A Reexamination of Aluminum in Alzheimer’s Disease: Analysis by Energy Dispersive X-ray Microprobe and Flameless Atomic Absorption Spectrophotometry

Roland W. Jacobs, Taihung Duong, Robert E. Jones, George A. Trapp and Arnold B. Scheibel

ABSTRACT: We have attempted to verify the presence of increased aluminum (Al) levels in Alzheimer’s disease (AD) brains by energy dispersive X-ray microanalysis (EDX) and flameless atomic absorption spectrophotometry (AAS). Tissue from seven AD brains, mounted on carbon polymerized coverslips, were stained with Congo-red or treated immunohistochemically to allow optical localization of AD-associated lesions during EDX. Despite a demonstrated sensitivity of 20-25 ppm, we were unable to detect Al in either plaque cores or neurons containing neurofibrillary tangles. For AAS, wet weight samples (ranging from 48-144 mg) from six of the seven AD brains and four controls were selected from regions similar to those studied under EDX, i.e., Brodmann areas 9, 11, 28, 46, 47, and the hippocampus. The tissue surrounding each sample site was sectioned and stained for thioflavin S. Both controls and AD samples revealed similar levels of Al ranging from undetectable to 1.80 ng/mg wet wt. (mean AD: 0.28 ± 0.39 (SD), control: 0.54 ± 0.58 (SD)), independent of degree of histopathology or age of the case. We conclude that the combined strengths of these two techniques on similar tissue specimens demonstrate that abnormal Al levels are not required to produce the histologic findings of AD and that this element may not accumulate in the aging brain. It is unlikely, therefore, that Al is essential in the etiology of pathogenesis of plaques and tangles in AD. Al’s role as a primary or secondarily associated event, when present, needs further delineation.
The induction of neuronal cytoplasmic “tangles” by Al in rabbits and reports of increases in brain Al in AD in the early 1970's initiated the current debate. These early findings were countered by subsequent studies which argued that: 1) brain Al content increases as a function of age irrespective of diagnosis; 2) the impact of atrophy and localized cell death were not considered in the AD group; the ultrastructure of induced rabbit neurofibrillary tangles (NFT) is significantly different from those of AD and they are formed from antigenically distinct cytoplasmic substrates.

Further controversy ensued with the demonstrated toxicity of Al in dialysis dementia and reports of Al in NFT-bearing hippocampal neurons in the amyotrophic lateral sclerosis-Parkinsonism dementia complex (ALS/PD) of the Pacific basin rim. Although the presence of Al in these disorders could indicate a common pathogenic process, the presence of early motor impairment in animal NFT-induction models, dialysis dementia, and ALS/PD argues against a pathogenic similarity with AD. By contrast, AD is characterized clinically and neuropathologically by relative sparing of motor systems (e.g. primary motor cortex, basal ganglia, and anterior horn motor neurons) until the latter stages.

In recent years, sophisticated probing techniques have been employed to counter the claim that gross Al brain content is solely a function of age. Perl and Brody presented evidence of modest but significant Al levels in hippocampal NFT-bearing neurons by energy dispersive X-ray microanalysis (EDX). Crapper et al demonstrated Al in chromatin and faulted prior controversial brain Al studies on selection of excessively large sample sizes, improperly diagnosed AD cases, and analytical methodology. Candy et al, using EDX and nuclear magnetic resonance, found alumino-silicates in amyloid plaque cores in AD brain tissue. Using similar techniques, however, Selkoe et al and Moretz et al were unable to demonstrate a significant presence of Al in the AD-associated lesions tested.

With access to an EDX, the Cameca Camebax, and a Varian AA-275 atomic absorption spectrophotometer (AAS), we set out to detect the presence of Al in unfixed, ultrafrozen AD diagnosed brains and in neurologically unimpaired controls. The protocol for the AAS includes a recently developed matrix modifier that significantly enhanced sensitivity and accuracy.

Similarly, with the technique described below for the EDX, we were able to improve its ability to detect trace elements. Thus, by combining the strength of site specificity of the EDX with the sensitivity of AAS between comparable brain regions, we demonstrated no increased presence of Al in AD. Our earlier results with EDX indicated the same, revealing a total of only one Al positive probe in 243 probes of 146 NFT, one in 667 probes of 282 nuclei, and one in 194 probes of 63 senile plaque cores.

**Materials and Methods**

Freshly frozen (−70°C; mean autolysis time of 13 hrs) human brain specimens from seven clinically diagnosed late stage probable AD cases (mean 80 yrs; range 68-102) and four controls (mean 66 yrs; range 58-78) were obtained from the National Neurological Research Bank, VA Wadsworth Medical Center. The AD cases were confirmed neuropathologically. Medical records were reviewed to insure that the controls were neurologically intact and maintained an unimpaired mental status until time of death.

**EDX Analysis**

Tissue blocks from the frontal cortex (Brodmann areas 8, 11, 46, and 47) and temporal lobe (area 28), including the hippocampus, were fixed by immersion in 4% buffered paraformaldehyde at 4°C for 24-48 hours and rinsed in 0.12 M Millonig’s phosphate buffer and 30% sucrose (pH 7.4) with gentle agitation for an additional 24 hours. The tissue was cut at 40 μm on a cryostat and collected in 0.1 M Tris buffer (pH 7.4). Alternative sections were stained with Congo red/toluidine blue and Bielchowsky’s reduced silver technique to identify selectively the plaques and tangles characteristic of AD. In addition, we introduced into the sequence a novel immunocytochemical label, amyloid P-component (AP-C), as recently described by Scheibcl et al. The advantage of this stain is its strong demonstration of senile plaques and NFT under standard light microscopy. During EDX analysis, both AP-C and Congo red were free from artifact and did not significantly alter the elemental profile as found with Bielchowski and other silver-containing, staining techniques. The prepared sections were mounted on glass slides or carbon polymerized plastic slides (Thermanox tissue culture cover slips: 5410, Lab-Tek Division, Miles Laboratories, Inc., Naperville, IL 60540).

We employed a Cameca Camebax scanning electron microscope (SEM) and electron microprobe in tandem with a Kevek 8000 energy detector (energy dispersive spectroscopy) with a standard beryllium window and a “take-off” angle of 40 degrees. The Cameca allows for simultaneous optical visualization of the tissue sample with a 400x binocular transmitted incandescent light microscope with polarizing capability. Mounted sections, lightly carbon-coated to avoid electron charging, were placed within the Cameca’s vacuum chamber and viewed by optical microscopy. Specific sites on identified neurons, plaques, and tangles were then selected under SEM for EDX. To ensure that “ghost cells” were not included in the sampling, only neurons with an identifiable nucleus were chosen. The microprobe data were obtained by narrowing an electron beam of specific current and voltage over the probe site for a specified length of time (acquisition time) during which X-ray counts were acquired.

Microprobe sensitivity was determined by preparing serial standard dilutions of Al (from reagent grade AlCl₃) and placing a known quantity over a weighed, predetermined area of a 40 μm thick cortical “control” tissue section mounted on a plastic slide. Sensitivity in parts per million (ppm) was obtained by determining the least amount of Al required to illicit consistently an unequivocal peak (Figure 1). In addition, homogenized pine needles from the National Bureau of Standards (standard reference material 1575) were microprobed (Figure 2a). The certified value of Al in the standard reference material is 545 ppm. Al peaks as determined by the various microprobe parameters utilized during this study are shown in Figure 1.

Initially, we began by utilizing the EDX technique as described by prior investigators. We employed the EDX technique in tandem with a Kevek 8000 energy detector (energy dispersive spectroscopy) with a standard beryllium window and a “take-off” angle of 40 degrees.

**Materials and Methods**

Freshly frozen (−70°C; mean autolysis time of 13 hrs) human brain specimens from seven clinically diagnosed late stage probable AD cases (mean 80 yrs; range 68-102) and four controls (mean 66 yrs; range 58-78) were obtained from the National Neurological Research Bank, VA Wadsworth Medical Center. The AD cases were confirmed neuropathologically. Medical records were reviewed to insure that the controls were neurologically intact and maintained an unimpaired mental status until time of death.
modest sized peaks of silicon were obtained. Under plastic
slides nearly all probes were negative for silicon.
Our final parameter (5 KeV, 40 nA, 400 sec) achieved a sen­
sitivity of 20-25 ppm. The lower voltage allowed the energy
detector to focus on low energy X-rays, including Al, by block­
ing out a significant amount of extraneous, competing signals.
Thus, when present, Al was proportionately stronger and rose
more quickly above background noise. Sampling by a
6.25 µm²/probe site was selected to search for Al’s presence
adequately in identified structures without extending into adja­
cent locations. In addition, the risk of possible elemental migra­
tion during data acquisition was avoided. One probe was per­
formed per structure identified, i.e., NFT, nuclei of NFT-bearing
neurons, and senile plaque cores. All probes were performed on
tissue mounted on plastic slides.

AAS Analysis
Freshly frozen grey matter samples, weighing 48-144 mg wet
wt., were taken from Brodmann areas 8, 11, 28, 45, 46, and hip­
pcampus from six of the original seven AD specimens and
from all four controls. Extreme care was exercised to minimize
possible Al contamination during handling of the tissue, includ­

![Figure 1](https://www.cambridge.org/core/terms). IP address: 54.70.40.11, on 10 Jun 2018 at 17:10:08, subject to the Cambridge Core terms of use, available at https://www.cambridge.org/core/terms. https://doi.org/10.1017/S0317167100029838
ing use of non-Al containing utensils, storage of deionized distilled water in nalgene containers, observing precautions against dust contamination, and washing of quartzware and plasticware in 6N HNO₃ and 0.01M EDTA solution. All solutions were stored in teflon bottles and reagents were either of analytical reagent grade or purified by a technique appropriate for that solution. Nitric acid was redistilled from quartz. Sulfuric and perchloric acids were trace metal grade purchased from GFS Chemicals, Columbus, Ohio.

The remaining tissue blocks from which the various samples were drawn were fixed in 10% buffered formalin. Selected 40 μm sections were stained and analyzed with thioflavin S for density of plaques and NFT. Each AD sample was then categorized, based on surrounding histopathology, as “light”, “moderate”, or “heavy” (Figure 3). All control samples were similarly analyzed and found to be free of pathology. This index allowed us to compare Al levels not only across diagnostic categories but by intensity of pathology.

Each sample was digested in a quartz test tube containing 250 μl of a mixture of HNO₃/HClO₄/H₂SO₄ (10/0.5/0.25). Temperature was slowly raised to 170°C to avoid foaming and to remove HClO₄. The digest was diluted with 200 μl of a matrix modifier composed of 5.0M NH₄NO₃, 0.2M Ca(NO₃)₂, 0.3M H₃PO₄, and LiNO₃. This matrix modifier has been shown to obviate the need for standard additions methodology.

Al was measured by atomic absorption spectrophotometry at 309.3 nm with a Varian AA-275 equipped for flameless technique with carbon rod atomizer CRA-90. The instrument was operated in the absorbance peak height mode and output peaks were recorded by a Varian 9176 strip chart recorder. Deuterium background correction was used. Sample size was 2 μl. Purge gas was argon and small amounts of hydrogen were added during the ash and atomize cycles. The dry cycle was 120°C X 40 sec, ash was 1500°C X 30 sec, and atomization was at 2500°C. The acid digestion blank was 5.8 ng Al ± 1.8 ng and was subtracted from experimental values. The sensitivity was 10 ng Al per tissue sample or two times the blank. Recovery from acid digests was 95-105%; analysis of NBS SRM 1575 (pine needles) yielded 95.8% ± 7.8% of expected value. Details of analysis of tissue for Al will be published elsewhere.

**RESULTS**

Brain specimens from severely demented AD patients and normal controls were analyzed for the presence of Al by two sensitive and complementary techniques. Energy dispersive X-ray microanalysis with an Al determination as sensitive as 20-25 ppm was employed in site specific analysis of the neuron and senile plaque. Flameless atomic absorption spectrophotometry of histopathologically characterized, low weight samples resulted in detection levels as sensitive as 10 ng Al per tissue sample or two times the blank.

No Al was detected by EDX (sensitivity: 20-25 ppm) in 105 NFT, 70 nuclei, and 56 senile plaque cores sampled from seven AD brain specimens mounted on plastic slides, representing Brodmann areas 8, 11, 28, 46, 47, and the hippocampus (Figures 2b, 2c). Minimal levels of silicon were occasionally found. With the 50 ppm sensitivity technique, probes of 45 NFT, 58 nuclei, and 14 plaques cores were also negative for Al. AAS failed to find any elevation of Al in samples taken from these areas in the six analyzed AD brains as compared to controls (Table 1). There was no correlation between density of plaques and or tangles and Al levels (Table 2). Similarly, no correlation was demonstrated between brain Al and age (Table 1).

**Table 1: AAS: Sample Comparison of AD to Control**

<table>
<thead>
<tr>
<th>Dx</th>
<th>Case #</th>
<th>Age</th>
<th>Sex</th>
<th># of samples</th>
<th>Mean Al</th>
</tr>
</thead>
<tbody>
<tr>
<td>AD</td>
<td>1</td>
<td>102</td>
<td>F</td>
<td>8</td>
<td>0.40 ± 0.56</td>
</tr>
<tr>
<td>AD</td>
<td>2</td>
<td>74</td>
<td>F</td>
<td>7</td>
<td>0.32 ± 0.32</td>
</tr>
<tr>
<td>AD</td>
<td>3</td>
<td>81</td>
<td>F</td>
<td>11</td>
<td>0.15 ± 0.22</td>
</tr>
<tr>
<td>AD</td>
<td>4</td>
<td>68</td>
<td>F</td>
<td>8</td>
<td>0.16 ± 0.25</td>
</tr>
<tr>
<td>AD</td>
<td>5</td>
<td>80</td>
<td>M</td>
<td>4</td>
<td>0.12 ± 0.15</td>
</tr>
<tr>
<td>AD</td>
<td>6</td>
<td>75</td>
<td>M</td>
<td>10</td>
<td>0.25 ± 0.34</td>
</tr>
<tr>
<td><strong>TOTALS</strong></td>
<td></td>
<td>48</td>
<td></td>
<td></td>
<td>0.28 ± 0.39</td>
</tr>
<tr>
<td>C</td>
<td>1</td>
<td>58</td>
<td>M</td>
<td>9</td>
<td>0.45 ± 0.52</td>
</tr>
<tr>
<td>C</td>
<td>2†</td>
<td>61</td>
<td>M</td>
<td>2</td>
<td>0.68 ± 0.75</td>
</tr>
<tr>
<td>C</td>
<td>3</td>
<td>71</td>
<td>M</td>
<td>6</td>
<td>0.44 ± 0.68</td>
</tr>
<tr>
<td>C</td>
<td>4</td>
<td>74</td>
<td>M</td>
<td>5</td>
<td>0.75 ± 0.66</td>
</tr>
<tr>
<td><strong>TOTALS</strong></td>
<td></td>
<td>22</td>
<td></td>
<td></td>
<td>0.54 ± 0.58</td>
</tr>
</tbody>
</table>

†Occasional plaque cores were present in Area 28. Remaining samples exhibited no pathology.
Our data fail to demonstrate the presence of elevated amounts of Al in AD brain tissue. Careful in situ analysis of neurons and senile plaque cores by a site-specified technique has not supported prior findings. Tandem investigation by a sensitive bulk analysis technique has also failed to support evidence that aluminum (Al) selectively accumulates in the neurofibrillary tangle-bearing neurons. One possible explanation for these unexpected results is that Alzheimer's disease and aging in our sample of ten with an age range of 58 to 102 yrs.

One possible explanation for these unexpected results is that brain Al may vary among AD patients and that we obtained seven that did not have abnormal amounts. We are led by our current findings to conclude that Al may not play an etiologic or pathogenic role in the formation of the characteristic "plaques and tangles" of Alzheimer's disease and, when present, may only be an associated phenomenon.

ACKNOWLEDGEMENTS

Tissue for this research was obtained from W.W. Tourtellotte M.D., Ph.D., director of the National Neurological Research Bank, VA Wadsworth Medical Center, Los Angeles, CA, U.S.A. 90073, which is sponsored by NINCDS/NIMH, NMSS, HD Foundation, and Veterans Administration. Sincere thanks to Iris Rosario R.N. and Randel Thomsen. Special thanks to Bob Jacobs for his helpful suggestions in preparation of the manuscript and to Enesto Pommier for superior technical assistance. Personal support for Roland Jacobs was provided by Valley Hospital Medical Center. Invetisgational support provided, in part, by the Eleanor Leslie Fund.

REFERENCES