable and yet have good stool-bulking ability, comparable to finely milled cereal brans. In spite of appreciable effects of processing, especially of blanching and canning, on the distribution of soluble and insoluble fibre and on texture and microscopic structure of carrots, the physiological effects of these raw or different processed carrots were very similar.

At first sight this finding appears to differentiate root vegetables from cereal fibre sources for which variable physiological effects have been attributed to processing. However, processing effects with cereals have mostly been studied after mechanical or heat treatments at low moisture contents or could in part be explained by formation of resistant starch. In addition, cereal fibres are normally less fermented than vegetable fibres (Wisker et al. 1988; Wisker & Feldheim, 1990) allowing probably a larger effect of processing to be seen than with highly fermentable fibre sources.

Finally, our combined findings from the two studies make it unlikely that carrots can exert a direct fibre-mediated cholesterol-lowering effect.

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