

## Review article

## Dietary nitrate in man: friend or foe?

G. M. McKnight<sup>1\*</sup>, C. W. Duncan<sup>1,2</sup>, C. Leifert<sup>3</sup> and M. H. Golden<sup>1,2†</sup><sup>1</sup>Department of Medicine and Therapeutics, Polwarth Building, Medical School, University of Aberdeen, Foresterhill, Aberdeen AB25 2ZD, UK<sup>2</sup>Boyd Orr Research Centre, Aberdeen Research Consortium, UK<sup>3</sup>Department of Soil Science, University of Aberdeen, Aberdeen AB25 1FX, UK

(Received 6 August 1997 – Revised 16 December 1998 – Accepted 17 December 1998)

Based on the premise that dietary nitrate is detrimental to human health, increasingly stringent regulations are being instituted to lower nitrate levels in food and water. Not only does this pose a financial challenge to water boards and a threat to vegetable production in Northern Europe, but also may be eliminating an important non-immune mechanism for host defence. Until recently nitrate was perceived as a purely harmful dietary component which causes infantile methaemoglobinemia, carcinogenesis and possibly even teratogenesis. Epidemiological studies have failed to substantiate this. It has been shown that dietary nitrate undergoes enterosalivary circulation. It is recirculated in the blood, concentrated by the salivary glands, secreted in the saliva and reduced to nitrite by facultative Gram-positive anaerobes (*Staphylococcus sciuri* and *S. intermedius*) on the tongue. Salivary nitrite is swallowed into the acidic stomach where it is reduced to large quantities of NO and other oxides of N and, conceivably, also contributes to the formation of systemic S-nitrosothiols. NO and solutions of acidified nitrite, mimicking gastric conditions, have been shown to have antimicrobial activity against a wide range of organisms. In particular, acidified nitrite is bactericidal for a variety of gastrointestinal pathogens such as *Yersinia* and *Salmonella*. NO is known to have vasodilator properties and to modulate platelet function, as are S-nitrosothiols. Thus, nitrate in the diet, which determines reactive nitrogen oxide species production in the stomach (McKnight *et al.* 1997), is emerging as an effective host defence against gastrointestinal pathogens, as a modulator of platelet activity and possibly even of gastrointestinal motility and microcirculation. Therefore dietary nitrate may have an important therapeutic role to play, not least in the immunocompromised and in refugees who are at particular risk of contracting gastroenteritides.

## Nitrate: Nitrite: Nitric oxide: Vegetables

Governmental concern about and public fear of nitrate has, over the last 50 years, been the *bête noir* of farmers and water boards, with increasingly stringent regulations being enforced to lower nitrate concentrations in food and water (World Health Organization, 1970, 1984; Department of the Environment, 1984). Green vegetables such as lettuce and spinach, root vegetables such as beetroot, and water, both municipal and well water, are the main sources of nitrate in our diet (Ministry of Agriculture, Fisheries and Food, 1987). In cooler temperate zones where vegetables are grown in low-light-intensity high-temperature glass houses, nitrate levels are much higher than in vegetables grown outside under natural sunlight (Steingrover *et al.* 1993). Moreover,

the nitrate concentration of ground water is gradually increasing through leaching of fertilizer (Addiscot, 1996). The financial burden on water boards attempting to maintain the concentration below the present maximum allowable limit (50 mg/l; World Health Organization, 1984; Ministry of Agriculture, Fisheries and Food, 1987) is becoming onerous, and regulations to limit food nitrate concentrations threaten to stop production of some vegetables in Northern Europe. The regulations are based on the premise that dietary nitrate has only detrimental effects, such as increasing the risk of gastric cancer (Sen *et al.* 1969; Armijo & Coulson, 1975; Cuello *et al.* 1976; National Research Council, 1981; Ohshima & Bartsch, 1981; Correa *et al.* 1990; Rademacher

\* Present address: Department of Clinical Pharmacology, QMW College, Charterhouse Square, London EC1M 6BQ, UK.

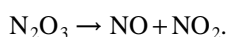
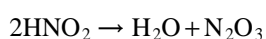
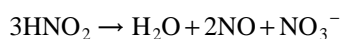
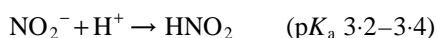
† Corresponding author: Professor Michael Golden, fax +44 (0)1224 699884, email m.golden@abdn.ac.uk

*et al.* 1992; Xu *et al.* 1992) and infantile methaemoglobinaemia (Comly, 1945; Donahoe, 1949), and has no known beneficial effects. However, recent studies show that the postulated risks of nitrate to man, extrapolated from animal studies, have not been substantiated epidemiologically (Armijo *et al.* 1981; Forman *et al.* 1985; Knight *et al.* 1987, 1990; Boeing, 1991; Sobala *et al.* 1991; Pobel *et al.* 1995) and suggest that nitrate may have beneficial effects on the physiology of the intestinal tract (Pique *et al.* 1989; Desai *et al.* 1991; Benjamin *et al.* 1994; Bilski *et al.* 1994; Duncan *et al.* 1994; Lundberg *et al.* 1994; McKnight *et al.* 1997) and that it protects us against food and water-borne pathogens (Dykhuis *et al.* 1996). From cohort mortality studies of healthy eaters (Key *et al.* 1996) and vegetarians (Burr & Sweetnam, 1982) who, by inference, have higher dietary nitrate levels, there appears to be a protective effect of such a diet on ischaemic heart disease.

### Metabolism of dietary nitrate

Understanding the metabolism of ingested nitrate is a prerequisite for evaluation of the hypothesized harmful and beneficial effects. Dietary nitrite is absorbed from the stomach and proximal small intestine into the plasma. It is actively concentrated, by a factor of ten, from the plasma into the saliva (Spiegelhalter *et al.* 1976; Tannenbaum *et al.* 1976; Duncan *et al.* 1995) and then re-secreted into the upper intestinal tract. Approximately 25% of dietary nitrate is thus recirculated into this enterosalivary circulation (Spiegelhalter *et al.* 1976; Tannenbaum *et al.* 1976).

The dorsal surface of the tongue harbours a specialized flora of symbiotic nitrate-reducing facultative anaerobic bacteria which rapidly reduce nitrate to nitrite under hypoxic conditions (Sasaki & Matano, 1979; Duncan *et al.* 1995). Antibiotic inhibition studies show that of the species that reduce nitrite in culture (Li *et al.* 1997), *Staphylococcus sciuri* and *S. intermedius* are likely to be the only organisms of physiological importance *in situ* (C Duncan, C Leifert and M Golden, unpublished results). In rats reared in a germ-free environment, lingual nitrite production from nitrate is absent; this confirms that the nitrate reduction is bacterial rather than an intrinsic mammalian system (Duncan *et al.* 1995). This special flora is induced by a high-nitrate diet and is more abundant under unhygienic husbandry conditions (Duncan, 1998). Thus, from the salivary and dietary nitrate, a high concentration of nitrite is generated in the mouth; this increases after ingestion of nitrate (Lundberg *et al.* 1994; McKnight *et al.* 1997). When swallowed, on exposure to the acidic conditions of the stomach, the nitrite is rapidly protonated to form HNO<sub>2</sub> (acid dissociation constant pK<sub>a</sub> 3.2–3.4). HNO<sub>2</sub>, in turn, decomposes to various nitrogen oxides spontaneously:



Additional reactions, with particularly rapid decomposition, occur in the presence of reducing equivalents, such as

ascorbic acid. Ascorbic acid is actively secreted by the gastric mucosa and is present in vegetables so that it will normally be present with nitrite in the stomach. The chemistry of this process is complex and the relative yields of the different products, under varying conditions of initial concentrations, pH and oxido-reductive potential, have not been determined.

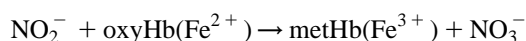
This highly refined and specific mechanism results in the chemical production of NO<sup>•</sup>, and other oxides of N, under acid conditions, in the mouth and upper intestine. The process of recirculation into the saliva has been previously described (Spiegelhalter *et al.* 1976; Tannenbaum *et al.* 1976); however, it was the recognition that high levels of NO were chemically generated in the lumen of the stomach (Benjamin *et al.* 1994; Lundberg *et al.* 1994; McKnight *et al.* 1997) that sparked a resurgence of interest in the role of the enterosalivary circulation of nitrate and the effect of high or low dietary nitrate concentrations on mammalian physiology. NO is a small gaseous molecule which is responsible for cell signalling and host defence in a number of mammalian tissues (Moncada & Higgs, 1993; Angaard, 1994; Brunelli *et al.* 1995; DeGroot *et al.* 1995). It is well established that mammalian cells produce NO enzymically from the amino acid L-arginine with a family of NO synthase enzymes (EC 1.14.13.39) (Moncada & Higgs, 1993). However, the chemical reactions of the upper intestine produce up to 10 000 times the concentrations that occur in tissues from enzymic synthesis (McKnight *et al.* 1997). When the oxides of N are absorbed they react with haemoglobin to regenerate nitrate.

In evolutionary terms, it is unlikely that nitrate would be purposefully concentrated and secreted into the mouth, followed by chemical recycling, if this was actually harmful, for example by causing carcinoma. Such a sophisticated mechanism has surely evolved for a net beneficial purpose.

### Nitrate the foe: potential harmful effects of nitrate

#### *Infantile methaemoglobinaemia*

Initial concern about the safety of nitrate arose in about 1945 when it was recognized that infantile methaemoglobinaemia was associated with the feeding of reconstituted baby food in rural areas where water was drawn from local wells. Some well waters are high in nitrate and this was identified as the usual oxidant causing methaemoglobinaemia (Comly, 1945; Donahoe, 1949). More recently it has been appreciated that infantile methaemoglobinaemia is associated with bacterially contaminated high-nitrate well water (Johnson & Kross, 1990) in which nitrate is reduced to nitrite by bacterial nitrate reductase enzymes. The ingested nitrite load then oxidizes the Fe<sup>2+</sup> of the young infant's oxyhaemoglobin to Fe<sup>3+</sup> of methaemoglobin:



Any oxidants, such as aniline dyes used as colourants or sulfite, may produce methaemoglobinaemia (Howarth, 1951). The condition is now very rare in Europe; sporadic cases do occur, for example, where spinach with its high nitrate content has been cooked at home and then inappropriately stored so that bacterial contamination reduces the nitrate to nitrite (Simon, 1966; Philips, 1971). Nitrate is

much less hazardous than nitrite because the young infant does not develop the dense growth of symbiotic lingual bacteria that reduce nitrate to nitrite until after weaning. Breast-fed infants are not at risk as nitrate is not concentrated into human milk. The age at which the salivary nitrate concentrating mechanism develops has not yet been investigated.

There are several reasons why young infants are more susceptible (Jaffé & Hultquist, 1989) than those in the post-weaning period, to methaemoglobinaemia: (1) young infants have a higher gastric pH ( $\text{pH} > 4$ ) and thus any ingested nitrite is less rapidly degraded in the stomach leaving more for absorption; (2) there are low concentrations of reducing agents such as ascorbic acid secreted by the immature stomach; these are derived in this age group from breast milk; (3) the intakes of water and food-energy are higher relative to body weight (about three times those of an adult); (4) the methaemoglobin reductase enzyme system does not mature until after weaning age; (5) in some races, congenital deficiency of glucose-6-phosphate dehydrogenase (EC 1.1.1.49) in the erythrocyte may lead to a secondary methaemoglobin reductase deficiency.

In particular, children and adults are far less susceptible to methaemoglobinaemia than young infants because of the induction of methaemoglobin reductase during the physiological post-weaning period. Furthermore, methaemoglobinaemia can be treated with reductants such as ascorbic acid (Carnrick *et al.* 1946); presumably a diet with a high ascorbate content will protect older infants and young children from methaemoglobinaemia.

No cases of methaemoglobinaemia have been reported in older children or adults from ingesting well water with a high nitrate concentration. Accidental and occasionally fatal poisoning has occurred where nitrite salts have been ingested (McQuiston & Belf, 1936; Gowans, 1990).

### Carcinogenesis

The second main anxiety about dietary nitrate is that it may be causally associated with gastrointestinal cancer. It has been postulated that when salivary nitrite is swallowed into the stomach the  $\text{HNO}_2$  may nitrosate secondary amines ingested in food to form nitrosamines (Sen *et al.* 1969) some of which are carcinogenic in animal studies (Low, 1974). The nitrosation may also occur exogenously, before food

intake, between nitrite which has been formed from nitrate by contaminating bacteria and amines and amides present in the same food. Many studies have been undertaken in an attempt to establish categorically whether or not nitrate intake *per se* is directly linked to the development of gastrointestinal cancer (Sen *et al.* 1969; Correa *et al.* 1970, 1990; Hill *et al.* 1973; Armijo & Coulson, 1975; Cuello *et al.* 1976; Geleperin *et al.* 1976; Zaldivar, 1977; Amadori *et al.* 1980; Davies, 1980; Juhasz *et al.* 1980; Armijo *et al.* 1981; National Research Council, 1981; Ohshima & Bartsch, 1981; Jensen, 1982; Vincent *et al.* 1983; Beresford, 1985; Forman *et al.* 1985; Knight *et al.* 1987; Boeing, 1991; Sobala *et al.* 1991; Rademacher *et al.* 1992; Xu *et al.* 1992; Gonzalez *et al.* 1994; Pobel *et al.* 1995). The results are conflicting; on balance there does not appear to be a direct causative role of nitrate in gastric cancer (Table 1). On the other hand, the indirect evidence from nutritional and epidemiological studies shows that diets with a high salad or vegetable content, and thus high nitrate content, are protective against some types of cancer, particularly gastric cancer (Haenszel *et al.* 1976; Gonzalez *et al.* 1991, 1994; Corella *et al.* 1996; Key *et al.* 1996).

Nevertheless, the toxicological data unequivocally show that preformed nitrosamines and nitrosamides cause carcinoma in animals (Magee & Barnes, 1967). The question thus arises whether carcinogenic nitrosamines are formed in the human stomach or food in doses sufficient to induce cancer when usual diets are taken. Whilst there is no direct human evidence to support the role of dietary carcinogenic nitrosamines in upper intestinal cancer where such nitrosamines are formed, we should assume that they are carcinogenic in man. This stricture, however, does not hold for nitrate which is not, in itself, carcinogenic. There is also no evidence of direct nitrite (other than through nitrosamine formation) carcinogenicity in animals (National Research Council, 1990).

Much of the recirculated nitrite is temporally separated from ingested amines. After ingestion of food the intragastric pH increases to a level that does not support nitrosamine formation for 0.5–2 h; nitrosation of secondary amines occurs most vigorously at acid pH (2.2–3.5; Bartsch *et al.* 1988). This may also explain why no increase in gastric cancer has been observed in patients on long-term acid suppressant therapy. Moreover, reducing substances such as

**Table 1.** Published studies giving evidence for and against the aetiological role of nitrate in gastric cancer

Studies suggesting a role for nitrate in the aetiology of gastric cancer		Studies giving evidence against a role for nitrate in gastric cancer	
Country	Reference	Country	Reference
UK	Hill <i>et al.</i> (1973)	USA	Geleperin <i>et al.</i> (1976)
Colombia	Correa <i>et al.</i> (1970)	UK	Davies (1980)
Colombia	Cuello <i>et al.</i> (1976)	UK	Beresford (1985)
Chile	Zaldivar (1977)	Chile	Armijo <i>et al.</i> (1981)
Hungary	Juhasz <i>et al.</i> (1980)	France	Vincent <i>et al.</i> (1983)
Italy	Amadori <i>et al.</i> (1980)	UK	Forman <i>et al.</i> (1985)
Denmark	Jensen (1982)	Italy	Knight <i>et al.</i> (1990)
		Spain	Gonzalez <i>et al.</i> (1994)
		France	Pobel <i>et al.</i> (1995)

ascorbic acid, and thiols, that are present in food prevent nitrosation and rapidly reduce nitrite and/or HNO<sub>2</sub> to NO. Indeed, the stomach itself secretes appreciable amounts of ascorbic acid in gastric juice (Sobala *et al.* 1989) which prevents endogenous nitrosamine formation. Even postprandially when the low gastric pH is re-established, nitrosamine formation is not particularly favoured as the reaction is second order with respect to nitrite concentration (Bartsch *et al.* 1988).

Physiologically, during digestion, there are no free amino acids liberated in the stomach so that their amino groups are generally protected within the peptide bond. Pepsin, the major gastric proteinase, only attacks peptide bonds where a tyrosine or phenylalanine provides the amino group; even in the small intestine less than 5% of amino acid residues are in the free state (Nixon & Mawer, 1970).

The most compelling evidence that nitrate, in the concentrations likely to be ingested from a high-nitrate diet, is not causally associated with gastric cancer is the repeatedly confirmed observation that vegetarians, who have three times the nitrate intake of omnivores (Ministry of Agriculture, Fisheries and Food, 1992), and those who have a high intake of fresh fruit and vegetables (and hence nitrate) have lower standardized mortality ratios for gastric cancer (Gonzalez *et al.* 1991; Key *et al.* 1996). Furthermore, whilst the worldwide incidence of gastric cancer is steadily declining (Correa & Chen, 1994), water nitrate content and the intake of green vegetables are both increasing.

#### *Teratogenesis*

As nitrosating substances may affect DNA alkylation and transcription, several groups, primarily Australian, have examined the hypothesis that chronic high nitrate ingestion by a mother may affect the fetus either by germ-cell mutation or by transplacental transmission of tumourigenic nitroso compounds. In animal studies where pregnant hamsters have been fed with large doses of sodium nitrite, an increase of neurogenic tumours has been seen in the fetus; however, such effects are not seen with chronic ingestion of large doses of nitrate (Inui *et al.* 1979). Very large doses of nitrate could cause fetal anoxia where the mother has high levels of methaemoglobin (Geleperin *et al.* 1971). Epidemiological studies in human subjects are equivocal; some workers report an increased incidence of congenital malformations in areas where water nitrate levels are high (Scragg *et al.* 1982; Dorsch *et al.* 1984) and others report a lower incidence in such areas than in the background population (Arbuckle *et al.* 1988). One group has suggested that the incidence of spontaneous abortion is diminished in regions where nitrate is detectable in drinking water (Aschengrau *et al.* 1989). Thus, extrapolation from animal studies suggests that a large dose of unprocessed nitrite may have mutagenic potential in man, but nitrate *per se* appears not to be teratogenic.

#### **Nitrate the friend: potential beneficial effects of nitrate**

##### *Nitrate and nitric oxide*

NO appears to have insinuated itself into most spheres of current research. This small gaseous molecule which readily

crosses membranes is involved in cell signalling (Moncada & Higgs, 1993), is a potent vasodilator (Furchgott & Zawadzki, 1980; Moncada & Higgs, 1993) and platelet aggregation inhibitor (Ignarro, 1989; Radomski *et al.* 1990, 1991), a non-specific acute-phase reactant (Geller *et al.* 1994) and has intrinsic antimicrobial activity (Brunelli *et al.* 1995; DeGroot *et al.* 1995). NO also reacts reversibly with thiols to form S-nitrosothiols (Stamler *et al.* 1992). S-nitrosothiols not only have a longer half-life and greater stability than NO but also have biological activity similar to that of NO (Radomski *et al.* 1992; deBelder *et al.* 1994). It is inherently difficult to separate their biological activities from those of NO *per se*. In general the roles of NO and S-nitrosothiols have only been studied in relation to enzymic synthesis at the cellular level. Our observation (McKnight *et al.* 1997) and that of others (Lundberg *et al.* 1994) that very much larger amounts of NO, which is readily diffusible through membranes, are generated chemically in the lumen of the gut after the oral ingestion of nitrate has led to a search for the physiological role of nitrate in the diet. The highly specialized mechanism of enterosalivary circulation and conservation of ingested nitrate may confer its advantage through the beneficial effects of upper intestinal NO formation which outweigh any theoretical harmful effect. It should be stressed that where nitrite, acid, O and NO coexist, other oxides of N will be present, such as peroxy-nitrite, that themselves have recognized biological roles, especially in their antimicrobial activity (Brunelli *et al.* 1995). The *in vivo* chemistry of nitrogen oxides is undoubtedly highly complex, even chemical modelling of relatively simple systems such as atmospheric nitrogen oxide reactions is far from being fully understood.

##### *A novel mechanism for host defence?*

A common perception of intestinal host defence to pathogens ingested in our food and water, is that it depends entirely on immune factors secreted in the saliva and gastric acidity itself. However, for many gastrointestinal pathogens such as *Campylobacter*, *Shigella* and *Salmonella*, even prolonged immersion in strong acid at pH 2, to emulate the normal fasting stomach, exerts only a bacteriostatic effect (Dykhuisen *et al.* 1996). Failure to kill these organisms means that viable pathogens would pass to the small intestine if the defence depended on acid alone. It has been presumed that for this reason man is susceptible to these organisms; however, for most pathogens the infecting dose is large and only a proportion of those people exposed develop illness. We suggest that acid alone is neither sufficient nor the only physiological mechanism for killing pathogenic bacteria in the upper intestinal tract. The addition of nitrite to acid, in concentrations found in the saliva after a meal containing green leafy vegetables, renders the gastric acid bacteriocidal (Dykhuisen *et al.* 1996). This synergistic enhancement of gastric acid's antibacterial activity by nitrite holds at any given pH, and, at a given pH, increases with the nitrite concentration and time of exposure. The concentrations of nitrite exerting these effects are within the physiological ranges of salivary nitrite (approximately 100 µM–1.5 mM) and gastric acid concentrations (pH 1.5–6.0). For short exposure times stronger acid and increasing nitrite concentrations are

required to ensure that no viable bacteria remain. With *Salmonella typhi*, for example, to achieve a bactericidal effect with acid alone  $\text{pH} < 3$  is required for several hours; as  $\text{pH}$  rises above this then nitrite in increasing concentrations is required to maintain the effect (0.6 mM at  $\text{pH}$  3.5; 1.3 mM at  $\text{pH}$  4.0). Above  $\text{pH}$  5, bacterial kill is achieved only with unphysiologically high ( $> 10$  mM) concentrations of nitrite within a reasonable gastric residence time; although they are killed by exposure to 1.0 mM for 24 h at this  $\text{pH}$ . Organisms exposed to gastric acid obtained endoscopically gave similar results with enhanced kill of organisms when nitrite-containing 'artificial' saliva was added. Alone, gastric acid exerted an antibacterial effect equal to that of acid of the same  $\text{pH}$  with 50  $\mu\text{M}$ -nitrite added (Ndebele, 1996).

In preliminary *in vivo* experiments in healthy volunteers, we have confirmed the *in vitro* results. Live attenuated oral *Salmonella typhi* Ty21a vaccine was enclosed in a short segment of dialysis tubing, impermeable to high-molecular-mass molecules such as immunoglobulins and interferon; the capsule was then swallowed into the stomach on a length of soft dental floss (Fig. 1). After exposure times of 5–20 min the capsules were retrieved, the  $\text{pH}$  was checked to ensure that the capsule had in fact been exposed to gastric contents, and the contents of the dialysis capsule were cultured to assess bacterial survival. The procedure was repeated after administration of a drink containing the amount of nitrate typically found in half a British lettuce, approximately 2 mmol, which caused the expected rise in salivary nitrate and nitrite. In five of six subjects, bacterial survival was shorter after the nitrate drink (Ndebele, 1996).

If these results are confirmed, dietary nitrate will have an important physiological role in augmenting salivary and gastric nitrite to concentrations that reduce the time required to kill ingested pathogens to within the normal residence time of food in the stomach. For food-borne, rather than water-borne pathogens this mechanism is particularly pertinent

because gastric  $\text{pH}$  rises after food ingestion to levels that are not bactericidal unless there is nitrite present. Whilst the  $\text{pH}$  may rise above the critical level for bacterial kill with nitrite present, in fact gastric  $\text{pH}$  recovers to about the  $\text{pK}_a$  of  $\text{HNO}_2$  within 30 min at which point the mechanism we propose for bacteriostasis, and subsequently kill, will come into effect. Food- and water-borne pathogens still represent a significant hazard to health, even in affluent societies, whereas in the developing world they are a major cause of mortality. *Escherichia coli* 0157 is more susceptible to acid plus nitrite than to acid alone, although it is more resistant than the other pathogens that we have tested (Dykhuizen *et al.* 1996). It is of interest that *E. coli* 0157 only emerged as a significant human pathogen after the introduction of regulations to prevent the addition of nitrate to processed meats.

The precise chemical species responsible for microbial damage by acidified nitrite remains unclear. It is clearly a product of protonated nitrite that is formed naturally in the stomach after a nitrate load has been ingested and converted to nitrite on the tongue. The variables of nitrite concentration, acidity, exposure time, presence of reductants, nitrosothiol formation and other anti-bacterial oxidants that are present in saliva (thiocyanate and iodide) will all affect the chemical species formed and the way in which pathogens are affected. It is known that NO complexes with the Fe-S centres of bacterial respiratory enzymes and also directly damages DNA (Wink *et al.* 1991). NO may also combine with superoxide to form peroxynitrite and the hydroxyl radical (Hogg *et al.* 1992) which may be involved in bacterial kill, however, whether such species are formed in the lumen of the intestine is unknown. Moreover,  $\text{HNO}_2$  which is an intermediate in the production of NO, may nitrosate N-, S-, and O-groups on bacterial surface molecules, thus exerting a toxic effect.

Evidence against a direct antimicrobial role of NO in the stomach is our finding that ascorbic acid, a powerful reducing agent which greatly enhances NO production, in fact reduces the antimicrobial activity of acidified nitrite against *Yersinia enterocolitica* (Fite, 1996). Iodide, a naturally-occurring abundant oxidant in saliva, augments the antimicrobial effect of nitrite and acid synergistically. Thiocyanate, which catalyses nitrosation at acidic  $\text{pH}$ , is concentrated in the saliva (Tenuovo & Makinen, 1976), particularly in smokers and those subsisting on plants such as yam and cassava, and dramatically increases the antimicrobial activity of acidified nitrite (Fite, 1996). This synergism may be of particular importance in developing countries where cassava (*Manihot* spp.) is the staple food and hygienic conditions are extremely poor. In such societies the rapid development of a symbiotic nitrate-reducing flora on the tongue and the avoidance of antibiotics at the time of weaning may be critical to prevent 'weanling diarrhoea'.

We suggest that the increased incidence of gastroenteritis associated with the use of gastric acid suppressant agents (Neal *et al.* 1994) is not simply due to the increased gastric  $\text{pH}$  *per se* but more to the effect of the  $\text{pH}$  alteration on the equilibrium of  $\text{HNO}_2$  and the oxides of N which will form far less readily above the  $\text{pK}_a$  of  $\text{HNO}_2$  and will not be available to halt the growth of pathogenic organisms. This provides a physiological explanation for the observation

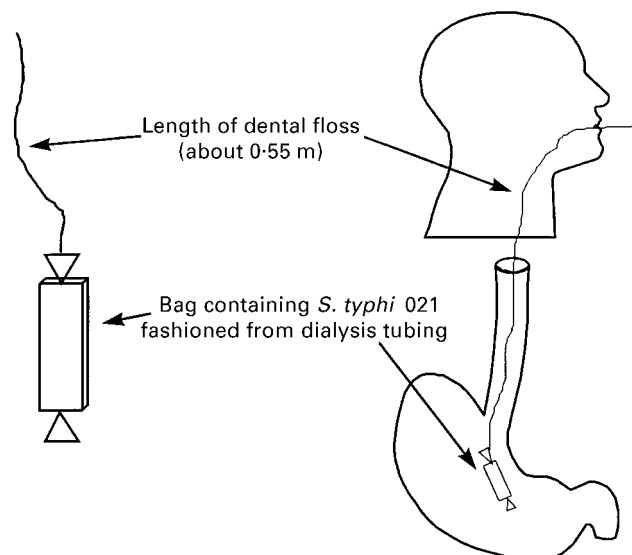


Fig. 1. Schematic representation of the method used to investigate the effect of dietary manipulation on the vaccine strain of *S. typhi* *in vivo*.

that antibiotic treatment predisposes treated individuals to *Salmonella* gastroenteritis (Neal *et al.* 1994), even when the *Salmonella* is sensitive to the antibiotic used, because the antibiotic will kill the symbiotic nitrate-reducing bacteria of the tongue and hence reduce salivary nitrite concentration. Apart from bacteria, yeasts such as *Candida albicans* are also sensitive to acidified nitrite (Dougall *et al.* 1995) and antibiotic treatment greatly increases the incidence of oral thrush (Walker *et al.* 1979); again we suggest that this is due to diminution of salivary nitrite concentration. Furthermore, nitrite-containing toothpaste delays tooth decay (Miller *et al.* 1994) whereas medications inhibiting salivary secretion accelerate tooth decay (McMahon *et al.* 1993; Dasanyake *et al.* 1995).

Many lines of evidence are now coming together to support a beneficial physiological role of dietary nitrate as a potent and essential component of host defence against many of the common pathogens of the gastrointestinal tract.

#### Cardiovascular protection

Many theories have been proffered over the last few decades in an effort to explain the evident cardioprotective effect of diets with a high content of fruit and vegetables. Undoubtedly, the antioxidant vitamins and folic acid that such a diet contains have important effects on the health of the cardiovascular system. However, they fail to explain completely the protection afforded by vegetables (Gaziano *et al.* 1995; Hennekens *et al.* 1995), and more recently phyto-oestrogens have emerged as an additional factor that remains to be fully investigated (Goldin & Gorbach, 1996). Platelet adhesion and aggregation are important in the pathogenesis of vascular occlusive disease (Davies & Thomas, 1981). Both NO and the S-nitrosothiol, S-nitroso-glutathione, are known to be potent inhibitors of platelet aggregation when administered systemically (Radomski *et al.* 1990, 1992; deBelder *et al.* 1994). However, we have shown (McKnight *et al.* 1997) that large quantities of easily diffusible NO gas are generated in the confined lumen of the stomach: a monoepithelialized, vascular viscus. Furthermore, the diet contains thiols which will be readily S-nitrosated in this acidic environment; thus, we would anticipate that absorption of these products would have systemic effects. An attractive hypothesis is that (1) locally, NO gas maintains the gastric microvasculature and (2) systemically, by nitrosating thiol groups which in the acidic stomach will be mainly in the reduced form, it will be absorbed and exert an inhibitory effect on platelet aggregation. Whilst the effect of systemically administered nitrosothiols has been shown (Radomski *et al.* 1992; deBelder *et al.* 1994), until now similar effects have not been shown after administration of oral nitrate. Recently in a small number of individuals ( $n = 6$ ) we have shown by aggregometry Born (1967) that there is inhibition of platelet aggregation following an oral nitrate load which is not seen after taking control anions such as chloride (Fig. 2; G McKnight, N Benjamin and MH Golden, unpublished results). The mechanism of this inhibition has still to be clarified. Nevertheless, these data show that oral nitrate has a rapid and pronounced systemic effect on platelet function which has the potential to prevent thrombotic episodes. We hypothesize that the nitrate content of

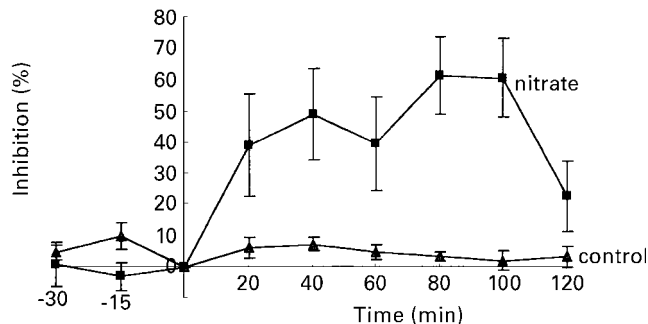


Fig. 2. Inhibition of platelet aggregation by 2 mM-KNO<sub>3</sub> (■) compared with 2 mM-KCl (▲) over a period of 2 h after dose administration.

vegetables actively contributes to their demonstrated cardio-protective properties.

#### Possible gastroprotective effect of high nitrate and/or vegetable intake

Whilst it is clear that NO may damage DNA and cause mutations in human cells *in vitro*, the epidemiological evidence suggests that a high-nitrate diet confers a gastrointestinal protective effect. We speculate that this effect may be partly explained by an enhanced gastric emptying and rapid recovery of gastric acidity associated with the enhanced nitrate content and gastric NO production with high-vegetable diets. However the preservation of the gastric microcirculation by chemically synthesized intra-gastric NO, and hence possible suppression of atrophic gastritis, may also play a part. The possibility that nitrite in the acidic stomach may contribute to the natural defence against *Helicobacter pylori* is under investigation *in vivo*; we have shown that *H. pylori* is susceptible to acidified nitrite *in vitro* (Dykhuisen *et al.* 1998), even in undisturbed biopsies with the mucus layer intact (A Fraser, C Leifert and M Golden, unpublished results). It is intriguing that the incidence of *H. pylori* in the developed world is reducing whilst dietary nitrate and standards of hygiene are increasing (Cave, 1996). Doubtless, gastric cancer is of multifactorial aetiology but a protective, rather than a detrimental, role for nitrate, by suppression of *H. pylori*, is possible. Ironically, as *H. pylori* is the most potent gastric carcinogen commonly present in man, even in terms of gastric cancer, nitrate may turn out to be more friend than foe, despite the theoretical risk of nitrosamine formation.

#### Other gastrointestinal effects

The potential physiological roles of this chemical, rather than enzymic, generation of luminal nitrogen oxides are now open for investigation and speculation. For example, the advice given to patients with irritable bowel syndrome includes instructions to eat more fibre. We speculate that, if this was in the form of green plant leaves with their nitrate-containing matrix capable of generating luminal NO, some amelioration of symptoms of spasm may be experienced. Certainly anecdotal evidence suggests that frequently these

patients' diets are as lacking in fruit and vegetables as in other forms of fibre.

### Conclusion

With the advent of interest in NO, dietary nitrate by way of its sophisticated enterosalivary recirculation and formation of NO and other oxides of N has recently been the subject of renewed scrutiny. Until recently, nitrate in the diet was stigmatized by its chemical relationship to nitrosamines, through nitrite, and there was no appreciation that nitrate in food and the nitrite subsequently formed may be qualitatively different from directly administered nitrite or preformed contaminating nitrite in stored foods. With increasing recognition that dietary nitrate and its recirculation may have evolved to confer an evolutionary advantage by way of provision of a non-immune mechanism of host defence against ingested gastrointestinal pathogens, modulation of microcirculation, platelet activity and possibly even gastrointestinal motility, nitrate should be liberated from the stigma of association with nitrosamine chemistry.

We envisage a future where antibiotic therapy may not be the treatment of choice for intestinal infection and where increased understanding of the luminal metabolism of components of the diet and of saliva will lead to interventions that will prevent illness of the mouth, stomach, intestine and cardiovascular system, and probiotic therapy with organisms active in the upper intestinal lumen will assume a therapeutic role. This would have implications for the control of antibiotic resistance and, at least initially, would be most useful in animal husbandry where antibiotic resistance is a major problem. In man the therapeutic importance of this mechanism in preventing oral and gastrointestinal infection may be particularly important in the immunocompromised and those who are unavoidably exposed to contaminated food and water. For example, the diets used to sustain refugees are devoid of nitrate and thiocyanate whilst epidemic gastrointestinal infection devastates the camps and there are no current affordable solutions being investigated. Before nitrate and probiotic therapy can become reality, the understanding of basic physiological N chemistry needs to be more profound, the active transporter at the salivary gland must be further characterized, along with the kinetics of the microbial nitrate reductase enzyme. The role of this system in the weanling child and the safety of nitrate at this age also have to be established. This knowledge should then provide the basis for entirely new therapeutic interventions and very cheap preventative strategies easily transferred to the impoverished areas of the Third World where intestinal infection is a common cause of death. This knowledge may also lead to a much needed re-evaluation of the limitations on nitrate intake and hence increase agricultural production and redirect funds into water management.

### Acknowledgements

We thank Alemu Fite and Nyaladzi Ndebele for making their unpublished data available and Nigel Benjamin, Lorna Smith, Rolf Dykhuizen and Hamish McKenzie for their contribution to the work of our group that is reported in this review. The work was supported by the Ministry of Agriculture, Fisheries and Food.

### References

- Addiscott TM (1996) Fertilizers and nitrate leaching. In *Agricultural Chemicals and the Environment, Issues in Environmental Science and Technology*, no. 5, pp. 1–26 [RE Hester and RM Harrison, editors]. Cambridge: Royal Society of Chemistry.
- Amadori D, Ravaioli A, Gardini A, Liverani M, Zoli W, Tonelli B, Ridolfi R & Gentilini P (1980) N-nitroso compound precursors and gastric cancer: preliminary data of a study on a group of farm workers. *Tumori* **66**, 145–152.
- Angaard E (1994) Nitric oxide: mediator, murderer and medicine. *Lancet* **343**, 1199–1206.
- Arbuckle TE, Sherman GJ, Corey PN, Walters D & Lo B (1988) Water nitrates and CNS birth defects; a population-based case-control study. *Archives of Environmental Health* **43**, 162–167.
- Armijo R & Coulson AH (1975) Epidemiology of stomach cancer in Chile: the role of nitrogen fertilizers. *International Journal of Epidemiology* **4**, 301–309.
- Armijo R, Gonzales A, Orellana M, Coulson AH, Sayre JW & Detels R (1981) Epidemiology of gastric cancer in Chile II. Nitrate exposures and stomach cancer frequency. *International Journal of Epidemiology* **10**, 57–62.
- Aschengrau A, Zierler S & Cohen A (1989) Quality of community water and the occurrence of spontaneous abortion. *Archives of Environmental Health* **44**, 283–290.
- Bartsch H, Ohshima H & Pignatelli B (1988) Inhibitors of endogenous nitrosation. Mechanisms and implications in human cancer prevention. *Mutation Research* **202**, 307–324.
- Benjamin N, O'Driscoll F, Dougall H, Duncan C, Smith L, Golden M & McKenzie H (1994) Stomach NO synthesis. *Nature* **368**, 502.
- Beresford SA (1985) Is nitrate in the drinking water associated with the risk of cancer in the urban UK? *International Journal of Epidemiology* **14**, 57–63.
- Bilski J, Konturek PC, Konturek SJ, Cieszkowski M & Czarnobilski K (1994) Role of endogenous nitric oxide in the control of gastric acid secretion, blood flow and gastrin release in conscious dogs. *Regulatory Peptides* **54**, 175–184.
- Boeing H (1991) Epidemiological research in stomach cancer: progress over the last ten years. *Journal of Cancer Research and Clinical Oncology* **47**, 133–143.
- Born GVP (1967) Quantitative investigations into the aggregation of blood platelets. *Journal of Physiology* **162**, 67P.
- Brunelli L, Crow JP & Beckmann JS (1995) The comparative toxicity of nitric oxide and peroxy-nitrite to *Escherichia coli*. *Archives of Biochemistry and Biophysics* **316**, 327–334.
- Burr ML & Sweetnam PM (1982) Vegetarianism, dietary fiber, and mortality. *American Journal of Clinical Nutrition* **36**, 873–877.
- Carnrick M, Plois BD & Klein T (1946) Methaemoglobinaemia: treatment with ascorbic acid. *Archives of Internal Medicine* **78**, 296–302.
- Cave DR (1996) Transmission and epidemiology of *Helicobacter pylori*. *American Journal of Medicine* **100**, 12S–17S.
- Comly HH (1945) Cyanosis in infants caused by nitrates in well water. *Journal of the American Medical Association* **129**, 112–116.
- Corella D, Cortina P, Guillen M & Gonzalez JI (1996) Dietary habits and geographic variation in stomach cancer mortality in Spain. *European Journal of Cancer Prevention* **5**, 249–257.
- Correa P & Chen VW (1994) Gastric cancer. *Cancer Surveys* **19**–**20**, 55–76.
- Correa P, Cuello C & Duque E (1970) Carcinoma and intestinal metaplasia of the stomach in Colombian migrants. *Journal of the National Cancer Institute* **44**, 297–306.
- Correa P, Haenszel W, Cuello C, Zavala D, Fontham E, Zarama G, Tannenbaum S, Collazos T & Ruiz B (1990) Gastric precancerous process in a high risk population: cross-sectional studies. *Cancer Research* **50**, 4731–4736.

- Cuello C, Correa P, Haenszel W, Gordillo G, Brown C, Archer M & Tannenbaum S (1976) Gastric cancer in Colombia. I. Cancer risk and suspected environmental agents. *Journal of the National Cancer Institute* **57**, 1015–1020.
- Dasanayake AP, Roseman JM, Caulfield PW & Butts JI (1995) Distribution and determinants of *mutans streptococci* among African-American children and association with selected variables. *Pediatric Dentistry* **17**, 192–198.
- Davies JM (1980) Stomach cancer mortality in Worksop and other Nottinghamshire mining towns. *British Journal of Cancer* **41**, 438–445.
- Davies MJ & Thomas T (1981) The pathological basis and microanatomy of occlusive thrombus formation in human coronary arteries. *Philosophical Transactions of the Royal Society of London, Series B: Biological Sciences* **294**, 225–229.
- de Belder AJ, MacAllister R, Radomski MW, Moncada S & Vallance PJ (1994) Effects of S-nitroso-glutathione in the human forearm circulation: evidence for selective inhibition of platelet activation. *Cardiovascular Research* **28**, 691–694.
- DeGroot MA, Granger D, Xu Y, Campbell G, Prince P & Fang FC (1995) Genetic and redox determinants of nitric oxide cytotoxicity in a *Salmonella typhimurium* model. *Proceedings of the National Academy of Sciences USA* **92**, 6399–6403.
- Department of the Environment (1984) *Nitrates in Water Supplies*. DOE Press Notice 180, 9 April 1984.
- Desai KM, Sessa WC & Vane JR (1991) Involvement of nitric oxide in the reflex relaxation of the stomach to accommodate food or fluid. *Nature* **351**, 477–479.
- Donahoe WE (1949) Cyanosis in infants with drinking water as a cause. *Paediatrics* **3**, 308–311.
- Dorsch MM, Scragg RKR, McMichael AJ, Baghurst PA & Dyer KF (1984) Congenital malformations and maternal drinking water supply in rural South Australia: a case control study. *American Journal of Epidemiology* **119**, 473–486.
- Dougall HT, Smith L, Duncan C & Benjamin N (1995) The effect of amoxicillin on salivary nitrite concentrations: an important mechanism of adverse reactions? *British Journal of Clinical Pharmacology* **39**, 460–462.
- Duncan C, Dougall H, Johnston P, Green S, Brogan R, Leifert C, Smith LM, Golden M & Benjamin N (1995) Chemical generation of nitric oxide in the mouth from the enterosalivary circulation of dietary nitrate. *Nature Medicine* **1**, 546–551.
- Duncan CW (1999) Nitrate to nitrite conversion on mammalian tongues. PhD Thesis, University of Aberdeen.
- Dykhuizen RS, Fraser A, McKenzie H, Golden M, Leifert C & Benjamin N (1998) *Helicobacter pylori* is killed by nitrite under acidic conditions. *Gut* **42**, 334–337.
- Dykhuizen RS, Frazer R, Duncan C, Smith CC, Golden M, Benjamin N & Leifert C (1996) Antimicrobial effect of acidified nitrite on gut pathogens: importance of dietary nitrate in host defence. *Antimicrobial Agents and Chemotherapy* **40**, 1422–1425.
- Fite A (1996) Modification of antimicrobial activity of acidified nitrite against *Yersinia enterocolitica* by physiologically active chemicals and hydroxyacids. MSc Thesis (Nutrition and Metabolism), University of Aberdeen.
- Forman D, Al-Dabbagh SA & Doll R (1985) Nitrates, nitrites and gastric cancer in Great Britain. *Nature* **313**, 620–625.
- Furchgott RF & Zawadzki JV (1980) The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature* **288**, 373–376.
- Gaziano JM, Manson JE, Branch LG, Colditz GA, Willett WC & Buring JE (1995) A prospective study of consumption of carotenoids in fruits and vegetables and decreased cardiovascular mortality in the elderly. *Annals of Epidemiology* **5**, 255–260.
- Geller DA, Freeswick PD, Nguyen D, Nussler AK, Di Silvio M, Shapiro RA, Way SC, Simmons RL & Billiar TP (1994) Differential induction of nitric oxide synthase in hepatocytes during endotoxaemia and the acute phase response. *Archives of Surgery* **129**, 169–171.
- Geleperin A, Jacobs EE & Kletke LS (1971) The development of methaemoglobin in mothers and newborn infants from nitrate in water supplies. *Illinois Medical Journal* **140**, 42–44, 82.
- Geleperin A, Moses VJ & Fox G (1976) Nitrate in water supplies and cancer. *Illinois Medical Journal* **149**, 251–253.
- Goldin BR & Gorbach SL (1996) Phytoestrogens: possible role in preventing human disease. *Nutrition* **12**, 216–217.
- Gonzalez CA, Riboli E, Badosa J, Batiste E, Cardona T, Pita S, Sanz JM, Torrent M & Agudo A (1994) Nutritional factors and gastric cancer in Spain. *American Journal of Epidemiology* **139**, 466–473.
- Gonzalez CA, Sanz JM, Marcos G, Pita S, Brullet E, Saigi E, Badia A & Riboli E (1991) Dietary factors and stomach cancer in Spain: a multi-centre case-control study. *International Journal of Cancer* **49**, 513–519.
- Gowans WJ (1990) Fatal methaemoglobinaemia in a dental nurse. A case study of sodium nitrite poisoning. *British Journal of General Practice* **40**, 470–471.
- Haenszel W, Correa P, Cuello C, Guzman N, Burbano LC, Lores H & Munoz J (1976) Gastric cancer in Colombia II. Case control epidemiologic study of precursor lesions. *Journal of the National Cancer Institute* **57**, 1021–1026.
- Hennekens CH, Gaziano JM, Manson JE & Buring JE (1995) Antioxidant vitamin-cardiovascular disease hypothesis is still promising, but still unproven: the need for randomized trials. *American Journal of Clinical Nutrition* **62**, 1377S–1380S.
- Hill MJ, Hawksworth GM & Tattersall G (1973) Bacteria, nitrosamines and cancer. *British Journal of Cancer* **28**, 562–567.
- Hogg N, Darley-Usmar VM, Wilson MT & Moncada S (1992) Production of hydroxyl radicals from the simultaneous generation of superoxide and nitric oxide. *Biochemical Journal* **281**, 419–424.
- Howarth BE (1951) Epidemic of aniline methaemoglobinaemia in newborn babies. *Lancet* **ii**, 934–935.
- Ignarro LJ (1989) Endothelium-derived nitric oxide: actions and properties. *FASEB Journal* **3**, 31–36.
- Inui N, Yoshisuke N, Taketomi M & Mori M (1979) Transplacental action of sodium nitrite on embryonic cells of Syrian Golden Hamster. *Mutation Research* **66**, 149–158.
- Jaffé ER & Hultquist DE (1989) Cytochrome b5 reductase deficiency and enzymopenic hereditary methemoglobinemia. In *The Metabolic Basis of Inherited Disease*, 6th ed., vol. 2, pp. 2267–2280 [CR Scriver, AL Beaudet WS Sly and D Valle, editors]. New York, NY: McGraw Hill.
- Jensen OM (1982) Nitrate in drinking water and cancer in northern Jutland, Denmark, with special reference to stomach cancer. *Ecotoxicology and Environmental Safety* **6**, 258–267.
- Johnson CJ & Kross BC (1990) Continuing importance of nitrate contamination of groundwater and wells in rural areas. *American Journal of Industrial Medicine* **18**, 449–456.
- Juhász L, Hill MJ & Nagy G (1980) Possible relationship between nitrate in drinking water and incidence of stomach cancer. *IARC Scientific Publications* **31**, 619–623.
- Key TJA, Thorogood M, Appleby PN & Burr ML (1996) Dietary habits and mortality in 11 000 vegetarians and health conscious people: results of a 17 year follow up. *British Medical Journal* **313**, 775–778.
- Knight TM, Forman D, Al-Dabbagh SA & Doll R (1987) Estimation of dietary intake of nitrate and nitrite in Great Britain. *Food and Chemical Toxicology* **25**, 277–285.
- Knight TM, Forman D, Piratsu R, Comba P, Iannarilli R, Cocco PL, Angotzi G, Ninu E & Schierano S (1990) Nitrate and nitrite



- exposure in Italian populations with different gastric cancer rates. *International Journal of Epidemiology* **19**, 510–515.
- Li H, Duncan C, Townsend K, Killum K, Smith L, Johnson P, Dykhuizen R, Kelly D, Golden MH, Benjamin N & Leifert C (1997) Nitrate-reducing bacteria on rat tongues. *Applied and Environmental Microbiology* **63**, 924–930.
- Low H (1974) Nitroso compounds: safety and public health. *Archives of Environmental Health* **29**, 256–260.
- Lundberg JON, Weitzberg E, Lundberg JM & Alving K (1994) Intra-gastric nitric oxide production in humans: measurements in expelled air. *Gut* **35**, 1543–1546.
- McKnight G, Smith LM, Drummond RS, Duncan CW, Golden MNH & Benjamin N (1997) The chemical synthesis of nitric oxide in the stomach from dietary nitrate in man. *Gut* **40**, 211–214.
- McMahon J, Parnell WR & Spears GF (1993) Diet and dental caries in preschool children. *European Journal of Clinical Nutrition* **47**, 794–802.
- McQuiston TAC & Belf MB (1936) Fatal poisoning by sodium nitrite. *Lancet* **ii**, 1153–1154.
- Magee PN & Barnes JM (1967) Carcinogenic nitroso compounds. *Advances in Cancer Research* **10**, 163–246.
- Miller S, Gaffar A, Sullivan R, Heu R, Truong T & Stronick M (1994) Evaluation of a new dentrifice for the treatment of sensitive teeth. *Journal of Clinical Dentistry* **5**, 71–79.
- Ministry of Agriculture, Fisheries and Food (1987) *Nitrate, Nitrite and N-nitroso Compounds in Food – 7. Food Surveillance Paper* no. 20. London: H. M. Stationery Office.
- Ministry of Agriculture, Fisheries and Food (1992) *Nitrate, Nitrite and N-nitroso Compounds in Food: Second Report*. London: H. M. Stationery Office.
- Moncada S & Higgs A (1993) The L-arginine–nitric oxide pathway. *New England Journal of Medicine* **329**, 2002–2012.
- National Research Council (1981) *The Health Effects of Nitrate, Nitrite and N-nitroso Compounds: Part 1 of a 2 Part Study by the Committee on Nitrite and Alternative Curing Agents in Food*. Washington, DC: National Academy Press.
- National Research Council (1990) *Implications for Reducing Chronic Disease Risk*, pp. 475–476. Washington, DC: National Academy Press.
- Ndebele N (1996) The *in vivo* sensitivity of *Salmonella typhi* TY21A to acidified nitrite in the stomach. MSc Thesis (Nutrition and Metabolism), University of Aberdeen.
- Neal RN, Brijj SO, Slack RCB, Hawkey CJ & Logan RFA (1994) Recent treatment with H<sub>2</sub> antagonists and antibiotics and gastric surgery as risk factors for salmonella infection. *British Medical Journal* **308**, 176.
- Nixon SE & Mawer GE (1970) The digestion and absorption of protein in man 2. The form in which digested protein is absorbed. *British Journal of Nutrition* **24**, 241–258.
- Ohshima H & Bartsch H (1981) Quantitative estimation of endogenous nitrosation in humans by monitoring *N*-nitrosoproline excreted in urine. *Cancer Research* **41**, 3658–3663.
- Philips WE (1971) Naturally occurring nitrate and nitrite in foods in relation to infant methaemoglobinaemia. *Food and Cosmetics Toxicology* **9**, 219–228.
- Pique JM, Whittle BJR & Esplugues JV (1989) The vasodilator role of endogenous nitric oxide in the rat gastric microcirculation. *European Journal of Pharmacology* **174**, 293–296.
- Pobel D, Riboli E, Cornee J, Hemon B & Guyader M (1995) Nitrosamine, nitrate and nitrite in relation to gastric cancer: a case control study in Marseille, France. *European Journal of Epidemiology* **11**, 67–73.
- Rademacher JJ, Young TB & Kanarek MS (1992) Gastric cancer mortality and nitrate levels in Wisconsin drinking water. *Archives of Environmental Health* **47**, 292–294.
- Radomski MW, Palmer RMJ & Moncada S (1990) An L-arginine/nitric oxide pathway present in human platelets regulates aggregation. *Proceedings of the National Academy of Sciences USA* **87**, 5193–5197.
- Radomski MW, Palmer RM & Moncada S (1991) Modulation of platelet aggregation by an L-arginine–nitric oxide pathway. *Trends in Pharmacological Sciences* **12**, 87–88.
- Radomski MW, Rees DD, Dutra A & Moncada S (1992) S-nitrosoglutathione inhibits platelet activation *in vitro* and *in vivo*. *British Journal of Pharmacology* **107**, 745–749.
- Sasaki T & Matano K (1979) Formation of nitrite from nitrate at the dorsum linguae. *Journal of the Food Hygiene Society of Japan* **20**, 363–369.
- Scragg RKR, Dorsch MM, McMichael AJ & Baghurst PA (1982) Birth defects and household water supply. Epidemiological studies in the Mount Gambier region of south Australia. *Medical Journal of Australia* **2**, 577–579.
- Sen NP, Smith DC & Schwinghamer L (1969) Formation of *N*-nitrosamines from secondary amines and nitrate in human and animal gastric juice. *Food and Cosmetics Toxicology* **7**, 301–307.
- Simon C (1966) L'intoxication par les nitrites après ingestion d'épinards (Poisoning by nitrites after ingestion of spinach). *Archives Françaises de Pédiatrie* **23**, 231–238.
- Sobala GM, Pignatelli B, Schorah CJ & Bartsch H (1991) Levels of nitrite, nitrate, *N*-nitroso compounds, ascorbic acid and total bile acids in gastric juice of patients with and without precancerous conditions of the stomach. *Carcinogenesis* **12**, 193–198.
- Sobala GM, Schorah CJ, Sanderson M, Dixon MF, Tompkins DS, Godwin P & Axon AT (1989) Ascorbic acid in the human stomach. *Gastroenterology* **97**, 257–363.
- Spiegelhalter B, Eisenbrand G & Preussmann R (1976) Influence of dietary nitrate on nitrite content of human saliva: possible relevance to *in vivo* formation of *N*-nitroso compounds. *Food and Cosmetics Toxicology* **14**, 545–548.
- Stamler JS, Simon DI, Osborne JA, Mullins ME, Jaraki O, Michel T, Singel DJ & Loscalzo J (1992) S-nitrosylation of proteins with nitric oxide: synthesis and characterization of biologically active compounds. *Proceedings of the National Academy of Sciences USA* **89**, 444–448.
- Steingrover EG, Steenhuizen JW & Vanderboon J (1993) Effects of low-light intensities on nitrate accumulation in lettuce grown on a recirculating nutrient solution. *Netherlands Journal of Agricultural Science* **41**, 13–21.
- Tannenbaum SR, Weisman M & Fett D (1976) The effect of nitrate intake on nitrite formation in human saliva. *Food and Cosmetics Toxicology* **14**, 549–552.
- Tenuovo J & Mäkinen KK (1976) Concentration of thiocyanate and ionizable iodine in saliva of smokers and nonsmokers. *Journal of Dental Research* **55**, 661–663.
- Vincent P, Dubois G & Leclerc H (1983) Nitrates in the drinking water and cancer mortality. An epidemiologic study in northern France. *Revue d'Epidémiologie et de Santé Publique* **31**, 199–207.
- Walker AM, Jick H & Porter J (1979) Drug-related superinfection in hospitalised patients. *Journal of the American Medical Association* **242**, 1273–1275.
- Wink DA, Kasprzak KS, Maragos CM, Elespuru RK, Misra M, Dunams TM, Cebula TA, Koch WH, Andrews AW, Allen JS & Keefer LK (1991) DNA deaminating ability and genotoxicity of nitric oxide and its progenitors. *Science* **254**, 1001–1003.
- World Health Organization (1970) *European Standards for Drinking-Water*. Geneva: WHO.
- World Health Organization (1984) *Guidelines for Drinking-Water Quality*. Vol. 1. *Recommendations*. Geneva: WHO.
- Xu G, Song P & Reed PI (1992) The relationship between gastric

mucosal changes and nitrate intake via drinking water in a high-risk population for gastric cancer in Moping County, China. *European Journal of Cancer Prevention* **1**, 437–443.

Zaldivar R (1977) Epidemiology of gastric and colorectal cancer in the United States and Chile with particular reference to the role

of dietary and nutritional variables, nitrate fertilizer pollution, and *N*-nitroso compounds. *Zentralblatt für Bacteriologie I Abt Originale B: Hygiene, Krankenhaushygiene, Betriebshygiene Preventive Medizin* **164**, 193–217.

© Nutrition Society 1999

---