Prenatal stress, immunity and neonatal health in farm animal species*

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The high pre-weaning mortality in farm animal species and poor welfare conditions of reproductive females question modern industrial farming acceptability. A growing body of literature has been produced recently, investigating the impact of maternal stress during gestation on maternal and offspring physiology and behavior in farm animals. Until now, the possible impact of prenatal stress on neonatal health, growth and survival could not be consistently demonstrated, probably because experimental studies use small numbers of animals and thus do not allow accurate estimations. However, the data from literature synthesized in the present review show that in ungulates, maternal stress can sometimes alter important maternal parameters of neonatal survival such as colostrum production (ruminants) and maternal care to the newborn (pigs). Furthermore, maternal stress during gestation can affect maternal immune system and impair her health, which can have an impact on the transfer of pathogens from the mother to her fetus or neonate. Finally, prenatal stress can decrease the ability of the neonate to absorb colostral immunoglobulins, and alter its inflammatory response and lymphocyte functions during the first few weeks of life. Cortisol and reproductive hormones in the case of colostrogenesis are pointed out as possible hormonal mediators. Field data and epidemiological studies are needed to quantify the role of maternal welfare problems in neonatal health and survival.

**Keywords:** maternal stress, neonate, health, mortality, immunity

**Implications**

Pre-weaning mortality rates in farm animals are high and variable between farms: 10% to 25% in lambs, 7% to 50% in kids, 0% to 50% in calves, 0% to 35% in foals, 5% to 35% in pigs (Mellor and Stafford, 2004), and important efforts are carried out by farmers to reduce these economically and ethically questionable losses. Concurrently, gestating females are often exposed to stressors or poor housing conditions that also question modern industrial farming acceptability. The present review shows that stress of pregnant females may impair neonatal health and survival in pigs and ruminants through its impact on maternal behavior as well as on passive, innate and acquired immunity of the offspring.

**Introduction**

Research has clarified much about the causes of neonatal mortality in farm animals and has provided practical means for decreasing it. The major mortality and health problems are due to hypothermia, hypoxia or starvation. Maternal undernutrition, mismothering, injury and infection represent a significant part of morbidity and mortality after the perinatal phase (Mellor and Stafford, 2004; Baxter et al., 2012; Walker et al., 2012). Management practices dedicated to improve neonatal survival and health are mainly focusing on the young animal itself, on its direct environment, or on the maternal qualities of the lactating mother. Less investment is devoted to the prenatal environment of the young, particularly the general health and welfare status of the pregnant mothers.

Besides genetic factors, environmental events acting prenatally on the developing fetus are important determinants for disorders later in life (Barker et al., 1993; Fowden et al., 2006). The concept that poor maternal nutrition during pregnancy can have long-term consequences on health and well-being of the offspring is broadly accepted (Reynolds and Caton, 2012). For example, gestating cattle submitted to drought give birth to calves with severe health and growth abnormalities (White et al., 2010), and nutritional strategies for pregnant females have been developed to favor fetal growth and neonatal survival (Le Floch et al., 2012; Reynolds and Caton, 2012). In this review, major physiological stressors...
such as malnutrition or materno-fetal infections will not be considered because they have obvious effects for the offspring that have been described elsewhere, and efforts are made in modern farm systems to suppress these two sources of problems. In this review, the term ‘stressor’ refers to less marked, non-nutritional and non-pathogenic environmental challenges that nevertheless impair maternal welfare and can induce a physiological stress response. These stressors are often related to housing or management practices. The effects of such stressors, occurring during gestation, on pre-weaning mortality and morbidity and the possible physiological mechanisms will be presented.

Maternal stress during gestation and prenatal stress in the offspring: relevance in farm animal species

Sources of maternal stress
Reproductive females in modern husbandry are often exposed to stressors during gestation. For example, in intensive pig husbandry, pregnant females have to face transportation and moving to a new environment, hunger because of food limitation, leg disorders, fear from human or social stress because of grouping with unfamiliar conspecifics and competition for the access to feed (Hemsworth, 2003; D’Eath et al., 2009; Spoolder et al., 2009). In cattle, climatic stress, handling, transportation, fear from humans and social stress are also common stressors (Hemsworth, 2003; Arnott et al., 2012). Maternal stressors induce a stress response, raising maternal endogenous levels of cortisol and catecholamines (De Leeuw and Ekkel, 2004; Roussel et al., 2005; Cooke et al., 2012). Maternal stress during pregnancy influences the developing fetus, affecting its birth weight and physiology, including immune system maturation after birth (Meriot et al., 2008). In rodents, these effects are partly mediated by the fetal exposure to high concentrations of glucocorticoids from maternal origin (Barbazanges et al., 1996). Indeed, the access of glucocorticoids to the fetal compartment is regulated by the placental enzyme 11β-hydroxysteroid dehydrogenase type 2 (11β-HSD2). Although a proportion of the cortisol produced by the mother during the stress response is inactivated by this enzyme, cortisol levels are still elevated in the fetus. In farm animals, this has been demonstrated in pig (Klemcke et al., 1995) and sheep (Hennessy et al., 1982). Trophic factors modulating placental physiology might also be involved. For example, stress stimulates adrenaline release and, high physiological levels of adrenaline reduce uterine artery blood flow in sheep, altering materno-fetal glucose exchanges (Jones and Gu, 1986).

Rodent data on prenatal stress cannot be simply transposed to farm animals because of major species differences. Species that give birth to non-premature offspring (rats, rabbits and mice) have a large proportion of immune development during late gestation and the postnatal period (Holladay and Smialowicz, 2000; Holsapple et al., 2003). By contrast, animals that give birth to precocious offspring (sheep, pig, cattle and primates) display development of the immune system predominantly in utero (Trebitsch et al., 1996; Sinkora et al., 2005). Furthermore, primates and rodents have a hemomonochorial and hemochorial placenta, whereas pigs, sheep and goats have an epitheliochorial placenta, which affects the route of acquisition of maternal immunity. Passive immunity is obtained through the placenta in rodents and primates, and via colostrum in ungulates. Thus, specific studies are needed to investigate whether prenatal stress can influence neonatal survival and the immune function in ungulate offspring.

Models of prenatal stress in farm animal species
Most of the studies investigating the influence of prenatal stress on factors influencing neonatal survival and immunity in farm animals focused on pigs, using behavioral stressors such as nose sling restraint that involves pain (Tuchscherer et al., 2002), rough treatment (Lay et al., 2011) or social stress (Jarvis et al., 2006; Couret et al., 2009b). Experiments were also conducted using artificial activation of the maternal corticotrope axis with adrenocorticotropic hormone (ACTH) injections (Hausmann et al., 2000; Otten et al., 2007) or mimicking corticotrope axis activation by providing pregnant sows with oral cortisol (Kranendonk et al., 2005). However, studies using synthetic glucocorticoids must be considered cautiously because these exogenous glucocorticoids are metabolized by the placenta less efficiently, and thus have more severe consequences on fetal development than natural glucocorticoids (Singh et al., 2012). In ruminants, studies using stressors such as transport also provide data concerning mortality, or birth weight of the offspring (Lay et al., 1997a; Roussel et al., 2005; Cooke et al., 2012).

Heat and cold stress are not behavioral stressors. However, because they both activate the corticotrope axis (Messias de Braganca et al., 1998; Webster et al., 2008) and are relevant challenges for animals kept outdoors as well as indoors, the consequences of maternal thermal stress during gestation will be also considered in this review. Studies on pigs also compared the consequences for the offspring of contrasted housing conditions of sows during their pregnancy, most often comparing individual stalls and collective pens of variable size. The welfare benefit of group housing is still in debate. Compared with individual stalls, group housing increases circadian cortisol secretion (Melchior et al., 2012), but not in response to an ACTH challenge (Broom et al., 1995; Boyle et al., 2002). Group housing decreases oral stereotypes and aggression (Broom et al., 1995); however, the impact on body lesions is uncertain and probably strongly
depends on the interaction with the presence of foraging material and the housing design (Boyle et al., 2002; Harris et al., 2006; Hulbert and McGlone, 2006). Although these studies do not provide contrasted groups in terms of maternal stress, they are included in the review because of their practical relevance for pig husbandry.

Effect of maternal stress during gestation on morbidity and mortality of the offspring

Pre-weaning mortality
Maternal stress has a negative impact on reproductive performances in rodents, such as a reduced litter size (Götz et al., 2008) or an increased mortality of neonates (Pollard, 1984; Lordi et al., 2000), but this effect was not consistently reported (Llorente et al., 2002; Kapoor and Matthews, 2005; Götz et al., 2008). In farm animals, stressors occurring during early gestation might lead to abortions or, in polytocous species such as the pig, to partial fetal losses without abortion (Von Borell et al., 2007). However, most of the time, the development of the fetuses continues until term (Von Borell et al., 2007). A majority of studies, mainly in porcine species, failed to show any link between prenatal stress and mortality during the neonatal period (Table 1). Only one study demonstrated a clear effect of prenatal stress on mortality. In this study, repeated nose sling restraint during the last third of gestation more than doubled the mortality of the piglets during the sucking period (Tuchscherer et al., 2002).

Most of the time individual penning of sows led to a pre-weaning mortality similar to group housing (Broom et al., 1995; Bates et al., 2003; Harris et al., 2006; Hulbert and McGlone, 2006), except for some studies where a lower (Cronin et al., 1996) or a higher (Jarvis et al., 2006) mortality were found in litters from individually housed sows. Providing straw bedding to group-housed sows (Von Borell et al., 1992; Spoolder et al., 1996) or combining group penning with provision of foraging materials (wood shavings) during pregnancy (Von Borell et al., 1992; Spoolder et al., 1996) did not influence pre-weaning mortality.

Pre-weaning morbidity and markers of inflammation
Human epidemiological data show that maternal stress during pregnancy may be a risk factor for congenital malformations, and respiratory, digestive, skin and genito-urinary diseases (Tegethoff et al., 2011). Few data are available on farm mammals. In pigs, treatment of the mother with ACTH during the last third of gestation did not affect the frequency of diseased piglets during the lactation period (Otten et al., 2007). However, a strong behavioral stressor involving pain applied at the same gestational period increased the number of diseased piglets per litter until weaning (Tuchscherer et al., 2002). Studies that did not measure the occurrence of disease events directly but assessed blood levels of health markers, such as acute phase proteins and inflammatory cytokines, failed to find a clear effect of prenatal stress. Indeed, repeated social disturbance of the mothers during late gestation did not affect the levels of haptoglobin in piglets around weaning (Cour et al., 2009a). Nose sling restraint stress during late gestation decreased blood levels of tumor necrosis factor-α (TNF-α) and interleukin-6 (IL-6), but not of C-reactive protein, serum amyloid A (SAA) and haptoglobin in 2-month-old pigs (Collier et al., 2011). When comparing the piglets born from gilts housed in stalls or in groups, Sorrells et al. (2006) did not find significant differences in TNF-α, haptoglobin and alpha-1-acid glycoprotein levels during the 1st month of life.

Table 1 Impact on neonatal survival of maternal stressors or pharmacological activation of maternal corticotrope axis in farm animals

<table>
<thead>
<tr>
<th>Species</th>
<th>Treatment</th>
<th>Gestational period</th>
<th>Pre-weaning mortality</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pig</td>
<td>ACTH</td>
<td>0.36 to 0.73</td>
<td>=</td>
<td>Haussmann et al. (2000)</td>
</tr>
<tr>
<td>Pig</td>
<td>ACTH</td>
<td>0.43 to 0.65</td>
<td>=</td>
<td>Otten et al. (2007)</td>
</tr>
<tr>
<td>Pig</td>
<td>ACTH</td>
<td>0.74 to 0.93</td>
<td>=</td>
<td>Otten et al. (2007)</td>
</tr>
<tr>
<td>Pig</td>
<td>Hydrocortisone</td>
<td>0.18 to 0.43</td>
<td>&gt; (when the 3 treatment periods were pooled)</td>
<td>Kranendonk et al. (2006)</td>
</tr>
<tr>
<td>Pig</td>
<td>Social stress</td>
<td>0.21 to 0.41</td>
<td>=</td>
<td>Cour et al. (2009c)</td>
</tr>
<tr>
<td>Pig</td>
<td>Social stress</td>
<td>0.34 to 0.56</td>
<td>=</td>
<td>Jarvis et al. (2006)</td>
</tr>
<tr>
<td>Pig</td>
<td>Social stress</td>
<td>0.34 to 0.56</td>
<td>=</td>
<td>Ringgenberg et al. (2012)</td>
</tr>
<tr>
<td>Pig</td>
<td>Social stress</td>
<td>0.67 to 0.90</td>
<td>=</td>
<td>Jarvis et al. (2006)</td>
</tr>
<tr>
<td>Pig</td>
<td>Social stress</td>
<td>0.67 to 0.91</td>
<td>=</td>
<td>Cour et al. (2009a)</td>
</tr>
<tr>
<td>Pig</td>
<td>Social stress</td>
<td>0.69 to 0.90</td>
<td>=</td>
<td>Cour et al. (2009c)</td>
</tr>
<tr>
<td>Pig</td>
<td>Snaring</td>
<td>0.73 to 0.97</td>
<td>&gt;</td>
<td>Tuchscherer et al. (2002)</td>
</tr>
<tr>
<td>Pig</td>
<td>Cold</td>
<td>0.90 to 1.0</td>
<td>=</td>
<td>Bate and Hacker (1985b)</td>
</tr>
<tr>
<td>Pig</td>
<td>Fear of humans</td>
<td>0 to 1</td>
<td>=</td>
<td>Hemsworth et al. (1999)</td>
</tr>
<tr>
<td>Cattle</td>
<td>Aggressive temperament</td>
<td>0 to 1</td>
<td>=</td>
<td>Cooke et al. (2012)</td>
</tr>
</tbody>
</table>

The gestational period when the treatment was applied is expressed in percentage of the total gestation length (e.g. 0 to 0.5 means ‘from conception to mid-gestation’).

Studies comparing different housing systems during the gestation of sows are not included in the table because they did not provide contrasted groups in terms of maternal stress.

2018
To conclude, data showing an effect of prenatal stress on mortality and morbidity are scarce. Furthermore, the conclusions of most of them are based on small numbers of experimental animals that did not allow an accurate evaluation of morbidity and mortality rates. Epidemiological data are lacking. Nevertheless, data from the literature suggest that prenatal stress might have an impact on important factors involved in the preservation of neonatal health. These factors can be either related to maternal biology or to true prenatal stress effects, that is, long-term alterations in fetal ontogeny (Figure 1). These two aspects are described below.

Effect of maternal stress during gestation on maternal qualities

Colostrum production

Consumption of colostrum provides energy for thermoregulation and passive immunity. Insufficient colostrum intake is one of the major causes of mortality until weaning in farm mammals (Tyler et al., 1998; Devillers et al., 2011). To the best of our knowledge, the impact of psychological stress on colostrum yield and gross composition has not been investigated. In pigs, psychological stress during the last third of pregnancy had no effect on IgG contents in colostrum (Tuchscherer et al., 2002; Couret et al., 2009a).

In pigs, heat stress during the last week of pregnancy decreased (Machado-Neto et al., 1987), whereas cold stress during the last 2 days increased (Bate and Hacker, 1985b) the circulating IgG levels in neonates. The possible causes for these variations are either an impaired ability of the neonate to suckle colostrum and absorb IgG (see below), or a decrease in the quantity and quality of the colostrum produced by the mother. Colostrum quality might be affected as heat stress tended to decrease colostrum IgG (Machado-Neto et al., 1987). The physiological mechanisms responsible for this alteration have not been investigated.

In cattle, heifers born from cows exposed to natural summer heat stress during the last 45 days of pregnancy had lower serum concentrations of IgG after colostrum consumption than heifers born from mothers that received a cooling treatment (Tao et al., 2012). Several studies investigated whether alteration in the transfer of maternal immunity from heat-stressed cows was related to a deterioration of the composition and/or the quantity of colostrum produced by the mothers. Nardone et al. (1997) found that heat stress during the last 3 weeks of gestation was associated with decreased energy and protein content in colostrum, including a reduction of colostral IgG and IgA. Adin et al. (2009) found that exposure to heat stress during the dry period reduced both the quantity of first colostrum and its IgG content. However, cows exposed to natural summer heat in Florida had higher colostral IgG concentrations than cows sampled during other months of the year (Shearer et al., 1992). Furthermore, cows exposed to a moderate heat stress owing to the hot Mediterranean summer had similar colostrum IgG concentrations as those delivering during spring (Lacetera et al., 2002). In these last two studies, the effect of heat was confounded with a more general seasonal effect.

In pigs and cattle, colostrum yield and composition are under hormonal control, with prolactin and steroids (progesterone, estradiol and glucocorticoids) playing a key role (Barrington et al., 2001; Farmer and Quesnel, 2009). Alterations in prolactin and progesterone concentrations during the prepartum period were associated with a decreased production of colostrum by primiparous sows (Foisnet et al., 2010). Moreover, prolactin was demonstrated to induce the cessation of IgG transfer into colostrum in cows, and glucocorticoids are also likely to be involved (Barrington et al., 2001). As heat stress increases prolactin levels during the last few weeks of gestation of cows (do Amaral et al., 2011), prolactin has been proposed as a candidate for the decreased colostrogenesis in heat-stressed cows (Tao et al., 2012). By contrast, in sows, repeated injections of ACTH during the third trimester of gestation had minor influence on maternal progesterone levels (Brüssow et al., 2005), and heat stress starting at 84 days of gestation did not affect maternal prolactin levels around parturition (Messias de Braganca et al., 1998). These observations fit with the negligible effect of maternal stress on colostrum production in sows. Further studies are required to determine
whether increased cortisol in stressed females played a role in terminating IgG transfer into colostrum.

**Gut flora of the neonate**

Maternal stress during late pregnancy can also alter the establishment of the newborn’s commensal intestinal flora. In Rhesus monkeys, maternal stress reduced the numbers of lactobacilli and bifidobacteria in the gut during the first 24 weeks of life of the offspring (Bailey et al., 2004). In rats, maternal cortisone injection reduced the concentrations of total and gram-negative bacteria in pups (Schiffrin et al., 1993). These alterations in the flora of the offspring are suspected to result from stress-induced changes in maternal gut flora or milk. Comparable studies in farm animals are lacking. Such changes could be particularly important because these commensal bacteria are involved in protection against diarrheal illness in piglets and calves (Abe et al., 1995).

**Maternal care**

Maternal care is a major element for perinatal survival in mammals. In pigs, two social reorganizations during mid or late pregnancy did not alter maternal behavior around parturition and until 6 h post parturition (Jarvis et al., 2006). However, more frequent social perturbations during mid-gestation increased the time lying ventrally in early lactation, thus restricting access to the udder and indicating a lower motivation of sows to nurse (Ringgenberg et al., 2012). In sheep, repeated transport associated with isolation stress during the last third of pregnancy reduced the dam’s motivation to stay in contact with her lamb when a human being was approaching (Roussel et al., 2006). The authors hypothesized that this might not result from a weaker mother–young bond but from increased fear of human, because of the aversive treatments that ewes experienced in the contact of humans during pregnancy. Accordingly, in another study, ewes subjected to daily aversive handling during the last 5 weeks of gestation displayed increased grooming of their lamb during the first 24 hours after parturition, but fear of humans disrupted their ability to follow their lambs closely when carried away by a human (Hild et al., 2011).

Prenatal stress might even influence maternal behavior of female offspring. Indeed, group-housed sows socially stressed during mid or late pregnancy gave birth to female offspring that were more responsive to newborn piglets that approached their head and tended to bite them more than control females (Jarvis et al., 2006).

**Effect of maternal stress on subclinical maternal health deterioration and consequences on maternal qualities**

**Immune status of the mothers and transmission of pathogens**

If stress during gestation weakens maternal immunity and worsens maternal health status, it might also affect the immune and microbial inheritance that the mother transmits to its progeny. In humans, maternal stress during gestation was associated with increased markers of inflammation in maternal plasma and increased responsiveness of maternal immune cells to inflammatory stimulation in vitro, supporting the view that prenatal stress alters maternal immune function (Coussons-Read et al., 2007). In gestating sows, repeated social stress increased salivary cortisol but did not alter either blood cell counts and lymphocyte proliferation, or the antibody response to an immunization (Couret et al., 2009b).

Regarding housing environment, most of the studies comparing individual stall v. group housing combined or not with provision of foraging materials failed to detect any impairment in the maternal immune markers investigated (Von Borell et al., 1992; Hulbert and McGlone, 2006; Sorrells et al., 2007). The lack of difference might be owing to the fact that the space allocated per animal in the small groups was not much bigger than in individual stalls (not more than 2.3 m² per sow). In opposite, sows housed in large groups on deep litter or with a lot of space per animal (9 m²) displayed improved lymphocyte counts during lactation (Broom et al., 1995) or neutrophil/lymphocyte ratio at the end of gestation (Karlen et al., 2007) than sows housed in individual stalls.

Individual housing is expected to limit the spread of pathogens that disseminate via direct contacts between animals or via the feces. Contrarily, epidemiological data suggest that individual housing during pregnancy may increase the transmission of pathogens from the mother to the fetus or neonate. Indeed, compared with group-housed sows, animals housed in individual pens during pregnancy were found to be more at risk for post-weaning multisystemic wasting syndrome development in their offspring (Rose et al., 2003) and shedding of Campylobacter coli in their feces during lactation (Denis et al., 2011). However, in these studies, the reason for the difference between stall and group housing in pathogen transmission is not known, and investigation is needed to determine whether maternal stress is involved.

**Colostrum production**

The impact of subclinical maternal problems during gestation on colostrum production and quality has been poorly explored. In cows, pre-partum mastitis reduced the volume of colostrum and the total amount of IgG exported into colostrum, along with total protein and fat contents (Maunsell et al., 1998). In an artificial model of mild chronic inflammation, cows were treated orally with interferon-alpha (IFN-α) during the last 2 weeks of pregnancy (Trevisi et al., 2009). At the first milk sampling, 3 days postpartum, the IFN-α group showed less 2 protein and fat contents in milk than did the control group, which suggests that the immunoglobulin protein fraction might have been also decreased during the first 3 days.

**Effect of maternal stress during gestation on the biology of the offspring**

**Birth weight**

Birth weight is a critical factor for neonatal survival and disease incidence in farm species (Tuchscherer et al., 2000;
Prenatal stress and neonatal health

Table 2 Impact on birth weight of maternal stressors or pharmacological activation of maternal corticotrope axis in farm animals

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Gestational period</th>
<th>Birth weight</th>
<th>Growth until weaning</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pig Cold stress</td>
<td>0.91 to 1.0</td>
<td>=</td>
<td>=</td>
<td>Bate and Hacker (1985a)</td>
</tr>
<tr>
<td>Pig ACTH</td>
<td>0.91 to 1.0</td>
<td>=</td>
<td>=</td>
<td>Bate and Hacker (1985b)</td>
</tr>
<tr>
<td>Pig ACTH</td>
<td>0.43 to 0.65</td>
<td>=</td>
<td>=</td>
<td>Otten et al. (2007)</td>
</tr>
<tr>
<td>Pig ACTH</td>
<td>0.74 to 0.93</td>
<td>=</td>
<td>=</td>
<td>Otten et al. (2007)</td>
</tr>
<tr>
<td>Pig ACTH</td>
<td>0.36 to 0.73</td>
<td>&lt; (tendency)</td>
<td>=</td>
<td>Haussmann et al. (2000)</td>
</tr>
<tr>
<td>Pig ACTH</td>
<td>0.37 to 0.66</td>
<td>=</td>
<td>=</td>
<td>Lay et al. (2008)</td>
</tr>
<tr>
<td>Pig Hydrocortisone</td>
<td>0.18 to 0.43</td>
<td>&lt;</td>
<td>=</td>
<td>Kranendonk et al. (2006)</td>
</tr>
<tr>
<td>Pig Hydrocortisone</td>
<td>0.44 to 0.70</td>
<td>=</td>
<td>=</td>
<td>Kranendonk et al. (2006)</td>
</tr>
<tr>
<td>Pig Hydrocortisone</td>
<td>0.70 to 0.95</td>
<td>&lt;</td>
<td>=</td>
<td>Kranendonk et al. (2006)</td>
</tr>
<tr>
<td>Pig Social stress</td>
<td>0.21 to 0.41</td>
<td>=</td>
<td>=</td>
<td>Cour et al. (2009c)</td>
</tr>
<tr>
<td>Pig Social stress</td>
<td>0.34 to 0.56</td>
<td>=</td>
<td>=</td>
<td>Ringgenberg et al. (2012)</td>
</tr>
<tr>
<td>Pig Social stress</td>
<td>0.34 to 0.56</td>
<td>=</td>
<td>=</td>
<td>Jarvis et al. (2006)</td>
</tr>
<tr>
<td>Pig Social stress</td>
<td>0.67 to 0.91</td>
<td>=</td>
<td>=</td>
<td>Cour et al. (2009a)</td>
</tr>
<tr>
<td>Pig Social stress</td>
<td>0.69 to 0.90</td>
<td>=</td>
<td>=</td>
<td>Cour et al. (2009c)</td>
</tr>
<tr>
<td>Pig Social stress</td>
<td>0.68 to 0.90</td>
<td>=</td>
<td>=</td>
<td>Jarvis et al. (2006)</td>
</tr>
<tr>
<td>Pig Rough handling</td>
<td>0.37 to 0.66</td>
<td>=</td>
<td>=</td>
<td>Lay et al. (2008)</td>
</tr>
<tr>
<td>Pig Snaring</td>
<td>0.73 to 0.97</td>
<td>=</td>
<td>=</td>
<td>Tuchscherer et al. (2002)</td>
</tr>
<tr>
<td>Pig Heat stress</td>
<td>0.87 to 1</td>
<td>=</td>
<td>=</td>
<td>Machado-Neto et al. (1987)</td>
</tr>
<tr>
<td>Pig Heat stress</td>
<td>0.88 to 1</td>
<td>=</td>
<td>=</td>
<td>Prunier et al. (1997)</td>
</tr>
<tr>
<td>Pig Heat stress</td>
<td>0.73 to 1</td>
<td>=</td>
<td>=</td>
<td>Messias de Braganca et al. (1998)</td>
</tr>
<tr>
<td>Cattle ACTH</td>
<td>0.21 to 0.49</td>
<td>=</td>
<td>=</td>
<td>Lay et al. (1997b)</td>
</tr>
<tr>
<td>Cattle Heat stress</td>
<td>0.31 to 1.0</td>
<td>=</td>
<td>=</td>
<td>Shell et al. (1995)</td>
</tr>
<tr>
<td>Cattle Heat stress</td>
<td>0.61 to 1.0</td>
<td>=</td>
<td>=</td>
<td>Collier et al. (1982)</td>
</tr>
<tr>
<td>Cattle Heat stress</td>
<td>0.80 to 1.0</td>
<td>=</td>
<td>=</td>
<td>Adin et al. (2009)</td>
</tr>
<tr>
<td>Cattle Heat stress</td>
<td>0.84 to 1.0</td>
<td>=</td>
<td>=</td>
<td>Tao et al. (2012)</td>
</tr>
<tr>
<td>Cattle Cold stress</td>
<td>0.0 to 1.0</td>
<td>=</td>
<td>=</td>
<td>Andreoli et al. (1988)</td>
</tr>
<tr>
<td>Cattle Transport</td>
<td>0.21 to 0.49</td>
<td>=</td>
<td>=</td>
<td>Lay et al. (1997b)</td>
</tr>
<tr>
<td>Goat ACTH</td>
<td>0.74 to 1.0</td>
<td>=</td>
<td>=</td>
<td>Roussel et al. (2005)</td>
</tr>
<tr>
<td>Goat Transport</td>
<td>0.74 to 1.0</td>
<td>=</td>
<td>=</td>
<td>Roussel et al. (2005)</td>
</tr>
<tr>
<td>Goat Rough handling</td>
<td>0.76 to 1.0</td>
<td>=</td>
<td>=</td>
<td>Hild et al. (2011)</td>
</tr>
</tbody>
</table>

The gestational period when the treatment was applied is expressed in percentage of the total gestation length (e.g. 0 to 0.5 means ‘from conception to mid-gestation’).

Bleul, 2011). Although human studies show that maternal stress exposure during mid-pregnancy is a risk factor for pre-term birth and small for gestational age children (Class et al., 2011), results from studies in farm animals are less clear.

In pigs, maternal treatment with ACTH or glucocorticoid reduced (Haussmann et al., 2000; Kranendonk et al., 2006), or increased the birth weight of the piglets (Otten et al., 2007). A variety of maternal psychological stressors have been studied (Table 2). None of them affected piglet birth weight except in one study where individual housing during gestation decreased litter weight (Machado-Neto et al., 1987). Heat stress during the last 10 or 30 days of pregnancy had no impact on individual and litter birth weight (McCrabb et al., 1993). Heat stress during the last third of gestation decreased the birth weight of calves (Lay et al., 1997a) but not of goat kids (Roussel et al., 2005). Behavioral stressors were mainly reported to have no impact or to increase birth weight (Table 2). Heat stress during the last third of pregnancy had no impact (Shell et al., 1995) or decreased birth weight in calves, lambs and goat kids (Collier et al., 1982; McCrabb et al., 1993; Mellado et al., 2000; Tao et al., 2012). Moreover, it has been shown in ewes that heat stress during mid-pregnancy had a delayed effect on fetal growth later during pregnancy, probably because of a long-lasting negative effect on placental growth (McCrabb et al., 1993). Exposure of dams to winter weather during the last third of gestation decreased the birth weight of calves relatively to those of dams housed at thermoneutrality (Andreoli et al., 1988).

Passive immunity
Impairment of the transfer of maternal immunity to the neonate has been described above. Besides effects on colostrum composition and quantity, late gestational stress can also decrease the ability of the offspring to acquire maternal immunity from colostrum. For example, decreased IgG contents were observed in piglets whose mothers were exposed to stress during late pregnancy, whereas IgG levels in the colostrum were not altered (Tuchscherer et al., 2002). There are no data investigating whether prenatal stress affects the amount of colostrum that neonates ingest, but indirect measurements suggest that this is unlikely the case. In pigs, treatment of the mother with ACTH during gestation did not affect neonatal vitality and latency of the...
first colostrum intake (Otten et al., 2007). Furthermore, the postnatal growth does not seem to be significantly deteriorated, which suggests that prenatally stressed neonates have at least normal milk consumption (Table 2).

Prenatal stress might affect the absorption of colostral IgG through the gut. Indeed, calves born from heat-stressed cows presented lower plasma IgG levels than control calves after ingestion of colostrum from their respective dams, despite the fact that calves were fed with similar amounts of colostrum, and that similar IgG levels were found in the colostrum of their mothers (Tao et al., 2012). The ability of neonatal gut to take up and transfer intact proteins from the epithelium into the circulation is a very time-specific process that develops close to term, and the signals inducing gut closure probably vary among species (Sangild, 2003). In pigs, glucocorticoids might be involved in that phenomenon (Bate and Hacker, 1985a and 1985b; Machado-Neto et al., 1987; Bate et al., 1991). Fetal glucocorticoid levels increase during the last few days before parturition and then decrease continuously after birth until reaching levels comparable to adult levels. The prenatal increase in cortisol has been proposed to be the signal for enhanced gut permeability for IgG, whereas the decrease after birth would favor gut maturation and closure (Bate et al., 1991). Therefore, a transient increase in maternal glucocorticoids because of maternal stress in late gestation may accelerate gut maturation, leading to impaired immunoglobulin acquisition after birth.

Innate immune function of the offspring

The inflammatory response has been investigated in pigs using in vivo lipopolysaccharide (LPS) challenges, but the results are contradictory. The fever response to LPS was increased in female pigs born from sows treated with glucocorticoids during pregnancy (de Groot et al., 2007). Collier et al. (2011) found that restraint stress during the last quarter of gestation altered pro-inflammatory cytokine and stress hormone response of 1-month-old piglets to an LPS injection, and that these changes were influenced by gender. Plasma IL-6 and SAA increased more in prenatally stressed offspring of both genders, but TNF-α and cortisol responses were decreased in prenatally stressed male offspring only (Collier et al., 2011). Conversely, regardless of the time of application (mid or late gestation), repeated maternal social stress did not affect cortisol and IL-6 plasma responses to an LPS challenge of male piglets (Couret et al., 2009c). However, when explored in vitro, immune cell responsiveness to LPS was decreased by late prenatal stress in both genders. Blood cell cultures stimulated with LPS tended to release less IL-6 in 2-month-old prenatally stressed piglets but not in younger ones (Couret et al., 2009a), and to proliferate less at 1 and 35 days of age (Tuchscherer et al., 2002). However, the ability of blood mononcytic cells from growing pigs to produce nitric oxide in response to LPS was not altered by rough handling or ACTH treatment occurring during mid-gestation (Lay et al., 2011).

Skin wound healing is closely related to inflammatory processes. It has been found accelerated in piglets born from ACTH-treated mothers (Haussmann et al., 2000; Lay et al., 2011). Apart from the inflammatory response, the non-specific immune response was nearly not explored in prenatally stressed farm animals. Only one study investigated natural killer cell cytotoxic responses in piglets and found that prenatal stress had no impact (Tuchscherer et al., 2002).

Acquired immune function of the offspring

Piglets stressed during late pregnancy were reported to decrease the thymus size at birth (Tuchscherer et al., 2002), blood lymphocyte and granulocyte numbers (Couret et al., 2009a), and blood CD4/CD8 ratio (Couret et al., 2009a) in the neonatal period relatively to unstressed counterparts. The functional impact of prenatal stress on immune function was investigated by in vivo challenges, which did not reveal any effect of prenatal stress. For example, antigen-specific responses of piglets to an immunization against ovalbumin during the lactation period were tested by measuring the blood anti-ovalbumin IgG levels and the ovalbumin-stimulated proliferative response of lymphocytes (Couret et al., 2009c). Results revealed no significant effect of prenatal stress. Similarly, the antibody production in response to immunization against sheep red blood cells was not altered in piglets born from sows that were ACTH-treated or roughly handled during mid-gestation (Lay et al., 2011).

In vitro investigations revealed that prenatal stress in pigs could either increase (Couret et al., 2009b; Couret et al., 2009c) or decrease (Tuchscherer et al., 2002) blood lymphocyte proliferation after in vitro mitogen stimulation. These alterations were observable until 35 days (Tuchscherer et al., 2002) or even 2 months of age (Couret et al., 2009a). Heat stress during late gestation in cows also reduced blood lymphocyte proliferation until 56 days of age in their female offspring (Tao et al., 2012). This effect might be specific to the blood because the lymphocyte proliferation was not affected in other compartments such as the spleen or thymus (Couret et al., 2009c). Prenatal stress may also increase the sensitivity of lymphocytes to the inhibitory effect of glucocorticoids. Indeed, in vivo ACTH treatment induced more severe inhibition of blood lymphocyte proliferation in prenatally stressed piglets (Tuchscherer et al., 2002). However, this hypothesis was partly confirmed when exposing thymocytes of prenatally or control piglets to physiological doses of cortisol in vitro: spontaneous proliferation of thymocytes was more inhibited by cortisol in prenatally stressed piglets than in controls, but this difference disappeared when thymocytes were stimulated with a mitogen in the presence of cortisol (Couret et al., 2009c).

Regarding the possible existence of windows of vulnerability to prenatal stress, late maternal stress seems to have a more pronounced disrupting influence than early prenatal stress on the immune system of pigs. Indeed, application of prenatal stress (Couret et al., 2009c) or pharmacological activation of the corticotrope axis during the first third of gestation (Otten et al., 2007) had no impact, whereas the same treatments applied during late pregnancy did have an impact.
Sex-specific effects of prenatal stress

Interestingly, the effects of prenatal stress differ among male and female offspring. Sex-specific effects of prenatal stress were reported in pigs concerning the lymphocyte proliferative response, which was more affected in females (Couret et al., 2009c) and the inflammatory response to LPS, which was more altered in males (Collier et al., 2011). In sheep, sex-specific effects of prenatal stress were reported on the emotional reactivity of goat kids (Roussel et al., 2005). In primates and rodents, sex-specific effects were also reported on behavior and neuroendocrine function (Weinstock, 2007) as well as on immune function, which seemed to be more consistently affected in male offspring (Klein and Rager, 1995; Coe et al., 1996). These differences might be due to a higher placental transfer of maternal glucocorticoids in males, as observed in rodents (Montano et al., 1991). In pigs (Klemcke and Christenson, 1996), sheep (Braun et al., 2009) and humans (Stark et al., 2009), female placentae seem to have a greater ability to increase their $11\beta$-HSD2 activity or expression in response to physiological stressors than male placentaes.

Conclusion

The data from the present review show that in pigs and ruminants, maternal stress alters important parameters related to neonatal survival such as transfer of passive immunity from the mother to the newborn and immune defences of the neonate. Impaired maternal care could also be involved. However, no effects on neonatal health, growth and survival could be consistently demonstrated. Field data from epidemiological studies are needed. Even if the effects of maternal stress during gestation on neonatal survival are weak, stress of pregnant females in modern husbandry remains an ethical question because of the deteriorated welfare in the mothers, and altered emotional and stress reactivity in the offspring (Braastad, 1998).

References


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