Dietary exposure biomarker-lead discovery based on metabolomics analysis of urine samples

Manfred Beckmann¹, Amanda J. Lloyd¹, Sumanto Haldar², Gaëlle Favé³, Chris J. Seal², Kirsten Brandt², John C. Mathers³ and John Draper¹*

¹Institute of Biological Environmental and Rural Sciences, Aberystwyth University, Aberystwyth SY23 3DA, UK
²Human Nutrition Research Centre, School of Agriculture, Food and Rural Development, Newcastle University, Newcastle upon Tyne NE1 7RU, UK
³Human Nutrition Research Centre, Institute for Ageing and Health, Campus for Ageing and Vitality, Newcastle University, Newcastle upon Tyne NE4 5PL, UK

Although robust associations between dietary intake and population health are evident from conventional observational epidemiology, the outcomes of large-scale intervention studies testing the causality of those links have often proved inconclusive or have failed to demonstrate causality. This apparent conflict may be due to the well-recognised difficulty in measuring habitual food intake which may lead to confounding in observational epidemiology. Urine biomarkers indicative of exposure to specific foods offer information supplementary to the reliance on dietary intake self-assessment tools, such as FFQ, which are subject to individual bias. Biomarker discovery strategies using non-targeted metabolomics have been used recently to analyse urine from either short-term food intervention studies or from cohort studies in which participants consumed a freely-chosen diet. In the latter, the analysis of diet diary or FFQ information allowed classification of individuals in terms of the frequency of consumption of specific diet constituents. We review these approaches for biomarker discovery and illustrate both with particular reference to two studies carried out by the authors using approaches combining metabolite fingerprinting by MS with supervised multivariate data analysis. In both approaches, urine signals responsible for distinguishing between specific foods were identified and could be related to the chemical composition of the original foods. When using dietary data, both food distinctiveness and consumption frequency influenced whether differential dietary exposure could be discriminated adequately. We conclude that metabolomics methods for fingerprinting or profiling of overnight void urine, in particular, provide a robust strategy for dietary exposure biomarker-lead discovery.

Dietary exposure: Metabolite fingerprinting: FFQ: Multivariate data analysis: Urine biomarkers

The accurate measurement of dietary exposure, which is an essential component of much health-related research, offers a challenging prospect. Specifically, dietary data can be subject to participant bias and can depend heavily upon food composition tables for the estimation of intakes of energy, nutrients and other food constituents. The chemical content of body fluids is a potentially rich source of information about dietary exposure as many foods contain distinctive metabolites which give rise to further chemical diversity following food ingestion, absorption and metabolism. However, to date, putative biochemical markers are available for only a relatively small number of specific foods and food components. The comprehensive analysis of metabolites in biological fluids using
Table 1. Recent publications in which non-targeted metabolite fingerprinting or metabolite profiling has been used to discover potential dietary biomarkers in food intervention and cohort studies

<table>
<thead>
<tr>
<th>Dietary exposure</th>
<th>Metabolomic technique</th>
<th>Potential biomarker-leads</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Acute food exposure studies (up to 24 h)</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Chicken and orange juice</td>
<td>FIE-MS</td>
<td>Chicken: anserine; orange juice: proline betaine and 4-hydroxy-proline betaine</td>
<td>20</td>
</tr>
<tr>
<td>Apple juice</td>
<td>HPLC–ESI–MS/MS</td>
<td>Phloretin O-glucuronides and eight (methyl) quercetin O-glucuronides, 5-caffeoylquinic acid, 4-p-coumaroarylpyruvic acid, caffeic acid, epicatechin, phloretin and quercetin</td>
<td>22</td>
</tr>
<tr>
<td>Cocoa powder with milk or water</td>
<td>HPLC-Q-ToF</td>
<td>Caffeine and theobromine (epicatechin, methyl-epicatechin, vanillic acid, vanilloxyglucine, phenylvaleric acid, phenylvalerolactone derivatives, 3,5-diethyl-2-methylpyrazine and hydroxycacetophenone) and processing-derived products (diketopiperazines), as well as trigonelline, hydroxykynotonic acid, nicotinic acid and tyrosine</td>
<td>23</td>
</tr>
<tr>
<td>Cocoa powder and milk</td>
<td>HPLC-Q-ToF</td>
<td>Methylguanine, vanillloylglycine, dihydroxyphenylvalerolactone glucuronide, furuoylglycine, 3- and 7-methylxanthine, theobromine and xanthurenic acid</td>
<td>24</td>
</tr>
<tr>
<td>Oily fish (salmon); a cruciferous vegetable (steamed broccoli); berry fruit (raspberries)</td>
<td>FIE-MS</td>
<td>Salmon: trimethylamine-N-oxide, methylhistidine, anserine. Raspberry: caffeoyl sulphate, methyl epicatechin sulphate, ascorbate, 3 hydroxyhipperic acid, naringenin glucuronide. Broccoli: ascorbate and breakdown products (xylonate/lyxonate, threitol/erythritol) naringenin and hesperitin glucuronide</td>
<td>25</td>
</tr>
<tr>
<td>Coffee</td>
<td>HPLC-PDA-MS</td>
<td>Caffeic acid sulphate, caffeoyquilinic acid lactone sulphate, caffeoyquilinic acid sulphate, dihydro(pseudo)ferulic acid glucuronide, dihydrocaffeic acid and glucuronide/sulphate, dihydroferulic acid and glucuronide/sulphate, feruloylglycine, feruloylquinic acid, iso/ferulic acid glucuronide/ sulphate</td>
<td>32</td>
</tr>
<tr>
<td><strong>Short-term food interventions (up to 2 weeks)</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Increased milk diet or an increased meat diet</td>
<td>1H NMR</td>
<td>Milk diet: increased hippurate; Meat diet: increased creatine, histidine and urea</td>
<td>17</td>
</tr>
<tr>
<td>High cruciferous vegetable (broccoli and Brussels sprouts)</td>
<td>1H NMR</td>
<td>S-methyl-L-cysteine sulphone (SMCSO) and three structurally related compounds</td>
<td>19</td>
</tr>
<tr>
<td>Mixed-fruit meal (apple, orange, grapes and grapefruit)</td>
<td>1H NMR</td>
<td>Citrus fruits: hippuric acid, proline betaine and tartaric acid</td>
<td>21</td>
</tr>
<tr>
<td>Dark chocolate</td>
<td>1H NMR and LC-MS</td>
<td>Increased levels of 4-hydroxyphenylacetate and several unassigned metabolites</td>
<td>28</td>
</tr>
<tr>
<td>‘Vegetarian’, ‘low meat’ and ‘high meat’ diets</td>
<td>1H NMR</td>
<td>High-meat diet: creatine, carnitine, acetylcarnitine and trimethylamine-N-oxide. Low-meat diet and vegetarian diet signatures: p-hydroxyphenylacetate</td>
<td>33</td>
</tr>
<tr>
<td>Low-phytochemical diet followed by a standard phytochemical diet</td>
<td>1H NMR</td>
<td>Hipurate increased in high phytochemical diet</td>
<td>35</td>
</tr>
<tr>
<td><strong>Long-term food interventions (&gt; 1 month)</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wheat bread v. rye bread</td>
<td>HPLC-Q-ToF</td>
<td>3-(3,5-dihydroxyphenyl)-1-propanoic acid sulphate, enterolactone glucuronide, azelaic acid, 2-aminophenol sulphate and 2,4-dihydroxy-1,4-benzoxazin-3-one</td>
<td>18</td>
</tr>
<tr>
<td>High protein (HP) or low protein (LP) diet</td>
<td>1H NMR</td>
<td>Increased nitrogen, creatinine, TMAO, creatine after the HP diet</td>
<td>31</td>
</tr>
<tr>
<td>High mixed nuts consumption</td>
<td>HPLC-Q-TOF-MS</td>
<td>10-hydroxy-decene-4,6-dynoic acid sulphate, tridecaenoic/tridecyanoic acid glucuronide, dodecanedioic acid, pyrogallol sulphate, p-coumaryl alcohol glucuronide, urolithin A glucuronide/sulphate/phosphoglucuronide, N-acetylsersotonin sulphate and hydroxyindoleacetic acid</td>
<td>34</td>
</tr>
</tbody>
</table>
metabolomics technology provides an objective approach for the discovery of dietary exposure biomarkers. Non-targeted metabolite fingerprinting, using either NMR or MS, and metabolite profiling using liquid chromatography MS have been used successfully for biomarker discovery using urine samples from various study designs (Table 1). In acute food intervention studies, participants were exposed to specific foods in known amounts and either postprandial urines sampled before the next meal or overnight or 24 h urines were collected. In other studies, participants were established on a specific diet for several days/weeks or longer term (1 month) before urine sampling. More recently there have been reports of the use of cohort studies in which participants consumed a freely-chosen diet. In these studies, the analysis of diet diary or FFQ information allowed classification of individuals in terms of their frequency of consumption of specific diet constituents. In the present paper, we illustrate these approaches for biomarker discovery with particular reference to two studies carried out by the authors.

The presence of substantial inter- and intra-individual variability in human metabolite profiles provides a challenge for both biofluid sampling and subsequent data normalisation in metabolomics studies seeking information on habitual diet. To address this problem, standardised methods have been validated recently both for the management of participants and for urine sampling in large-scale food interventions involving free-living individuals and also for acute postprandial studies in a controlled environment. Key features of these study protocols include behavioural restrictions, e.g. no alcohol and the consumption of a standardised evening meal in the evening before a clinic visit to provide a fasting urine sample. It was anticipated that the latter would provide a ‘normalised’ background against which differences in urine chemistry resulting from either previous habitual dietary intake prior to clinic visit or acute food intake during the test day would be detectable.

With effective protocols in place for volunteer management and urine sampling there was now an opportunity to determine whether changes in urine chemistry could reflect dietary exposure. In an acute feeding ‘proof of principle’ study, urine samples were analysed from individuals participating in the MEtabolomics to characterise Dietary Exposure (MEDE) research programme. As part of the MEDE project, twenty-four healthy participants consumed a ‘test’ breakfast, in which the cereal component of a standardised breakfast was replaced by one of four foods of high public health importance, followed by the collection of postprandial urine samples for metabolome analysis. Once candidate food biomarkers had been identified, there was then the opportunity to validate their potential usefulness to monitor habitual diet in the independent GrainMark study (http://www.ncl.ac.uk/afrd/research/project/2287). This large-scale dietary intervention study, involving free-living individuals, aimed to discover potential biomarkers of dietary wholegrain exposure. After a washout period of 4 weeks the participants (sixty-eight in total) were asked to consume three servings of...
Proceedings of the Nutrition Society

either wholegrain rye foods or wholegrain wheat foods per d for 4 weeks and subsequently doubled their intake of the same foods for a further 4 weeks. At baseline, and at the middle of each 4-week intervention period (washout, three servings and six servings of wholegrain rye/wheat foods per d), volunteers completed a validated FFQ (four in total) based on the European Prospective Investigation into Cancer and Nutrition FFQ (7), which recorded consumption of foods for a 7 d period, within 7–14 d of sampling (37).

Using data from both the MEDE and GrainMark studies it has been shown that analysis of overnight void urine samples can provide a rich source of potential biomarkers of habitual diet as reported in FFQ(26,27).

Postprandial urine composition reflects recent dietary exposure

Table 1 lists several recent studies in which non-targeted metabolite fingerprinting or metabolite profiling has been used to discover dietary biomarkers in both acute and short-term food intervention studies(17,19–25,28,32,33,35). The basic design principles are illustrated in Fig. 1 with reference to the MEDE study in which fasted participants consumed specific foods (Fig. 1(a)) as part of a standardised breakfast. Metabolome fingerprints representing postprandial urine(25) were then generated using non-targeted, nominal mass flow injection electrospray–ionisation MS(15,20,25). Figure 1(b) illustrates a typical flow injection electrospray-ionisation MS urine fingerprint of a mass range from 

\[ m/z \]

100 to 800, which shows that urine fingerprints are both complex and information-rich. The question of whether a postprandial urine sample contains chemicals distinctive of exposure to specific foods can be evaluated by subjecting the urine fingerprint data to powerful supervised multivariate analysis including principal component-linear discriminant analysis. Figure 1(c) shows a scores plot from a typical principal component-linear discriminant analysis (16) of flow injection (d) Annotation of ions by targeted ultra-high accurate mass analysis by FT-ICR-MS and MS/MS

(e) Potential biomarkers from MEDE study

Citrus fruits:
- Proline
- Betaine

Oily fish:
- Anserine

Fig. 1. (colour online) Typical analytical strategy for food biomarker discovery based on an acute food intervention. All elements of this example workflow are derived from data from experiments previously reported(25). (a) The standardised breakfast consisted of orange juice, a cup of tea with milk and sugar, a butter croissant and cornflakes with milk, while in the test breakfasts the cornflakes and milk were replaced with smoked salmon, steamed broccoli or raspberries. Postprandial urine was collected. (b) Typical flow infusion electrospray-ionisation MS (FIE-MS) fingerprints of postprandial urine. (c) Scores plot from a typical principal component-linear discriminant analysis (PC-LDA) of the metabolite fingerprinting data (m/z 100–800) derived from analysis postprandial urine. (d) Fourier transform-ion cyclotron resonance ultra-mass-spectrometry (FT-ICR-MS) plot of the nominal mass 'bin' m/z 241. (e) Potential biomarkers from the Metabolomics to characterise Dietary Exposure (MEDE) study. PCA, principal component analysis; RF, random forest; MS/MS, flow infusion electrospray-ionisation tandem MS.
electrospray–ionisation MS fingerprints representing urine samples from volunteers exposed to a standard breakfast or to a breakfast in which the cereal component of the standard breakfast was replaced by smoked salmon, broccoli or raspberries\(^{(25)}\). With an eigenvalue (\(T_w\)) of 2.55 in the dimension of maximal discrimination (Fig. 1(c)), it is evident that the ‘test’ foods (particularly smoked salmon) are adequately discriminated from the standard breakfast. A range of feature selection methods\(^{(16)}\) can then be employed to determine the flow injection electrospray–ionisation MS signals responsible for the discrimination of each food and, under most circumstances, it has been found that agglomerative decision trees such as random forest perform consistently well\(^{(20, 25, 26)}\). The detailed analysis of highlighted nominal masses can be undertaken by targeting them for further investigation on MS instruments capable of ultra-high mass resolution\(^{(25–26)}\). The Fourier transform-ion cyclotron resonance ultra–MS plot shown in Fig. 1(d) represents the analysis of the nominal mass m/z 241, a signal linked with the consumption of oily fish, indicating that a signal with a mass of 241.12958 is a likely biomarker candidate for oily fish exposure\(^{(25)}\). The accurate mass information can be used to generate an elemental formula and to predict candidate metabolites that could yield the measured ion using annotation tools such as MZedDB\(^{(39)}\), which take into account anticipated ionisation behaviour. Further experiments in which the selected ion is fragmented and the resulting spectrum compared with that of a chemical standard are required to assign a putative structure. For example, in the MEDE study these subsequent targeted analyses suggested that anserine (nominal mass m/z 241) is a candidate biomarker for oily fish consumption and proline betaine for citrus foods (Fig. 1(e))\(^{(25, 26)}\).

Exposure to citrus foods has provided a paradigm for discovery of dietary biomarkers by analysis of dietary data

Citrus fruits and citrus fruit juices are a distinctive and frequently consumed (i.e. often once per d) component of the UK diet and thus represent a paradigm for validation of biomarkers of habitual dietary exposure. Studies using either NMR\(^{(21)}\) or ESI–MS fingerprinting\(^{(20)}\) of post-prandial urines demonstrated initially that proline betaine was a potential biomarker of acute exposure to citrus foods. There was thus an opportunity to determine whether the urinary concentration of this metabolite reflected habitual exposure to the same foods. Recent publications described a multivariate modelling strategy for habitual dietary biomarker discovery dependent on the comparison of urine metabolome fingerprints from groups of individuals reporting differential exposure to specific dietary components\(^{(26, 27, 29, 30)}\). An initial analysis\(^{(26)}\) of habitual citrus exposure data in the MEDE study (n = 24) indicated that there were sufficient individuals reporting citrus consumption to allow the assignment of volunteers into three broad exposure levels (High (two–three citrus portions per d), Medium (about one citrus portion per d) and Low (less than two citrus portions per week)). With the larger number of participants in the GrainMark study it was possible to assign individuals to a more quantitative scale of habitual dietary exposure (i.e. never, 1 per week, 2–4 per week, 5–6 per week, 1 per d, 2–3 per d)\(^{(27)}\). Overnight void urine samples available from both studies were subjected to metabolite fingerprinting and the m/z signals responsible for discriminating higher v. lower habitual consumption levels were identified using random forest feature selection\(^{(16)}\). In both studies the majority of the top twenty highest ranked signals responsible for the classification of High v. Low habitual citrus consumption were identified as ionisation products of proline betaine\(^{(26, 27)}\). Although not strictly quantitative, the relative intensity of one of the major ions (m/z 144; [M+H\(^+\)]) reflected the level of habitual citrus exposure reported by individual participants in both MEDE and GrainMark studies. A similar result has been reported for NMR signals associated with recent exposure to citrus foods that were found to be derived from the presence of proline betaine in 24 h urine samples associated with recent exposure to citrus foods\(^{(21)}\).

**Habitual consumption frequency for individual foods impacts on the ability to detect differential dietary exposure using overnight urine samples**

Following the success with citrus foods there have been several recent reports in which dietary data have been analysed from studies where, in most cases, participants have eaten freely-chosen diets (Table 1)\(^{(26, 27, 29, 30)}\). These meta-data allowed the assignment of individuals into different food consumption frequency classes and could be used to study habitual exposure for a range of other foods. In the GrainMark study, as expected, habitual consumption frequency differed greatly between individual foods, and these food-specific consumption patterns were generally consistent with the MEDE study\(^{(27)}\). Overall, the patterns of intake of individual foods could be summarised in four general exposure categories (Fig. 2) ranging from foods consumed very infrequently (e.g. liver or kidneys) to those consumed, on average, more than once per d (e.g. coffee). Although habitual citrus exposure could be modelled adequately using data from the MEDE study\(^{(26)}\), with only twenty-four participants the present strategy was not suitable for biomarker discovery for foods which are consumed very infrequently by most people. Assignment to habitual consumption frequency ranges for the remaining foods (grouped into High skewed, Normal distribution or Low skewed consumption patterns) identified sufficient numbers of individuals within the GrainMark study to develop higher v. lower frequency exposure groups for multivariate classification\(^{(27)}\). The likelihood of discovering potential biomarkers for each food was assessed by determining the ‘goodness’ of class discrimination using random forest margin values and area under the receiver operating characteristic curve values as robust classification statistics\(^{(16)}\). Figure 2 illustrates a clear trend that classification efficiency (as assessed by random forest margins and area under the receiver operating characteristic curve values) is generally higher in foods that are consumed...
more frequently. We suggest that such foods will form likely candidates for monitoring using a biomarker strategy if it can be proven that the presence of potential biomarker signals in urine can be linked to the ingestion and chemical composition of specific dietary components. An example ‘data-driven’ strategy for the discovery of potential dietary biomarkers based on the analysis of GrainMark FFQ structure and data is summarised in Fig. 3.

Structural analysis of potential biomarkers representing three selected foods shows that their presence in overnight void urine reflects original food chemical composition

Metabolome models (based on overnight void urine samples) of three distinctive foods representing examples of Low skewed, Normal distribution and High skewed habitual dietary exposure groups have been investigated in more detail to determine whether the selected metabolites could be correlated with known food chemistry\(^\text{[10,26,32,33,40–51]}\). Groups of participants were identified for each dietary component to represent High, Medium and Low exposure classes within the consumption ranges described for each food in the GrainMark study. Thus for oily fish (Low skewed) the Low consumption category represents less than one portion per week, Medium is approximately one per week and High is greater than or equal to two to four portions week. For tomato (Normal distribution), the Low consumers ate less than or equal to one portion per week, Medium consumers ate greater than two to four portions per week, while High consumers ate up to five to six portions per week. High consumers of coffee (High skewed) drank more than one cup per d while Medium consumers had greater than or equal to two to four cups per week and Low consumers more than or equal to one per week. Despite the wide range of consumption frequencies each of these distinctive foods generated adequate classification models (Fig. 4(a)).

In the GrainMark study, dihydrocaffeic acid-3-O-glucuronide was highly ranked as a potential biomarker of habitual coffee exposure (Fig. 4(b)). Previous studies on acute exposure to coffee reported the presence in urine and/or plasma of at least ten phenolic compounds which represented metabolic endpoints of the biotransformation of chlorogenic acid and caffeic acid which is present at high levels in this beverage\(^\text{[10,32,43,49]}\). Although chlorogenic acid and caffeic acid are found in many fruits and vegetables and it is unlikely that dihydrocaffeic acids will prove to be unique biomarkers of coffee consumption, it is interesting that other phenolic metabolites identified in acute exposure studies were not highlighted by this analysis. Investigation of top ranked signals for oily fish exposure showed that methyl-histidine (probably derived from anserine, which was also highly ranked, by the action of carnosinase\(^\text{[40–42]}\)) was an excellent biomarker candidate (Fig. 4(c)). Thus, the identity of both of these potential biomarkers could be linked directly to the previous chemical analysis of each food. Although hippuric acids (presumably derived from colonic fermentation of the hydroxycinnamic acid content of tomato fruits\(^\text{[48]}\)) were indicative of tomato exposure, many highly ranked signals
proved to be dihydroxyphenylvalerolactone conjugates that commonly result from the colonic fermentation of flavonoids, such as flavanols (Fig. 4(d)). As tomato contains insignificant amounts of such polyphenols it is most likely that these compounds are associated with foods strongly co-consumed in meals containing tomatoes. This observation highlights the requirement for careful validation of any potential food biomarkers in the context of the whole diet and emphasises the utility of the test meal approach (as used in the MEDE study) for the identification of putative biomarkers which are causally linked with food exposure. In addition, such data-driven strategies for putative biomarker discovery are constrained by the robustness of the original dietary intake data which may contain inherent biases. Further, both conceptual and practical challenges are likely in cases where there are strong correlations between intakes of foods with related chemistries.

Conclusion

The advent of non-targeted metabolomics technology for global chemical fingerprinting/profiling of human biofluids has offered an opportunity to accelerate research on food biomarker discovery. Recent data support the concept that metabolomics analysis of 24 h or overnight void urine samples, in particular, will provide a productive strategy for the identification of candidate dietary exposure biomarkers. The demonstration that biomarker discovery using high throughput metabolomics is feasible using urine samples derived from populations of free-living individuals (26,27,29,30) supports the ultimate objective of using such biomarkers in epidemiological studies. Evidence is now accumulating that a range of relatively frequently consumed (i.e. one to two portions per week) and more distinctive foods can be considered good candidates for biomarker discovery using biofluid samples from cohort studies (26,27,29,30). Many other foods forming major components of composite meals, or less frequently consumed food items, may be targets for biomarker discovery but would probably require well designed, controlled food intervention studies to identify candidate metabolites. In future studies we propose testing this hypothesis by the analysis of overnight urine samples collected at home by individuals consuming carefully constructed weekly menus designed both to provide adequate exposure to specific dietary components and to offer foods in specific combinations in order to expose any biomarker redundancy that could confound dietary exposure measurement.

Ensuring the generation of accurate dietary information in epidemiological studies requires considerable effort from both researchers and study participants. Despite the
Fig. 4. (colour online) Habitual consumption of distinctive foods eaten in a range of consumption frequencies is detectable in overnight urine samples by flow infusion electrospray–ionisation MS (FIE-MS) fingerprinting. The data illustrated in this figure are derived from experiments previously reported (27). (a) Principal component-linear discriminant analysis (PC-LDA) of the metabolite fingerprinting data (m/z 100–550; oily fish, positive ionisation mode; tomato and coffee, negative mode) derived from the analysis of overnight 'PRE' urine from participants classified as representative of low/medium/high exposure ranges (ten to twelve per exposure group) based on the average FFQ score for exposure to these different foods. Oily fish: low, less than one per week; medium, one per week; high, greater than or equal to two to four per week; tomato: low, less than or equal to one per week; medium, greater than two to four per week to less than five to six per week; high, greater than or equal to five to six per week; coffee: low, less than or equal to one per week; medium, greater than two to four per week to less than five to six per week; high, greater than one per d; eigenvalues (Tw values) are given in parentheses. Markers confirmed using Fourier transform-ion cyclotron resonance ultra-mass-spectrometry (FT-ICR-MS), flow infusion electrospray-ionisation tandem MS and MZedDB. (b) Signals highly ranked in habitual coffee exposure models. (c) Signals highly ranked in habitual oily fish exposure models. (d) Signals highly ranked in habitual tomato exposure models.
advent of increasingly sophisticated digital tools\(^1\), diet recording remains an inherent source of major uncertainty, even in studies where individuals recording food exposures are well trained and carefully monitored. Following validation in epidemiological and/or controlled dietary intervention studies, it is expected that putative food intake biomarkers can be translated into practical measurements that can complement, or in some cases replace, more traditional methods of assessing dietary exposure. We conclude that in the not too distant future, urine biomarker technology may allow objective monitoring of the levels of intake of several key foods and strengthen the evidence for causal links between dietary exposure and health outcomes.

Acknowledgements

None of the authors has a conflict of interest with respect to this manuscript. This research programme was supported by the UK Food Standards Agency Projects N05073 and N05075 and MRC Programme Grant MR/J010308/1. The authors’ contributions to the work were as follows: M. B. developed urine extraction procedures, designed metabolite fingerprinting experiments, supervised MS support staff, pre-processed data for analysis and edited the manuscript; A. J. L. was involved in data analysis, produced figures, researched the literature and wrote the manuscript; G. F. and S. H. undertook volunteer recruitment, coordinated the project, supervised research in Newcastle University, designed volunteer handling protocols and edited the manuscript; J. D. coordinated the project, supervised research in Aberystwyth, designed figures and wrote the manuscript.

References

Food biomarkers in the urine metabolome 361