Quantitative significance of measuring trimethylselenonium in urine for assessing chronically high intakes of selenium in human subjects†

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The purpose of the present study was to investigate the effects of Se restriction on the excretion of Se in men who had consumed high levels of this element during their entire lives. With the use of stable isotopes of Se as selenite, the excretion of methylated Se in urine was investigated in Chinese men (n 10) who had habitual chronic high intakes of this element. The relationship between either urine Se or trimethylselenonium (TMSe) to the estimated long-term Se intake was not linear over the entire range of intake, which was also true for the infusion of labelled selenite. A non-linear relationship was also found between urine TMSe and urine Se both for TMSe arising from catabolism of endogenous body Se and that from infused selenite. The data suggest a close precursor–product relationship of urine Se and its TMSe component based on the nearly identical specific activities for these two selenocompounds. Although dimethylselenide in breath was not measured in the present study, combining urinary TMSe with this breath test may be more useful in the assessment of long-term Se status.

Selenium: Chinese men: Urine: Stable isotopes

Little accurate information is available on the relationship between long-term Se intake in different chemical forms, the resultant body status of Se, and potential indices that could be used to monitor Se status. The ability to monitor Se status during long-term supplementation is important because of the need to monitor biological efficacy and concerns for toxicity. Since much of total body Se is present as selenomethionine (Semet) in body proteins (Sunde, 1990), measurement of an index related to total body Se would not be useful as the required index. In contrast, an index related quantitatively to whole body flux of Se2+ (Fig. 1) may provide the needed information. Of the various selenocompounds that may be available in human subjects by non-invasive sampling, hair and toenails may be the least useful indicators of body status of biologically active Se because a large fraction of their Se content may be from non-specific incorporation of dietary Semet in their structural proteins (Fig. 1). Of the three selenocompounds present in plasma (Deagen et al. 1993) (albumin-bound Se, glutathione peroxidase (EC 1.11.1.9), and selenoprotein P) albumin-bound Se is also not likely to be a useful indicator of body status of biologically active Se because the major portion of its Se has been shown to be in the form of Semet (Deagen et al. 1991), while the Se content of glutathione peroxidase and selenoprotein P is saturable over the nutritional range of intake (Combs & Combs, 1986; Burk et al. 1991; Burk & Hill, 1994) and is not likely to respond to the wide range of Se intake involved in long-term supplementation. On the other hand, both urine Se and its methylated derivative(s) may have potential as quantitatively useful indices of biologically active Se (Ip et al. 1991; Vadhanavikit et al. 1993). However, for either urine Se, or any methylated derivatives excreted in urine to be able to function as quantitative indices of biologically active body Se, their excretion must bear a quantitative relationship to whole body flux of Se2+ over the applicable range of long-term Se supplementation. In order to understand the relationship between Se status and disposition of Se2+ where the body’s capacity for its incorporation has been exceeded, the present investigation was undertaken to study the excretion of trimethylselenonium (TMSe) in men who had consumed excess Se their entire lives. The experimental data presented

Abbreviations: SA, specific activity; Semet, selenomethionine; TMSe, trimethylselenonium.

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in this manuscript provide some initial insight into this important relationship.

Experimental methods

Protocol

Ten adult male volunteers who had been life-long residents of the village of Luojiaba (Jianshi County, Hubei Province, People’s Republic of China; estimated daily Se intake: 197–1230 μg/d; Table 1) were recruited for this investigation. A description of the people living in this high-Se area has been presented previously (Xia et al. 1992). The protocol was approved by the Oregon State University Committee for the Protection of Human Subjects and by a special Institutional Review Board convened at the Chinese Academy of Preventive Medicine in Beijing, and the subjects signed informed consent before participation in the study. Each subject was paid an equivalent of US$300 for participation in this study. After baseline samples were obtained (day 0), informed consent before participation in the study. Each subject was paid an equivalent of US$300 for participation in this study. After baseline samples were obtained (day 0), the men were moved to Changping village (Lichuan County, Hubei Province), approximately 300 km from their native village, where they stayed for 70 d. The latter village is in the Keshan disease area, which during 1969–83 had incidence and mortality rates of 103 per 100 000 and 45.2 % respectively. These rates were the highest recorded among villages in Lichuan County for the period. During their 70 d stay, they consumed locally grown food providing 43.3 μg Se/d (range 30–45 μg Se/d) based on 3 d dietary survey. Anthropometric and Se baseline data for the subjects are summarized in Table 1.

On day 1 of the protocol, each subject was infused with 105 μg Se labelled with the stable isotope 74Se which was prepared specifically for intravenous administration in the form of selenite (see later), by adding the stable isotope solution to a 500 ml bag of intravenous glucose solution (50 g/l). Infusion rate was adjusted (by gravity) to last for 5 h (07.00–12.00 hours). Complete 24 h urine collections were made daily for the first 7 d after infusion, and at days 22, 43, and 62 afterwards. An identical infusion procedure was repeated on day 64, with the exception that the total amount of Se infused was 113 μg. Similarly to the first infusion period, 24 h urine collections were made daily for 7 d following the second infusion at 64 d. The subjects were closely monitored by the medical and research staff during the course of the investigation.

Preparation of stable isotope for infusion

Elemental Se (13.6 mg) enriched with respect to 74Se (74Se 54.4 % by weight; 77Se 4.406 % by weight; R74/77 12.35 % by weight; the corresponding values for unenriched Se were 0.815, 7.38 and 0.110 % respectively) was dissolved in 2–3 ml 16 M-HNO3. The resulting solution was heated gently until almost dry. The remaining liquid was transferred with deionized water into an acid-washed plastic container and volume adjusted with deionized water. This solution had a concentration of 99.83 μg Se/ml and pH of 3.0. A measured portion of this solution was prepared for infusion as follows. Solution pH was adjusted with NaOH to 7.40 and diluted with saline solution (9 g/l). It was then filtered through a 0.45 μm filter and 2.0 ml portions dispensed into sterile glass ampoules. All ampoules were then sterilized by autoclaving at 120° for 20 min. Three of the ampoules were analysed for Se (52.5 μg/2 ml ampoule). Freedom from pyrogenicity was tested in five rabbits with three randomly selected ampoules using standard

Table 1. Anthropometric and baseline selenium variables for the male subjects (n 10)

<table>
<thead>
<tr>
<th>Subject no.</th>
<th>Age (years)</th>
<th>Weight (kg)</th>
<th>Packed cell volume (%)</th>
<th>Estimated long-term Se intake* (μg/d)</th>
<th>Urine Se (μg/d)</th>
<th>Plasma Se (ng/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>50</td>
<td>67</td>
<td>45.7</td>
<td>507</td>
<td>356</td>
<td>379</td>
</tr>
<tr>
<td>2</td>
<td>45</td>
<td>69</td>
<td>44.4</td>
<td>363</td>
<td>199</td>
<td>309</td>
</tr>
<tr>
<td>3</td>
<td>49</td>
<td>65</td>
<td>47.7</td>
<td>439</td>
<td>325</td>
<td>347</td>
</tr>
<tr>
<td>4</td>
<td>19</td>
<td>59</td>
<td>43.9</td>
<td>402</td>
<td>218</td>
<td>329</td>
</tr>
<tr>
<td>5</td>
<td>44</td>
<td>64</td>
<td>46.2</td>
<td>358</td>
<td>255</td>
<td>306</td>
</tr>
<tr>
<td>6</td>
<td>41</td>
<td>61</td>
<td>45.7</td>
<td>348</td>
<td>185</td>
<td>301</td>
</tr>
<tr>
<td>7</td>
<td>19</td>
<td>57</td>
<td>47.8</td>
<td>197</td>
<td>89</td>
<td>212</td>
</tr>
<tr>
<td>8</td>
<td>25</td>
<td>60</td>
<td>46.2</td>
<td>463</td>
<td>270</td>
<td>359</td>
</tr>
<tr>
<td>9</td>
<td>22</td>
<td>65</td>
<td>47.8</td>
<td>492</td>
<td>244</td>
<td>372</td>
</tr>
<tr>
<td>10</td>
<td>25</td>
<td>71</td>
<td>43.9</td>
<td>1230</td>
<td>433</td>
<td>654</td>
</tr>
</tbody>
</table>

* Long-term Se intake was calculated as: log(Se intake) = 1.624 log(plasma Se) + 3.389, in which units are μg/d and μg/ml for intake and plasma concentrations respectively (Yang et al. 1989).
procedures. A similar batch was prepared for the second infusion with a final Se concentration of 56.5 μg/ml ampoule.

**Analytical procedures**

The volume of each 24 h urine collection was determined and measured portions were processed for the measurement of total urine Se using HNO₃–HClO₄ digestion (Sun et al. 1987). In addition, measured portions of urine were processed for the isolation of TMSe using the previously published KOH-thermolysis procedure (Sun et al. 1987). Each resulting solution was then analysed for Se content by the fluorometric method (Yang et al. 1987) and the 74 Se : 77 Se isotope ratio (R 74/77 ) with hydride generation-inductively coupled plasma mass spectrometry (Ting et al. 1989).

**Calculations**

Experimental data for each sample for Se and R 74/77 were combined to calculate that portion of Se in the sample originating from the enriched infusate, according to equations 1, 2 and 3:

\[
R_{74/77} = \frac{(74 \text{ Se}^\circ + 74 \text{ Se}^\ast)}{(77 \text{ Se}^\circ + 74 \text{ Se}^\ast)} = \frac{(0.00814 \text{ Se}^\circ + 0.544 \text{ Se}^\ast)}{(0.0738 \text{ Se}^\circ + 0.0441 \text{ Se}^\ast)},
\]

and

\[
\text{Se}^\ast = \text{Se} - \text{Se}^\circ,
\]

where R 74/77 is the measured isotope ratio in the sample (on a weight for weight basis), Se is the Se content of sample, measured fluorometrically, Se^\circ is the Se content of sample of unenriched origin, Se^\ast is the Se content of sample of enriched origin, and the constants are the appropriate weight fractions of each isotope in either unenriched Se, or infusate Se.

Simultaneous solution of equations 1 and 2 yields the desired quantity:

\[
\text{Se}^\ast = \frac{(0.0738 \times R_{74/77} - 0.00815 \times \text{Se})}{(0.02974 \times R_{74/77} + 0.53585)}.
\]

**Results**

Total Se and TMSe levels were determined in urine from all ten men (Figs. 2, 3 and 4) but stable isotopes of Se as total Se^\circ and TMSe were determined in urine from only five of these men (Figs. 5 and 6). Since there were no statistical differences between the corresponding available data between the 10-subject universe and the subset of five subjects, the data shown in Figs. 5 and 6 are representative of all the subjects.

**Relationship between long-term selenium intake and urine excretion**

Long-term Se intake was estimated from the expression

\[
\log(\text{Se intake (μg/d)}) = 1.624\log(\text{plasma Se (μg/ml)}) + 3.389 \quad (\text{Yang et al. 1989}).
\]

The ten subjects of this investigation showed a wide range of long-term Se intake (Table 1), in the range 2–12 times the average long-term intake for North American adults (Levander, 1987). The relationship between the estimated Se intake and urine Se (or TMSe) excretion is shown in Fig. 2. In Fig. 2(A), we have plotted daily urine excretion as average value (±1 SEM) for days 2–7 for each subject against his estimated long-term daily intake of Se, as well as the average value for days 22, 43, and 62. As seen from the data of Fig. 3(A), in which the average daily urine excretion of all subjects as a function of time is shown, there was little systematic decrease in daily urine excretion during the first 7 d, but marked decreases thereafter. Thus, we consider average daily urine excretion for days 2–7 to be a better estimate of the initial endogenous urine excretion than baseline values (day 0). Similar data for urine TMSe are shown in Fig. 2(B) as a function of the estimated long-term intake, and in Fig. 3(B) as a function of time. The average of the ratio in each subject for urine excretion of Se for days 22, 43, and 62 to that for the corresponding value of day 2–7 (Fig. 2(A)) for all subjects.

**Fig. 2.** Relationships between urine selenium (A), or urine trimethyl-selenonium (TMSe) (B), and the estimated long-term Se intake of the male subjects (n 10). (A), Mean urine variable days 2 to 7 (baseline); ( ), mean urine variable days 22, 43 and 62 following intravenous infusions of 105 μg selenium as selenite on day 0. Values are means with standard errors of the mean represented by vertical bars.
was 0.462 (SEM 0.023) and that for TMSe (Fig. 2(B)) was 0.437 (SEM 0.073). Therefore, during days 22–62 each subject, regardless of his initial body status, lost on average 46.2% Se in urine compared with that for days 2–7. Similarly, loss of TMSe decreased to 43.7% of its initial value.

**Relationship between urine selenium and urine trimethylselenonium**

In Fig. 4(A) we have plotted the observed relationship between urine Se and urine TMSe for all urine samples regardless of from whom and when they were collected. In Fig. 4(B), we have presented the data according to the day of collection, and in Fig. 4(C) according to an incremental increase in urine Se content for each 50 μg increment. The relatively large inter-individual variations in urine Se (and TMSe), due to large variations in long-term Se intake of the subjects (Table 1), are apparent in Fig. 4(B) and partially mask the non-linear relationship between the two variables as long-term Se intake increases. However, when the data are presented according to incremental increases in urine Se (Fig. 4(C)), the non-linear relationship between urine TMSe and urine Se becomes clearer. If the polynomial expression given in Fig. 4(C) is accepted to represent the experimental data, then the slope of the relationship at 100 μg Se excretion per day in urine is 0.030 (μg TMSe/μg Se), while the corresponding slope at 300 μg Se excretion per day in urine...
is 14.4 (μg TMSe/μg Se). This indicates a large increase in TMSe excretion with increase in urine Se as long-term intake is increased.

**Relationship between labelled urine selenium and urine trimethylselenonium**

The data showing the relationship between urine excretion of labelled Se (urine Se*) and its TMSe component (TMSe*) are given in Fig. 5. The data are presented in the same general format as in Fig. 4 for chemically measured Se. Data given in Fig. 5(C) show that similarly to the corresponding data for chemically measured Se (Fig. 4(C)) the relationship between urine TMSe* and urine Se* also appears to be non-linear, with the slope increasing as long-term Se intake is increased.

**Relationship of specific activities for urine selenium and trimethylselenonium**

The specific activity (SA) for each sample (urine Se or urine TMSe) was calculated as $R_{74/77} = R_{74/77}$, which corresponds to the $\text{Se}^{74}/\text{Se}^{77}$ ratio. The value of $R_{74/77}$ was obtained by averaging all available data from baseline samples. The observed relationship between specific activities of urine Se and urine TMSe are shown in Fig. 6. In Fig. 6(A), we have plotted average ($\pm$ 1 SEM) SA for daily urine for all subjects.
forms of Se in the body and Se \(^2\) currently accepted relationship between various chemical
1991; Vadhanavikit
consists of two metabolically distinct pools: Semet, which
generates Se \(^2\) (Janghorbani 1988; Sunde, 1990). While the selenite-exchangeable meta-
bolism of Se in human subjects (Sunde, 1990) indicates that the most likely candidate(s) result from generation of
the labile selenocompound, Se\(^{2-}\) (Sunde, 1990; Ip et al. 1991; Vadhanavik 1993). As noted earlier, the
currently accepted relationship between various chemical forms of Se in the body and Se \(^2\) is shown in the simplified scheme of Fig. 1. According to this scheme, body Se consists of two metabolically distinct pools: Semet, which
can be substituted non-specifically for methionine in body proteins, and a selenite-exchangeable metabolic pool
(Janghorbani et al. 1990). Dietary or supplemental Se is added to both pools depending on its chemical forms and
methionine status of the subject (Waschulewski & Sunde, 1988; Sunde, 1990). While the selenite-exchangeable meta-
bolic pool interacts reversibly with Se\(^{2-}\), the Semet pool only generates Se\(^{2-}\). At steady state, there is an average whole body half-life of 102 d for the selenite-exchangeable meta-
bolic pool and 252 d for Semet (Patterson & Levander, 1997). The turnover characteristics of the two pools are
primarily determined by the chemical form consumed (Waschulewski & Sunde, 1988). The steady-state Se content of the selenite-exchangeable metabolic pool is probably at its saturation level in subjects who consume a nutritionally adequate amount of Se (Thomson & Robinson, 1980) as is the case for healthy North Americans (Combs & Combs, 1986; National Research Council, 1989). In contrast, the steady-state Se content of Semet is directly proportional to daily consumption of Semet.
Se\(^{2-}\) is either utilized for synthesis of constituents of the selenite-exchangeable metabolic pool such as the glu-
thurione peroxidases (Sunde, 1990, 1997), selenoprotein P (Burk & Hill, 1993) selenoprotein W (Vendeland et al. 1995), and iodothyronine deiodinases (Sunde, 1997), or is excreted, and does not accumulate in the body. Two known excreatory products are dimethyl selenide and TMS\(\text{e}^\text{\text{e}}\) (Ip et al. 1991; Vadhanavik et al. 1993) which are expired in air (Hsieh & Ganther, 1977) or excreted in urine (Sun et al. 1987) respectively.

As seen from the data of Fig. 2, the relationship between urine Se and the estimated long-term Se intake is more linear than that for TMS\(\text{e}^\text{\text{e}}\) over the entire range of intake. This may indicate that other excretory pathways in addition to TMS\(\text{e}^\text{\text{e}}\) play increasingly significant roles in eliminating excess catabolized-tissue Se. In light of the quantitative significance of respiratory dimethyl selenide under sub-
toxic-to-toxic intakes (Hsieh & Ganther, 1977), this path-
way may account for some of the non-linearity observed in urine Se (or TMS\(\text{e}^\text{\text{e}}\)) at the high end of long-term Se intake in our subjects. This non-linearity appears to hold not only for data obtained during the initial 7 d following dietary Se restriction (Fig. 2), but also for data obtained for days 22, 43, and 62 (Fig. 2), indicating possibly that even after approximately an initial month of dietary Se restriction, non-excretion may still be an important loss pathway in our subjects. Furthermore, the value of the ratio of daily urine Se excretion for the latter days (average value for days 22, 43, and 62; Fig. 2) to the corresponding value for the initial days (average value for days 2–7; Fig. 2) appeared to be independent of long-term Se intake in our subjects (average for all subjects: 0.462 (SEM 0.023)). The same was also true for TMS\(\text{e}^\text{\text{e}}\) excretion (average ratio for all subjects: 0.437 (SEM 0.073)). One interpretation for this observation is that whole-body turnover of Se in these subjects was not affected by the 1–2 month dietary Se restriction of our experiment.

Urinary Se and TMS\(\text{e}^\text{\text{e}}\) excretion did not differ during the first 7 d after the life-long high exposure to Se (Fig. 3(A) and (B) days 2–7). However, after the dose of labelled-Se at day 64, both Se and TMS\(\text{e}^\text{\text{e}}\) excretion declined during the following 7 d (days 64 to 71). This suggests that enough body Se had been depleted to affect the excretion of urinary Se and TMS\(\text{e}^\text{\text{e}}\).

Data given in Figs. 4 and 5 indicate the possible non-
linear relationship between urine TMS\(\text{e}^\text{\text{e}}\) and urine Se, both for TMS\(\text{e}^\text{\text{e}}\) arising from catabolism of endogenous body Se (Fig. 4) and that from infused selenite (Fig. 5). Data shown in Fig. 4(C), and to a lesser extent also Fig. 5(C), show that the major determinant of this non-linear relationship is perhaps the flux of Se\(^{2-}\) generated from catabolism of body selenocompounds, and other subject-specific factors may play a more minor role. However, even in subjects with very high long-term Se intake urine TMS\(\text{e}^\text{\text{e}}\) still constitutes a quantitatively small fraction of urine Se. Therefore, the combination of the two observations that: (1) rate of urine excretion of Se decrease per unit increase in long-term Se intake, and (2) TMS\(\text{e}^\text{\text{e}}\) and urine Se are non-linearly related, indicate that either urine Se or urine TMS\(\text{e}^\text{\text{e}}\) alone may not provide a linear representation of whole body flux of Se\(^{2-}\). It may be that the combination of TMS\(\text{e}^\text{\text{e}}\) excretion in urine and dimethyl selenide in expired air provides the most quantita-
tively accurate measure of whole body Se\(^{2-}\) flux and thus the most accurate measure of the ability of whole body Se to maintain tissue fluxes of biologically active Se for chemopreventive purposes. The validity of this hypothesis needs further exploration.

Finally, the nearly identical SA observed for urine Se and
its TMSe component (Fig. 6) indicate the very close precursor–product relationship for these two components. This may indicate that most urine Se is related to methylation of tissue Se\(^{2-}\). Equally importantly, this may also indicate identical metabolism of infused selenite and catabolized endogenous Se. If this is the case, urine excretion of infused labelled-selenite may also provide as satisfactory a means for monitoring whole body flux of endogenous Se\(^{2-}\) as endogenously produced TMSe, or urine Se.

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