Optimum nutrition: thiamin, biotin and pantothenate

David A. Bender
Department of Biochemistry and Molecular Biology, University College London, Gower Street, London WC1E 6BT, UK

The metabolism of glucose is deranged in thiamin deficiency, but once any deficiency has been corrected there is no further effect of increased thiamin intake on the ability to metabolize glucose through either pyruvate dehydrogenase (EC 1.2.4.1) and the citric acid cycle, or the pentose phosphate pathway, in which transketolase (EC 2.2.1.1) is the thiamin-dependent step. It has been suggested that the Wernicke-Korsakoff syndrome is associated with a genetic variant of transketolase which requires a higher than normal concentration of thiamin diphosphate for activity. This finding would suggest that there may be a group of the population who have a higher than average requirement for thiamin, but the evidence is not convincing. There are no estimates of biotin requirements, but either coenzyme saturation of erythrocyte pyruvate carboxylase, or the excretion of 3-hydroxy-isovalerate (perhaps after a test dose of leucine) could be used to assess requirements in depletion–repletion studies. Biotin deficiency leads to impaired glucose tolerance, but it is unlikely that glucose tolerance could be used to assess optimum biotin status, since other more common factors affect glucose tolerance to a greater extent. Plasma triacylglycerol and non-esterified fatty acids are moderately elevated in pantothenic acid deficiency. However, this is unlikely to be useful in assessing pantothenate status, since again, other more common factors affect plasma lipids. To date there are no biochemical indices of adequate pantothenate nutrition, and no estimates of requirements.

Thiamin status: Biotin status: Pantothenate status: Transketolase activation:

Erythrocyte pyruvate carboxylase: 3-Hydroxy-isovalerate excretion

In the absence of any clear definition of optimum nutrition, and given the impracticability of performing whole-life studies, we have to investigate the metabolic functions of the nutrients in order to determine which, if any, may provide markers of a level of intake that could be considered to be desirable or optimum, rather than being simply adequate to prevent deficiency. It is also relevant to consider whether inadequate intake of the nutrient in question may be sufficiently important to warrant investigation, or whether average intakes are so obviously greater than optimum levels of intake that further investigation is probably irrelevant.

Thiamin

Thiamin has two distinct functions. Thiamin diphosphate (TDP) is the coenzyme for pyruvate dehydrogenase (EC 1.2.4.1) and transketolase (EC 2.2.1.1) in carbohydrate metabolism, or-ketohutyrate dehydrogenase (EC 1.2.4.2) in the citric acid cycle, and branched-chain keto-acid dehydrogenase (EC 1.2.4.4) in the metabolism of the branched-chain amino acids. Thiamin triphosphate (TTP) acts in nerves (and possibly also muscle) to activate a chloride ion channel.

Bettendorf et al. (1993b, 1994) showed that the formation of TTP in brain-membrane-vesicle preparations was correlated with chloride uptake. In neuroblastoma cells in culture the addition of TTP results in activation of the chloride ion channel, which is not reversed by washing out, suggesting that it acts by phosphorylation of the chloride-channel protein (Bettendorf et al. 1993a). Impaired formation of TTP may be a factor in the neurological signs of both beriberi and Wernicke-Korsakoff syndrome, although Thornber et al. (1980) reported that TTP was preserved in the brains of thiamin-deficient lambs, at the expense of TDP and free thiamin. There are no reports of TTP concentration in analyses of brains of patients with Wernicke-Korsakoff syndrome, presumably because of its instability post mortem, and at present there is no way of using TTP or membrane chloride permeability to assess thiamin nutritional status.

The role of TDP in pyruvate dehydrogenase was elucidated in the 1930s. Thiamin deficiency results in
Impaired oxidation of pyruvate, and hence accumulation of lactate and pyruvate, especially after a glucose load and moderate exercise; for many years this process was the basis of assessed thiamin nutritional status (Horwitt & Kreisler, 1949). It is unlikely, however, that additional thiamin, above the amount required to prevent deficiency, will increase aerobic capacity and prevent the development of lactic acidosis after vigorous exercise, or that lactic acidosis after exercise can be used as an index of optimum thiamin status.

In maximum exertion the limiting factor is not the activity of pyruvate dehydrogenase, but the rate at which O₂ can be delivered to muscle. Furthermore, the activity of pyruvate dehydrogenase is regulated by phosphorylation and dephosphorylation in response to the intracellular NADH : NAD⁺, GABA after intracerebroventricular injection of [¹⁴C]glutamate (as assessed by the specific activity of glutamate), and may provide the basis of a test of functional capacity, since the rate of O₂ uptake, plasma lactate, lactate threshold, heart rate during exercise to exhaustion, or the time to complete 2 km on a cycle ergometer.

α-Ketoglutarate dehydrogenase is very similar to pyruvate dehydrogenase, and is similarly impaired in thiamin deficiency (Butterworth et al. 1993). However, there seems to be little or no impairment of citric acid cycle activity, as might be expected if there were reduced activity of this enzyme. Page et al. (1989) suggested that this lack of impairment was because the γ-amino butyric acid (GABA) shunt provides an alternative pathway. α-Ketoglutarate can undergo transamination to glutamate, followed by decarboxylation to GABA, which undergoes transamination to succinate semialdehyde, and oxidation to succinate. They demonstrated that in the brains of thiamin-deficient rats there was significantly greater metabolic flux through the GABA shunt (as assessed by the specific activity of GABA) than in control animals. There is no simple way of assessing the activity of the GABA shunt in vivo, since the same C atom is lost as CO₂ whether α-ketoglutarate undergoes decarboxylation directly or by way of glutamate formation.

Thiamin deficiency also impairs carbohydrate metabolism through the pentose phosphate pathway, as a result of reduced activity of transketolase. This reduction in activity may provide the basis of a test of functional capacity, since metabolic flux through the pentose phosphate pathway, compared with that through glycolysis, can be estimated by the release of [¹³C]CO₂ from [¹-¹³C]- and [⁶-¹³C]glucose. However, although transketolase has a significant control coefficient (0.74) in the non-oxidative part of the pentose phosphate pathway (Berrthou et al. 1992), the entry of glucose-6-phosphate into the pathway is controlled by glucose-6-phosphate dehydrogenase (EC 1.1.1.49), largely in response to NADPH : NADP⁺, and the demands of NADPH for lipogenesis and GSH reductase (EC 1.6.4.2), as well as the need for pentoses for nucleotide synthesis (Stryer, 1995). Thus, although there may be apo-transketolase in tissues, and complete saturation of the enzyme with TDP may provide an index of more than minimally adequate thiamin status, this factor is unlikely to increase metabolic flux through the pentose phosphate pathway.

**The Wernicke-Korsakoff syndrome**

Impaired activity of transketolase has been implicated as a factor in the central nervous system lesions of Wernicke’s encephalopathy, a condition that may be considerably more prevalent than has been believed. Harper (1979) reported that Wernicke’s encephalopathy had only been diagnosed in 13 % of a series of patients in whom brain lesions were detected post mortem. There is considerable international variation in the prevalence of the Wernicke-Korsakoff syndrome, which is not accounted for by variations in alcohol consumption, and a body of evidence to suggest genetic susceptibility (Zubaran et al. 1997). There are two genes for transketolase: the most investigated is on chromosome 3, but Cay et al. (1996) reported a second transketolase gene on the X chromosome. If this X-linked gene were associated with susceptibility to the Wernicke-Korsakoff syndrome, there would be a sex-linked (maternal) pattern of inheritance. However, most of the evidence suggests autosomal inheritance (Mukherjee et al. 1987), so it is likely that it is the gene on chromosome 3 that is important.

Blass & Gibson (1977) reported that the affinity of transketolase for TDP was approximately 10-fold lower in cultured fibroblasts from patients with Wernicke-Korsakoff syndrome than in those from control subjects, suggesting that there may be a genetic polymorphism in transketolase that would result in a subgroup of the population having higher than average thiamin requirements. This finding is akin to the thiamin-responsive variant of maple-syrup-urine disease, where the affinity of branched-chain keto-acid dehydrogenase is considerably lower than normal; in this case thiamin supplements of approximately 100 mg/d are required (Scrivert et al. 1971).

In support of the suggestion of a variant transketolase associated with the Wernicke-Korsakoff syndrome, Jeyasingham et al. (1987) reported that a significant number of elderly patients with acute dementia, as well as chronic alcoholics, showed not only a higher than normal transketolase activation coefficient (indicative of thiamin deficiency), but also showed a further increase in enzyme activity when erythrocyte lysates were incubated with 3 mmol thiamin/d, compared with the usual 0.3 mmol/l used for the transketolase activation test. By contrast, Nixon et al. (1984) reported no difference in the affinity of transketolase for TDP between patients with Wernicke-Korsakoff syndrome and controls. They demonstrated a variety of different patterns of electrophoretically-separable forms of transketolase in normal subjects; one pattern was found in thirty-nine of forty-two patients with the Wernicke-Korsakoff syndrome, but only eight of thirty-six control subjects. However, Blansjaar et al. (1991) reviewed a series of investigations and concluded that there was little evidence to support a genetic abnormality of transketolase associated with Wernicke-Korsakoff syndrome.

McCool et al. (1993) and Martin et al. (1995) demonstrated that multiple electrophoretically-separable forms of transketolase were not the result of variant alleles, tissue-specific iso-enzymes or differential splicing of mRNA, and suggested that there were differences in either the assembly of the functional holoenzyme or post-synthetic modification
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do you not estimates of biotin requirements, and no reference
nutrient intake or recommended dietary allowance; the ‘safe
and adequate range of intake’ is derived from observed
average intakes, which are, obviously, more than adequate
to prevent deficiency.

There is some evidence that inadequate biotin nutrition
may be a factor in sudden infant death. Johnson et al. (1980)
reported that the liver biotin content in children who had
died from no known cause was approximately 75 % that in
children who had died from a known cause. The fatty liver
and kidney syndrome, which can cause sudden death of
flocks of chicks, is associated with biotin deficiency
(Bannister, 1976), and biotin deficiency can lead to skin
and hoof lesions in pigs.

Biotin functions as the coenzyme for four carboxylases:
pyruvate carboxylase (EC 6.4.1.1), acetyl-CoA carboxylase
(EC 6.4.1.2), propionyl-CoA carboxylase (EC 6.4.1.3) and
methylcrotonyl-CoA carboxylase (EC 6.4.1.4). Much of our
knowledge of the metabolic disturbances that may be
associated with biotin deficiency, and which might therefore
provide sensitive markers of status, has come from studies
of children with multiple carboxylase deficiency as a result
of genetic diseases which lead to functional deficiency
despite an adequate dietary intake. Holocarboxylase
synthetase (EC 6.3.4.10) catalyses the covalent attachment
of biotin to a lysyl residue in the carboxylase apo-enzymes;
lack of this enzyme results in total absence of all four
carboxylases, and is fatal in early life (Nyhan, 1987). When
biotin-containing enzymes are catalysed, the biotin is
released as biocytin (biotinyl ε-aminolysine); this is
normally hydrolysed by biotinidase (EC 3.5.1.12), and there
is considerable conservation of the biotin released in this
way. Children who lack biotinidase again suffer from multi-
carboxylase deficiency, but can be treated with relatively
large supplements of biotin, to replace that lost as biocytin
rather than it being salvaged. However, they do suffer some
long-term neurological damage (Hymes & Wolf, 1996).

In addition to its coenzyme role, biotin also has effects on
gene expression; it reacts with, and binds covalently to,
histones. Biotinidase seems to be important here, acting
both as an intracellular biotin-binding protein and to
catalyse the covalent attachment to histones (Hymes et al.
1995; Hymes & Wolf, 1996). There is some evidence that
biotinidase of histones is important in embryological
development; biotin deficiency is strongly teratogenic in
experimental animals (Watanabe & Endo, 1984; Watanabe
et al. 1995).

Chauhan & Dakshinamurti (1991) have shown that biotin
also acts, relatively specifically, to induce the synthesis
of glucokinase in fasted rats. Glucokinase (EC 2.7.1.2) is an
isoenzyme of hexokinase (EC 2.7.1.1) with a high Michaelis
constant (Km) for glucose in the liver, glucokinase is
responsible for the increase in glucose uptake and metabo-
lism in the fed state; hexokinase has a Km of 0.15 mmol/l,
and is therefore saturated with glucose, and acting at a
constant rate under all physiological conditions, whereas
glucokinase, with a Km of 20 mmol/l, only has significant
activity when the concentration of glucose in the portal
blood rises after a meal (Stryer, 1995). Apart from the liver,
the only other tissue in which glucokinase is known to be
expressed is the β-islet cells of the pancreas. Freguia
et al.
(1985) showed that in biotin-deficient chickens there was a greater proportion of the dose was exhaled as 14CO2. It is unlikely that this finding could be exploited as a test of optimum biotin nutritional status. Kopinski et al. (1989) showed that the impairment of fatty acid elongation in biotin-deficient pigs with normal desaturase activity led to an increase in the monounsaturated fatty acid value in liver lipids. While there may be similar effects on plasma lipid composition, this ratio is probably too susceptible to the effects of dietary fat intake to provide a useful index of biotin status.

Propionyl-CoA arising from a number of sources, including the metabolism of isoleucine, methionine, the side-chain of cholesterol and (rare) dietary odd-chain fatty acids, is normally carboxylated by propionyl-CoA carboxylase to yield methylmalonyl-CoA, which then undergoes isomerization to succinyl-CoA, a citrate cycle intermediate. Impairment of propionyl-CoA carboxylase leads to urinary excretion of propionate and onward metabolites such as hydroxypropionate, propionyl glycine and methyl citrate. Detection of these acids in urine may therefore provide an index of biotin deficiency; they are not normally detectable.

Thus, it is possible that the ability to metabolize a test dose of [13C]propionate may provide a sensitive index of status. Barshop et al. (1991) reported that control subjects exhaled between 46 % and 70 % of a test dose of [13C]propionate as 13CO2; however, apart from the endogenous sources of propionate, intestinal bacteria provide a considerable but variable amount, which would lead to unpredictable dilution of the isotope and confound interpretation of the results.

Propionyl-CoA can also compete with acetyl-CoA for incorporation into fatty acids, leading to the formation of odd-chain fatty acids. Mock et al. (1988) reported an increased proportion of 15:0 and 17:0 fatty acids in plasma lipids of patients who were biotin deficient as a result of prolonged total parenteral nutrition, which was normalized by administration of biotin. Proud et al. (1990) showed that the proportion of odd-chain fatty acids from 15:0 to 25:0 was increased in skin lipids from biotin-deficient rats. This finding suggests that measurement of odd-chain fatty acids in skin or serum lipids might be a useful index of optimum biotin status. However, dietary intake is probably more important than endogenous synthesis of odd-chain fatty acids. Watkins (1988) measured 17:0 levels in liver and heart triacylglycerol from biotin-deficient animals fed on four different dietary sources of fat: maize oil; hydrogenated soya bean oil; a mixture of trisolein, tristearin and tripalmitin; spent restaurant grease. The differences due to dietary fat were considerably greater than those due to biotin deficiency.

Methylcrotonyl-CoA is an intermediate in the metabolism of leucine; it is normally carboxylated, leading eventually to the formation of acetyl-CoA and acetoacetate. Impairment of methylcrotonyl-CoA carboxylase activity leads to urinary excretion of methylcrotonate and two onward metabolites, methylcrotonyl glycine and 3-hydroxyisovalerate. The latter acid is an early marker of experimental biotin deficiency in rats (Mock & Mock, 1992). 3-Hydroxyisovalerate is normally excreted in detectable amounts (some 25–42μmol/24h). In a human study in which avidin was fed in order to cause biotin depletion, Mock et al. (1997) showed that urinary 3-hydroxy-...
isovalerate excretion increased 4-fold, while plasma biotin was still (just) within the reference range. Measurement of 3-hydroxy-isovalerate excretion, perhaps combined with a loading dose of its precursor leucine, may provide a sensitive test of optimum biotin nutritional status.

**Pantothenate**

As with biotin, pantothenate deficiency is more or less unknown. The so-called burning foot syndrome (nutritional melalgia) in severely-malnourished prisoners of war in the Far East is often assumed to have been due to pantothenate deficiency, but for obvious reasons they were repleted with yeast extract as a source of all B-vitamins rather than being used for more precise experimental studies.

Early studies showed that there was loss of fur colour in black and brown rats fed on a pantothenate-deficient diet, and at one time it was known as the anti-grey hair factor. Despite the fact that there is no evidence that loss of hair colour with age in human subjects is related to pantothenate nutrition, it is still added to shampoos. Matsumoto et al. (1994) reported that an early sign of pantothenate deficiency in rainbow trout (Oncorhynchus mykiss) is anorexia; neither lack of anorexia nor a voracious appetite could sensibly be used as an index of optimum nutrition.

Pantothenate is pantoyl γ-alanine; the next higher homologue, pantoyl γ-aminobutyrate (pantoyl GABA, homopantothenate or hopanthenate), is used in Japan to enhance cognitive function, especially in Alzheimer’s disease. It acts via GABA receptors to increase acetylcholine release and cholinergic function in the cerebral cortex and hippocampus (Nakahiro et al. 1985). A rare side effect is the development of hepatic encephalopathy (Noda et al. 1991), and the excretion of a variety of dicarboxylates which arise by way of CoA-independent α- and γ-oxidation of fatty acids (Matsumoto et al. 1991). Both the encephalopathy and the dicarboxylic aciduria are reversed by pantothenate; suggesting that homopantotheinate may cause pantothenate deficiency. If this suggestion is verified, dicarboxylic aciduria may provide a marker of pantothenate status. However, it is equally possible that the adverse effects are due to a toxic action of homopantotheinate which is antagonized by pantothenate.

Pantothenate has two well-defined metabolic roles: in CoA in fatty acid oxidation acetate metabolism, cholesterol and steroid synthesis, and acetylation of drugs; as the prosthetic group of acyl carrier protein in fatty acid synthesis. Thus, it would seem that the ability to acetylate drugs such as sulphamidimine and isoniazid might be a useful test for pantothenate status, and certainly deficiency does impair drug acetylation (Petrizik et al. 1975). There is a genetic polymorphism in the N-acetyltransferase (EC 2.3.1.5) that is involved in drug acetylation, and in different ethnic groups between 20 and 60 % of the population are genetically slow acetylators. Vas et al. (1990) showed that the administration of 1100 mg pantothenate daily for 7 d had no effect on the pharmacokinetics of sulphamidimine, and did not affect the determination of acetylator phenotype.

It is unlikely that fatty acid oxidation would be significantly affected by pantothenate status. Reibel et al. (1982) showed that despite a very considerable reduction in tissue pantothenate in deficient rats, there was little or no fall in tissue CoA. Both fasting and induction of diabetes by administration of alloxan led to similar increases in tissue CoA in control and pantothenate-deficient rats. Smith et al. (1987) did report a small fall in tissue CoA in pantothenate-deficient rats, and a greater depletion of glycogen reserves during exercise, but the deficient animals showed normal ketogenesis and ketone utilization in exercise. They noted that deficient animals became exhausted more rapidly during exercise, but this is unlikely to provide a useful test of pantothenate status for the human population at large.

The preservation of tissue CoA in pantothenate deficiency is presumably the result of both control over CoA synthesis and efficient conservation of phosphopantetheine arising from catabolism of CoA and acyl carrier protein. Köchbaum et al. (1990) showed that the increase in liver CoA in fasting in both control and genetically-diabetic mice was due to changes in the activity of pantothenate kinase (EC 2.7.1.13). This enzyme is subject to feedback inhibition by CoA and acyl-CoA, repression by insulin (in the fed state when the need for CoA is lower) and induction by glucagon (in the fasting state when there is a greater requirement for CoA). The availability of pantothenate is unlikely to affect the rate of incorporation into CoA, since the Km of the enzyme is 18 µmol/l, and even in severe experimental deficiency most tissues maintain a higher concentration. There is rapid turnover of the prosthetic group of acyl carrier protein (a half-life of the order of 6 h) compared with a half-life of apo-acyl carrier protein (6-4; Volpe & Vagelos, 1973). The phosphopantetheine released from acyl carrier protein is incorporated into CoA, which in turn activates apo-acyl carrier protein as required; presumably the enzymes responsible for this cycling of phosphopantetheine are under hormonal regulation.

Wittwer et al. (1990) showed that both plasma triacylglycerol and non-esterified fatty acids are moderately elevated in pantothenate-deficient rats, however, this finding is unlikely to provide the basis for a test of pantothenate status in human subjects.

**Summary**

Especially if there is a genetic polymorphism in transketolase that affects thiamin requirements, there is a need to re-examine current estimates of requirements; probably the only index of status that is available is the transketolase activation assay. While Thurnham (1981) questioned the physiological significance of marginal vitamin deficiency as determined by enzyme activation assays, transketolase activation is at least a marker that can be used to assess saturation of one physiological system. In the case of biotin, it is obviously desirable to have an estimate of requirements or desirable levels of intake; either saturation of erythrocyte pyruvate-CoA carboxylase with its coenzyme or, preferably, 3-hydroxy-isovalerate excretion (perhaps after a test dose of leucine) will provide an index of adequacy.

There still seems to be no way of assessing pantothenate status.


