Livestock veterinarians at high risk of acquiring
methicillin-resistant Staphylococcus aureus ST398

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SUMMARY

The prevalence and risk factors associated with livestock-associated MRSA (LA-MRSA) carriage
was examined in Danish and Belgian veterinarians. The MRSA and LA-MRSA carriage rates
were 9.5% (95% CI 5.3–15.6) and 7.5% (95% CI 3.8–13.1) for MRSA and LA-MRSA,
respectively, in Belgium and 1.4% (95% CI: 0.17–5.05) in Denmark (all Danish MRSA isolates
belonged to the LA-MRSA genotype). All LA-MRSA isolates were resistant to tetracycline and
53.4% (7/13) showed a multi-resistant phenotype. LA-MRSA was significantly associated with
veterinarians in contact with livestock ($P = 0.046$). In the multivariable analysis, working with
small animals in a veterinary clinic seems to be negatively associated (OR 0.15, 95% CI 0–1.0,
$P = 0.05$) and a strong direct association was found for LA-MRSA acquisition and exposure
to live pigs (OR 12.1, 95% CI 1.6–548.5, $P = 0.01$). Since carriage of MRSA ST398 may increase
the risk of complications during hospitalization, our results underline that preventive measures
may need to be developed for veterinary professionals, particularly for livestock veterinarians.

Key words: Exposure to live pigs, livestock-associated MRSA, pig veterinarians, risk factors.

INTRODUCTION

In 2003, a new methicillin-resistant Staphylococcus aureus (MRSA) clone emerged in livestock, particularly pigs, and in individuals in contact with livestock [1, 2]. Subsequently, this livestock-associated MRSA (LA-MRSA) clone has been identified throughout Europe [3], Canada [4], and the USA [5]. The analysis of LA-MRSA isolates identified four features that are characteristic for LA-MRSA: (i) they are non-typable by pulsed-field gel electrophoresis (PFGE) using SmaI [6]; (ii) they belong to clonal complex (CC) 398 by multi-locus sequence typing (MLST) [7]; (iii) the staphylococcal chromosomal cassette (SCC)mec types IV and V predominate but are structurally different from the corresponding types carried by MRSA genotypes endemic in community and healthcare settings [8]; and (iv) most isolates lacks toxins such as Panton–Valentine leukocidin (PVL) and other enterotoxins [9].
LA-MRSA seems to be less virulent and less transmissible than community- and healthcare-associated MRSA [10–12], although secondary transmission to household members, people and animals in contact has been reported [13]. In a Danish case-control study, a relatively high proportion of case-patients reported skin and soft tissue infections (10/21), one patient developed sinusitis, and another developed bacteraemia after knee surgery [14].

In Europe, geographical variation in the number of human LA-MRSA cases (both asymptomatic carriage and infections) is directly linked to pig and veal calf densities [15]. In areas where LA-MRSA is endemic in the agricultural setting, the carriage rate in pig farm workers is higher than in any other population, including patients with predisposing risk factors such as exposure to foreign healthcare facilities [10]. In countries with a low incidence of MRSA, this epidemiological change threatens existing control policies developed to prevent spread of MRSA in the community and into hospitals. For example, a Dutch survey conducted during 2002–2006 found that 32% of patients carried MRSA on admission to a hospital located in an area with a high pig density compared to a nationwide incidence of 0.03% in 2000 [10].

In this study, we assessed the prevalence of LA-MRSA in veterinarians from two countries, Belgium and Denmark, representing areas with high and low proportions (> tenfold difference) of positive pig farms, respectively, as previously reported [3]. We hypothesized that veterinarians working with livestock constitute an occupational risk group due to frequent contact with livestock and that the individual risk for LA-MRSA carriage is correlated with the proportion of LA-MRSA in the agricultural setting. For appropriate adaptation of control and protective measures specific for veterinarians (and indirectly for their household members), a variety of potential risk factors modified according to their profession were also examined.

MATERIALS AND METHODS

Study design and population

A cross-sectional prevalence study was conducted from February to April 2010 in Belgium and from February to October 2010 in Denmark. A total of 1215 veterinarians (Belgium, n = 800; Denmark, n = 415) were invited to participate in the study after receiving full information from the investigators. In Belgium, veterinarians were randomly selected from two registers, Nederlandstalige Orde der Dierenartsen and Conseil regional francophone de l’Ordre de Médecin Vétérinaires. In Denmark, two groups of veterinarians were invited to participate in the study: all veterinary practitioners with specialization for particular animals (i.e. pigs, cattle, horses, companion animals) registered with the Danish Veterinary Association (n = 279) and all veterinary meat inspectors, responsible for inspection of live animals before slaughter, registered with the Danish Veterinary and Food Administration (n = 136).

The study was approved by the Ethics Committee of the ULB-Erasme Hospital (protocol no. P2009/244) or the Danish National Committee on Biomedical Research Ethics (protocol no. H-4-2009-112), and the Danish Data Protection Agency (protocol no. 2009-54-0821). The volunteers signed an informed consent form and were asked to agree to self-administered nasal swabbing and to answer a standard questionnaire, which was designed to capture demographic, lifestyle, and environmental data as well as information on known/identified risk factors for MRSA (including medical history, antibiotic usage, contact sports, and travel) and animal-related exposures (including living or working on a farm, living with a farm worker, intensity and type of exposure to animals, and hygienic/protective measures). Nasal swabs (Venturi Transystem, Copan Innovation, Italy) were taken by the veterinarians themselves according to the manufacturer’s instructions and were sent to Erasme Hospital (Brussels, Belgium) or to Statens Serum Institut (Copenhagen, Denmark) in transport medium.

MRSA isolation and characterization

Nasal swabs were analysed individually. MRSA isolation was performed using standard methods described previously [13]. Identification of all S. aureus isolates, including presence of the mecA gene (confering resistance to β-lactam antibiotics) and lukF-lukS genes (encoding PVL), was performed using a multiplex PCR assay [16]. Isolates were further characterized by MLST typing [7], spa typing [17], and SCCmec typing [18]. The presence of 10 resistance genes [tet(K), tet(M), aac(6')-Ie + aph(2'), ant(4')-Ia, aph(3')-IIIa, aadC, ermA, ermC, msrA/mrsB, vatB] was investigated in all isolates, using multiplex PCR assays and primers that have been described previously [19–21].
Antimicrobial susceptibilities

The antimicrobial susceptibility profiles (spectinomycin, gentamicin, kanamycin, tobramycin, rifampin, trimethoprim-sulfamethoxazole, clindamycin, erythromycin, linezolid, chloramphenicol, mupirocin, ciprofloxacin, minocycline, tetracycline, fusidic acid) of all isolates were determined by the Kirby–Bauer disk diffusion method using Neo-Sensitabs (Rosco, Denmark), in accordance with the Clinical Laboratory Standards Institute (CLSI) guidelines with modifications for interpretation according to the manufacturer’s instructions (details can be found in the manufacturer’s database) [22]. Vancomycin susceptibility was determined onto brain heart infusion agar (BHIA). Briefly, 10 μl of a 0.5 McFarland suspension was spotted onto BHIA supplemented with 2 and 3 μg/ml vancomycin and incubated at 35 °C for 48 h. Any growth was interpreted as positive. Multidrug resistance was defined as resistance to ≥4 non-β-lactam antimicrobial classes.

Statistical analysis

In a first model, it was attempted to identify risk factors using a univariable analysis. Therefore a single-factor exact logistic regression was performed for the presence or absence of MRSA/ST398 as binary outcome. The variables with a P value <0.25 were withheld for the multivariable regression model. The multivariable model was built forwards, gradually including the variables to the model and only retaining the significant factors. Interactions were checked for all significant main factors in the model. A median-unbiased estimator (MUE) was used to estimate odds ratios in case of null values.

For both models, the significance level was set at 5% (P≤0.05). All models were built in Stata version 10.0 (Stata Corporation, USA).

RESULTS

Study population

A total number of 289 veterinarians signed the informed consent form and returned the nasal swab and questionnaire. The response rates were 18.3% (146/800) and 34.4% (143/415) for Belgium and Denmark, respectively. In Belgium, volunteering rates differed between occupational groups and were lower for veterinarians lacking exposure to livestock. The proportions of veterinarians working with livestock (defined in this study as professional contact on a daily basis with pigs, ruminants, and poultry) were 71.9% and 67.8% for veterinarians from Belgium and Denmark, respectively.

Molecular and phenotypic characterization

Of the 289 veterinarians, 16 carried MRSA (Belgium, n=14; Denmark, n=2) of which 13 belonged to MLST type ST398 (Belgium, 11/14; Denmark, 2/2), whereas the remaining three isolates from Belgian veterinarians belonged to other MLST types (ST1, ST5, ST45) (Table 1). All isolates lacked the lukF-lukS genes (encoding PVL). The spa and SCCmec types as well as resistance gene and antibiotic susceptibility profiles were heterogeneous within ST398 (Table 1). An important characteristic of these MRSA ST398 isolates was uniform resistance to tetracycline (13/13) due to the presence of the tet(M) determinant (13/13). Of note, the proportion of multidrug-resistant isolates was higher in ST398 MRSA isolates (7/13) than in non-ST398 MRSA isolates (0/3). However, this difference was not statistically significant (P=0.0901).

Risk factors for LA-MRSA carriage

The MRSA and LA-MRSA carriage rates were 9.5% (95% CI 5.3–15.6) and 7.5% (95% CI 3.8–13.1) for MRSA and LA-MRSA, respectively, in Belgium and 1.4% (95% CI 0.17–5.05) in Denmark (all Danish MRSA isolates belonged to the LA-MRSA genotype) which supports that Belgian veterinarians have a higher risk of MRSA carriage in general and of LA-MRSA carriage in particular than Danish veterinarians (Table 2).

Due to a lack of variance in the descriptive results for the Danish veterinarians combined with their low carriage prevalence found, further analysis was limited to the Belgian data. The female/male ratio and the mean age of Belgian participants were 0.52 and 45.1 years, respectively. Univariable analysis, with nasal carriage of LA-MRSA as the endpoint, showed that working with livestock was a significant risk factor (Table 3). In contrast, neither exposure to other animals nor risk factors defined by demographic, lifestyle, environmental characteristics, medical history, or antimicrobial exposure were found to be significantly associated with nasal carriage of LA-MRSA, except for male sex and living with a healthcare worker. Veterinarians working with pigs were at higher risk of LA-MRSA carriage than those
working with cattle. The highest risk was associated with exposure to live pigs. Interestingly, exposure to cattle within 24 h before sampling was a significant risk factor, whereas exposure to pigs within 24 h before sampling was not associated with carriage. No association was found to exposure to other farm animals, sheep, goats, poultry, neither to horses nor to companion animals. None of the following non-related occupational exposure variables, like smoking, living in the countryside, living with a farm worker, travel abroad, contact with healthcare institutions in the last 12 months, use of antimicrobials drugs during

### Table 1. Molecular and phenotypic characteristics of methicillin-resistant Staphylococcus aureus isolates from veterinarians in Belgium and Denmark

<table>
<thead>
<tr>
<th>Isolate ID</th>
<th>MLST type</th>
<th>spa type</th>
<th>SCCmec type</th>
<th>Resistance profile</th>
<th>Resistance genes</th>
</tr>
</thead>
<tbody>
<tr>
<td>BE-1</td>
<td>ST398</td>
<td>011</td>
<td>IV</td>
<td>GEN TOB KAN ERY CLI TET</td>
<td>aac(6’)-aph(2’) ermC tet(M)</td>
</tr>
<tr>
<td>BE-2</td>
<td>ST398</td>
<td>011</td>
<td>IV</td>
<td>GEN TOB KAN ERY CLI TET</td>
<td>aac(6’)-aph(2’) mrsA/mrsB ermC tet(M)</td>
</tr>
<tr>
<td>BE-3</td>
<td>ST398</td>
<td>011</td>
<td>IV</td>
<td>GEN TOB KAN SPC ERY CLI TET CHL</td>
<td>aac(6’)-aph(2’) ermC tet(K) tet(M)</td>
</tr>
<tr>
<td>BE-4</td>
<td>ST398</td>
<td>011</td>
<td>IV</td>
<td>GEN TOB KAN SPC ERY CLI TET</td>
<td>aac(6’)-aph(2’) ermC tet(M)</td>
</tr>
<tr>
<td>BE-5</td>
<td>ST398</td>
<td>011</td>
<td>IV</td>
<td>TOB KAN ERY CLI TET</td>
<td>aacD ermC tet(M)</td>
</tr>
<tr>
<td>BE-6*</td>
<td>ST398</td>
<td>011</td>
<td>V</td>
<td>TOB ERY CLI TET</td>
<td>tet(K) tet(M) aacD</td>
</tr>
<tr>
<td>BE-7</td>
<td>ST398</td>
<td>011</td>
<td>V</td>
<td>CIP TET</td>
<td>tet(K) tet(M)</td>
</tr>
<tr>
<td>BE-8</td>
<td>ST398</td>
<td>011</td>
<td>V</td>
<td>ERY CLI TET</td>
<td>ermC tet(M)</td>
</tr>
<tr>
<td>BE-9</td>
<td>ST398</td>
<td>034</td>
<td>IV</td>
<td>ERY CLI TET</td>
<td>ermC tet(K) tet(M)</td>
</tr>
<tr>
<td>BE-10</td>
<td>ST398</td>
<td>01451</td>
<td>V</td>
<td>ERY CLI TET</td>
<td>ermC tet(K) tet(M)</td>
</tr>
<tr>
<td>BE-11</td>
<td>ST398</td>
<td>16408</td>
<td>IV</td>
<td>GEN TOB KAN ERY CLI CIP TET</td>
<td>aac(6’)-aph(2’) ermC tet(M)</td>
</tr>
<tr>
<td>BE-12</td>
<td>ST1</td>
<td>127</td>
<td>IV</td>
<td>KAN, ERY, TET</td>
<td>tet(K) ermC aph(3’)</td>
</tr>
<tr>
<td>BE-13</td>
<td>ST5</td>
<td>002</td>
<td>I</td>
<td>TOB, KAN, TET, FUS</td>
<td>tet(K) aacD</td>
</tr>
<tr>
<td>BE-14</td>
<td>ST45</td>
<td>038</td>
<td>IV</td>
<td>CIP</td>
<td>None</td>
</tr>
<tr>
<td>DK-1</td>
<td>ST398</td>
<td>034</td>
<td>V</td>
<td>ERY CLI MIN TET</td>
<td>ermC tet(K) tet(M)</td>
</tr>
<tr>
<td>DK-2</td>
<td>ST398</td>
<td>034</td>
<td>V</td>
<td>CLI MIN TET</td>
<td>tet(K) tet(M)</td>
</tr>
</tbody>
</table>

BE, Belgium; DK, Denmark; MLST, multi-locus sequence typing; SCCmec, staphylococcal chromosomal cassette mec; GEN, gentamicin; TOB, tobramycin; KAN, kanamycin; SPC: spectinomycin, ERY, erythromycin; CLI, clindamycin; CHL, chloramphenicol; CIP, ciprofloxacin; MIN, minocycline; TET, tetracycline; FUS, fusidic acid.

* Erythromycin and clindamycin resistance could not be detected with the PCR used.

### Table 2. Carriage rates of methicillin-resistant Staphylococcus aureus isolates among veterinarians from Belgium and Denmark

<table>
<thead>
<tr>
<th>Exposure variable</th>
<th>No. (%)</th>
<th>OR (95% CI)</th>
<th>P value*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Belgium</td>
<td>Denmark</td>
<td></td>
</tr>
<tr>
<td>All veterinarians</td>
<td>(n = 146)</td>
<td>(n = 143)</td>
<td></td>
</tr>
<tr>
<td>MRSA ST398</td>
<td>11 (7.5)</td>
<td>2 (1.4)</td>
<td>5.7 (1.2–54.0)</td>
</tr>
<tr>
<td>MRSA non-ST398</td>
<td>3 (2.1)</td>
<td>0 (0)</td>
<td></td>
</tr>
<tr>
<td>Total MRSA</td>
<td>14 (9.6)</td>
<td>2 (1.4)</td>
<td>7.5 (1.7–68.6)</td>
</tr>
<tr>
<td>Veterinarians working with livestock</td>
<td>(n = 105)</td>
<td>(n = 97)</td>
<td></td>
</tr>
<tr>
<td>MRSA ST398</td>
<td>11 (10.5)</td>
<td>2 (2.1)</td>
<td>5.6 (1.2–52.6)</td>
</tr>
<tr>
<td>MRSA non-ST398</td>
<td>2 (1.9)</td>
<td>0 (0)</td>
<td></td>
</tr>
<tr>
<td>Total MRSA</td>
<td>13 (8.9)</td>
<td>2 (2.1)</td>
<td>6.7 (1.5–62.4)</td>
</tr>
<tr>
<td>Veterinarians not working with livestock</td>
<td>(n = 41)</td>
<td>(n = 46)</td>
<td></td>
</tr>
<tr>
<td>MRSA ST398</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td></td>
</tr>
<tr>
<td>MRSA non-ST398</td>
<td>1 (2.4)</td>
<td>0 (0)</td>
<td></td>
</tr>
<tr>
<td>Total MRSA</td>
<td>1 (2.4)</td>
<td>0 (0)</td>
<td></td>
</tr>
</tbody>
</table>

OR, Odds ratio; CI, confidence interval.

* Significant exposure variables are shown in bold (based on a P value of ≤0.05).
the last month before sampling and skin disorders could be associated with LA-MRSA carriage. The univariable analysis could not demonstrate that hygiene and protective measures (e.g. hand washing, use of gloves and mask) had a protective effect due to incomplete data for this variable. Statistically, working in a small animal veterinary clinic was protective. In the multivariable analysis, a negative association was observed between LA-MRSA acquisition and working in a small animal veterinary clinic (OR 0.15, 95% CI 0–1.0, \( P = 0.05 \)); however, the limited sample size does not allow us to clearly show a direct association. The strongest significant direct association found was between LA-MRSA acquisition and working with live pigs (OR 12.1, 95% CI 1.6–548.5, \( P = 0.01 \)).

**DISCUSSION**

The MRSA and LA-MRSA carriage rates were significantly higher for Belgian veterinarians (9.6% and 7.5%, respectively) than for Danish veterinarians (1.4%), with LA-MRSA accounting for 78.6% and 100% of the cases, respectively. The difference in carriage rate may be explained by geographical variations in MRSA–animal host associations and animal population structures. Moreover, considering the MRSA status of the Belgium food production system, it seems that large differences exist between both countries. In 2008, it was reported that the proportion of positive pig farms was tenfold higher for Belgium than for Denmark (40.0% and 35.9% vs. 0.0% and 3.5% for breeding and production holdings, respectively) [3]. Comparative data from Belgium and Denmark on MRSA prevalence in veal farms are currently lacking; however, in a recent study it has been shown that LA-MRSA was highly prevalent in Belgium veal calf farms (64%) [23]. Therefore, it seems that Belgium has a large potential LA-MRSA reservoir in its production system that may in turn represent a threat to people in contact with it. The MRSA carriage rates observed in this study are in accord with previous findings of veterinarians from other European countries, with carriage rates ranging from 3% to 17.9% depending on the country and the type of practice [24–26]. Moreover, the observed carriage rates were about tenfold higher than those in the general population (Belgium, 0.5%; Denmark, 0.2%) (R. Skov, unpublished data) and were comparable to the levels found in Belgian nursing-home residents (19.5%) [29]. Thus, from a healthcare point of view, this segment of the population can be considered as a group of concern and measures to protect them and the healthcare system should be considered.
In the univariable analysis, the predominant factors affecting nasal carriage of LA-MRSA among Belgian veterinarians were exposure to pigs and, to a lesser extent, cattle. This is in accord with previous findings [13, 14, 30]. However, in the multivariable analysis, exposure to cattle was not retained as a risk factor and only exposure to live pigs was positively associated. Beside this, our results suggest that working with small animals in a veterinary clinic is negatively associated with LA-MRSA acquisition. This observation is in agreement with a recent Belgium study in which LA-MRSA ST398 could not be detected in dogs [31] and suggests that the small animal population does not seem to constitute a significant LA-MRSA reservoir. However, those findings should be viewed with caution and require further studies including larger populations. In our study the sample size was limited to a small number and the proportion of participants in the category ‘working with small animals in a veterinary clinic’ was about twofold lower compared to the category of veterinarians working with livestock.

The LA-MRSA isolates identified in this study shared characteristics with the main genotypes reported in Europe [8, 24–26, 30]. The majority of LA-MRSA isolates (53.8%) showed a multidrug-resistant phenotype, which supports the hypothesis that this clone has adapted to the antimicrobial pressure encountered in intensive animal husbandry [32, 33]. In particular, the fact that genes resistant to a large variety of antibiotics are present in zoonotic bacteria is of great concern for public health since it may lead to limit and failure of human therapy when treating infection.

The non-ST398 MRSA strains found in this study belong to genotypes commonly associated with hospital- or community-acquired infections in humans, which have recently been reported in pigs and poultry. However, the limited number of isolates does not allow us to suggest any association.

This study has a limitation. Since veterinary practices appear to differ within and between countries, comparison of MRSA prevalence by animals species could not be accurately evaluated and MRSA prevalence was determined by category, livestock contact vs. no livestock contact, therefore prevalence has to be interpreted with caution.

In conclusion, veterinarians are at high risk of acquiring LA-MRSA through exposure to pigs, and particularly to live pigs, rather than other animal species. The fact that carriage was not dependent on a recent exposure to pigs may suggest that veterinarians could become chronic carriers and therefore they could be implicated in the spread of LA-MRSA between farms. Long-term epidemiological studies are currently underway to investigate this hypothesis.

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DECLARATION OF INTEREST

None.

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