

Proceedings of the Nutrition Society (2019), **78**, 34–44 © The Author 2018 First published online 13 September 2018

The Nutrition Society Scottish Section Meeting was held at the Royal College of Physicians and Surgeons, Glasgow on 26–27 March 2018

### Conference on 'Nutrient-nutrient interaction' Symposium 2: Nutrient interactions and their role in protection from chronic diseases

# Multiple nutritional factors and thyroid disease, with particular reference to autoimmune thyroid disease

#### Margaret P. Rayman

Department of Nutritional Sciences, Faculty of Health and Medical Sciences, University of Surrey, Guildford GU2 7XH, UK

Hashimoto's thyroiditis (HT) and Graves' disease (GD) are examples of autoimmune thyroid disease (AITD), the commonest autoimmune condition. Antibodies to thyroid peroxidase (TPO), the enzyme that catalyses thyroid-hormone production and antibodies to the receptor for the thyroid-stimulating hormone, are characteristic of HT and GD, respectively. It is presently accepted that genetic susceptibility, environmental factors, including nutritional factors and immune disorders contribute to the development of AITD. Aiming to investigate the effect of iodine, iron and selenium in the risk, pathogenesis and treatment of thyroid disease, PubMed and the Cochrane Library were searched for relevant publications to provide a narrative review. *Iodine*: chronic exposure to excess iodine intake induces autoimmune thyroiditis, partly because highly-iodinated thyroglobulin (Tg) is more immunogenic. The recent introduction of universal salt iodisation can have a similar, although transient, effect. Iron: iron deficiency impairs thyroid metabolism. TPO is a haem enzyme that becomes active only after binding haem. AITD patients are frequently iron-deficient since autoimmune gastritis, which reduces iron absorption and coeliac disease which causes iron loss, are frequent co-morbidities. In two-thirds of women with persistent symptoms of hypothyroidism despite appropriate levothyroxine therapy, restoration of serum ferritin above 100 µg/l ameliorated symptoms. Selenium: selenoproteins are essential to thyroid action. In particular, the glutathione peroxidases remove excessive hydrogen peroxide produced there for the iodination of Tg to form thyroid hormones. There is evidence from observational studies and randomised controlled trials that selenium, probably as selenoproteins, can reduce TPO-antibody concentration, hypothyroidism and postpartum thyroiditis. Appropriate status of iodine, iron and selenium is crucial to thyroid health.

Autoimmune thyroid disease: Autoimmune thyroiditis: Nutrition: Iodine: Iron: Selenium

The thyroid gland is the organ most commonly affected by autoimmune disease<sup>(1)</sup>. Lymphocytic infiltration of the thyroid is a frequent post-mortem finding in some 40 % of white females and 20 % of white males in the USA, with similar percentages of British white males and females being affected<sup>(2)</sup>. In black Americans and Japanese, the occurrence of lymphocytic thyroid infiltration was less than half that in Caucasians<sup>(2)</sup>. Autoimmune thyroid disease (AITD) was probably first

described in 1912 when a Japanese physician, Hakaru Hashimoto, reported a condition where the thyroid was infiltrated by lymphocytes resulting in the production of anti-thyroid antibodies<sup>(3)</sup>.

AITD, also known as autoimmune thyroiditis, has a multifactorial aetiology involving both genetic, environmental and nutritional factors<sup>(4)</sup>. It includes a spectrum of thyroid conditions ranging from hypothyroidism, most notably Hashimoto's thyroiditis (HT) at one end,

**Abbreviations:** AITD; autoimmune thyroid disease; DIO; deiodinases; GD; Graves' disease; GPX; glutathione peroxidases; HT; Hashimoto's thyroiditis; ID; iron deficiency; L-T4; levothyroxine; RCT; randomised controlled trial; T3; tri-iodothyronine; T4, thyroxine; Tg; thyroglobulin; TPO; thyroid peroxidase; TSH; thyroid-stimulating hormone.

Corresponding author: Professor Margaret P. Rayman, email m.rayman@surrey.ac.uk



to hyperthyroidism, most commonly Graves' disease (GD) at the other end<sup>(4)</sup>. In HT, the thyroid gland is gradually destroyed resulting in reduced production of thyroid hormones and triggering symptoms that include fatigue, weight gain, constipation, increased sensitivity to cold, dry skin, depression, muscle aches and reduced exercise tolerance<sup>(5)</sup>. HT affects more than 15% of females over 60 years and 2% of males<sup>(6)</sup>. It is defined by the presence of antibodies to thyroid peroxidase (TPO), the thyroid enzyme that oxidises iodide to iodine for thyroid hormone synthesis<sup>(4)</sup>. In addition, antibodies to thyroglobulin (Tg), the protein on which thyroid hormones are synthesised by iodination of its tyrosine residues, are frequently present<sup>(4)</sup>. In GD, the major autoantigen (thyroid-stimulating hormone (TSH)-antibody) is to the receptor for the TSH, which causes overproduction of thyroid hormones, resulting in symptoms such as irritability, rapid heartbeat, weight loss, poor tolerance of heat and orbitopathy)<sup>(4,7,8)</sup>. bulging eyes (Graves'

Nutritional factors that affect thyroid function include the micronutrients iodine, iron and selenium<sup>(9)</sup>; these will be discussed later. Though vitamin D has been postulated to affect thyroid function, the evidence is insufficient<sup>(9)</sup> to include it in the present review.

PubMed and the Cochrane Library were searched for publications up to March 2018 using the search terms 'autoimmune thyroiditis' OR 'autoimmune thyroid disease' in combination with 'iodine', 'selenium', 'iron' and 'nutrition OR diet'. Articles were filtered by the relevance of the title, abstract and finally the full text. Relevant conclusions or results were extracted from each article to provide a narrative review.

#### **Iodine**

Role of iodine in the thyroid

Iodine is a key constituent of the thyroid hormones, thyroxine (T4, pro-hormone) and tri-iodothyronine (T3, active hormone), as shown in Fig. 1, which also depicts the key players in thyroid hormone synthesis that takes place in the thyroid follicular cells<sup>(10)</sup>.

#### Iodine and autoimmune thyroid disease

The association between iodine intake and the presence of circulating thyroid antibodies is complex with iodine intake both below and above the recommended level being associated with an increase in circulating antibodies<sup>(11)</sup>. Circulating TPO-antibodies and Tg-antibodies are common both in populations with a stable high iodine intake and those with mild and moderate iodine deficiency (ID)<sup>(12)</sup>. Deficient iodine intake can lead to nodular goitre in which thyroid antigens are released from the abnormal gland, resulting in the presence of thyroid antibodies in the circulation<sup>(13)</sup>. However, excess iodine intake or a rise in intake following iodine fortification of an iodine-deficient population also gives an increased risk of thyroid autoimmunity, as attested by studies in many countries<sup>(14–24)</sup>. In China, for instance, 3 years after the introduction of

salt iodisation in 1996, the prevalence of AITD was  $0.5\,\%$  in an area of mildly deficient iodine intake,  $1.7\,\%$  in an area of more-than-adequate iodine intake and  $2.8\,\%$  in an area of excessive iodine intake<sup>(14)</sup>. In Denmark, formerly a region of mild-to-moderate ID (median urinary iodine concentration 61 µg/l), 5 years after the mandatory iodine fortification of salt, iodine status had significantly improved (median urinary iodine concentration 101 µg/l), but the prevalence of thyroid antibodies had risen, i.e. TPO-antibody >30 IU/ml increased from 14 to 24 % and Tg-antibody >20 IU/ml increased from 14 to 20 %<sup>(18)</sup>. However, despite the short-term adverse effects on thyroid autoimmunity, raising iodine intake from a deficient to an optimal intake-level ultimately results in the decreased prevalence of AITD; Denmark is an example of this<sup>(11,25)</sup>.

Potential mechanisms by which high or increased iodine intake raises autoimmune thyroid disease risk

The increase in circulating antibodies associated with iodine fortification is probably due to a number of factors including the strong immunogenicity of highly iodinated Tg which may trigger an immune reaction against the thyroid gland<sup>(25,26)</sup>. An additional factor may be that excess iodine intake increases the expression of the intercellular adhesion molecule-1, on the thyrocyte causing accelerated mononuclear cell infiltration and inflammation<sup>(26)</sup>. This has been demonstrated in the NOD.H2h4 mouse model of autoimmune thyroiditis where iodide treatment enhanced the transcription of intercellular adhesion molecule-1 triggered by reactive oxygen species and, in particular, by hydrogen peroxide generated in the thyrocyte for the organification of iodine (26,27). Other likely effects of high iodine intake in susceptible individuals are an increased production of thyroid-infiltrating T helper 17 cells, inhibition of T regulatory cell development and an abnormal expression of TNF-related apoptosis-inducing ligand in thyrocytes, resulting in apoptosis and tissue destruction<sup>(28)</sup>.

Iodine-intake recommendations to reduce autoimmune thyroid disease risk

With regard to autoimmune thyroiditis, as can be seen from the earlier section, there is more evidence for an association with iodine excess than with deficiency, especially in genetically susceptible individuals (14,28,29). It is therefore important to ensure, as far as possible, that iodine intake falls within the recommended levels<sup>(11)</sup> (see Table  $1^{(30-32)}$ ). On a population basis, this would be represented by a median urinary iodine concentration in adults of 100–200 µg/l. Authorities introducing iodine fortification of the food supply in a country (e.g. universal salt iodisation) need to ensure that such fortification is introduced very cautiously; Denmark provides an excellent example of how this can be done<sup>(33)</sup>. Individuals living in a country that does not have an iodine-fortified food supply who avoid the main food sources of iodine, i.e. milk and dairy products, seafood, most notably haddock, cod, crab, large/Dublin-bay prawns (often called scampi) and eggs, and do not use iodised salt, should be advised to take a daily supplement containing 140-150 µg iodine for thyroid protection, particularly if 36 M. P. Rayman

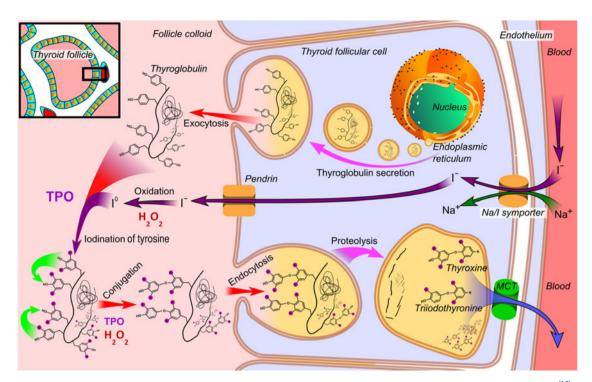


Fig. 1. (Colour online) Synthesis of the thyroid hormones in the thyroid follicle (modified from Häggström 2014<sup>(10)</sup>). Thyroglobulin is synthesised in the rough endoplasmic reticulum and follows the secretory pathway to enter the colloid in the lumen of the thyroid follicle by exocytosis. Meanwhile, a sodium-iodide (Na/I) symporter pumps iodide (I<sup>-</sup>) actively into the cell, which previously has crossed the endothelium by largely unknown mechanisms. This iodide enters the follicular lumen from the cytoplasm by the transporter pendrin, in a purportedly passive manner. In the colloid, iodide ( $I^-$ ) is oxidised to iodine ( $I^0$ ) by hydrogen peroxide ( $H_2O_2$ ) with the help of an enzyme called thyroid peroxidase (TPO). Iodine (I<sup>0</sup>) is very reactive and iodinates the thyroglobulin at tyrosyl residues in its protein chain (in total containing approximately 120 tyrosyl residues). In conjugation, adjacent tyrosyl residues are paired together, again under the influence of TPO and H<sub>2</sub>O<sub>2</sub>. The entire complex re-enters the follicular cell by endocytosis. Proteolysis by various proteases liberates thyroxine and triiodothyronine molecules, which enter the blood via a monocarboxylate transporter (MCT).

planning pregnancy (34-36). Although high in iodine, intake of brown seaweed (e.g. kelp/kombu), or brownseaweed supplements should be avoided as it may result in excessive intake<sup>(37)</sup>.

#### Iron

#### Role of iron in the thyroid

A haem-dependent enzyme, TPO, that has iron at its active centre, is required for thyroid hormone synthesis, as illustrated in Fig. 1<sup>(38,39)</sup>. TPO becomes active at the apical surface of thyrocytes only after it binds a prosthetic haem group<sup>(40)</sup>, hence an adequate iron status is required for the synthesis of thyroid hormones.

Co-morbidity of autoimmune thyroid disease and other autoimmune conditions

It is not always appreciated that ID is common in people with AITD owing to the frequent co-morbidity of other autoimmune conditions such as coeliac disease(41-43) and autoimmune gastritis (44–47) that often cause ID. Patients with subclinical hypothyroidism or HT

Table 1. lodine intake requirements by life stage according to various authorities

Age	EFSA AI (μg/d) <sup>(30)</sup>	USA RDA (μg/d) <sup>(31)</sup>	ICCIDD/UNICEF/ WHO RNI (μg/d) <sup>(32)</sup>
0-6 month	_	110 (AI)	90
7-12 month	70	130 (AI)	90
1–6 year	90	90	90
7-10 year	90	90-120	120
11-14 year	120	120-150	120-150
15-17 year	130	_	-
15-50 year	_	150	150
≥18 year	150	_	_
Pregnancy	200	220	250
Lactation	200	290	250

AI, adequate intake; RNI, recommended nutrient intake.

frequently have lower serum iron concentration and a higher prevalence of ID than do healthy controls (48,49). A symbiotic relationship exists between active thyroid hormone concentration and the formation of erythrocytes; T3 is needed to stimulate the proliferation of erythrocyte precursors, both directly and by enhancing the production of erythropoietin<sup>(50)</sup>.



#### Dependence of thyroid function on iron status

ID reduces thyroid hormone production by decreasing the activity of TPO (38–40). Evidence of the dependency of thyroid function on iron status comes from both animal and human studies. In rodents, ID, with or without anaemia, decreased serum T4 and T3 concentrations, lowered 5'-deiodinase (DIO) activity and reduced the ability to thermoregulate in response to a cold environment (39,51–53). In US women with mild ID anaemia (Hb, 110 g/l), serum T3 and T4 were significantly lower than in iron-sufficient controls<sup>(54)</sup>. Furthermore, ID predicts poor maternal thyroid status in pregnancy; in a study on 365 Swiss pregnant women in the second and third trimesters with borderline ID (median urinary iodine concentration 139 µg/l), concentrations of TSH, total T4 and urinary iodine were measured. Body iron stores, calculated from blood Hb concentration, mean corpuscular volume, serum ferritin, and transferrin receptor were highly significant predictors of TSH and total T4  $(P < 0.0001)^{(55)}$ . We also know that ID is associated with hypothyroxinaemia; serum free T4 concentrations were significantly lower in both 3340 pregnant and 1052 non-pregnant Chinese women with ID than in iron-adequate women<sup>(56)</sup>.

#### Effect of low iron stores on the efficacy of treatment for hypothyroidism

It is important to recognise that low iron stores may contribute to symptom persistence in patients treated for hypothyroidism in 5–10 % of whom symptoms remain despite being treated with levothyroxine (L-T4)<sup>(57)</sup>. An example is afforded by a small study in twenty-five Finnish women with persistent symptoms of hypothyroidism, despite appropriate L-T4 therapy, who became symptom-free when treated with oral iron supplements for 6–12 months<sup>(58)</sup>. None of the women had anaemia or erythrocyte indices outside the reference range although all had serum ferritin <60 µg/l. Restoration of serum ferritin above 100 µg/l ameliorated the symptoms in two-thirds of the women. At least 30–50 % of hypothyroid patients with persisting symptoms despite adequate L-T4 therapy may, in fact, have covert  $ID^{(58)}$ .

## Supplementation with thyroid hormone can improve iron status

An interesting fact is that supplementation with thyroid hormone in patients with subclinical hypothyroidism improves iron status. Early experiments in hypothyroid rats showed diminished gastrointestinal iron absorption that was restored to normal on supplementation with  $T3^{(59)}$ . In iron-deficient women with subclinical hypothyroidism treated for 1 year with T4, the frequency of anaemia decreased (P = 0.001) while ferritin, iron and Hb levels slightly increased (P > 0.05)<sup>(49)</sup>. In untreated women, a further decrease in ferritin level and increase in anaemia occurred<sup>(49)</sup>. In two randomised controlled trials (RCT) in patients with coexisting ID anaemia and sub-clinical hypothyroidism, treatment with iron and L-T4 together was considerably more effective in improving iron status than was treatment with iron alone<sup>(60,61)</sup>.

Recommendations for iron intake in thyroid patients

Patients with AITD or hypothyroidism should be routinely screened for ID. If either ID or serum ferritin below 70  $\mu g/l$  is found<sup>(58)</sup>, coeliac disease or autoimmune gastritis may be the cause and should be treated. Medication that reduces the acidity of stomach contents (e.g. proton pump inhibitors) may also cause reduced iron absorption<sup>(62)</sup>. If ID anaemia is present, haematological testing can be used to rule out the anaemia of chronic disease as the cause. In the absence of the latter, supplementation should be begun to restore iron sufficiency and prevent its deleterious effects on thyroid function<sup>(54,63)</sup>.

Once iron sufficiency is restored, assuming there is no underlying clinical cause of the deficiency, patients need to be told how to optimise their dietary iron intake. Foods with relatively high iron concentration include meat, fish, cereals, beans, nuts, egg yolks, dark green vegetables, potatoes and fortified foods<sup>(64)</sup>. However, iron is inefficiently absorbed, its bioavailability from different foods being markedly variable; bioavailability has been estimated to be in the range of 14-18 % for mixed diets and 5-12 % for vegetarian diets in individuals with no iron stores<sup>(65)</sup>. Absorption depends on a number of dietary and host-related factors; haem iron (from animal tissues) is considerably better absorbed than nonhaem iron, although the latter constitutes 90 % of the iron in a mixed diet. Dietary factors that reduce nonhaem iron absorption include phytate, polyphenols and calcium, while those that increase it include ascorbic acid and muscle tissue<sup>(65)</sup>. Following dietary advice, iron status should be checked regularly.

#### Selenium

Role of selenium in the thyroid: selenoproteins

The thyroid contains the highest concentration of selenium in the human body and is able to retain it even under conditions of severe deficiency<sup>(66)</sup>. A number of selenoproteins are expressed in thyrocytes<sup>(67)</sup>, those named later being particularly important to thyroid function.

The deiodinases. DIO1 and DIO2 can activate T4 by transforming it into T3 by removal of the 5'-iodine, while DIO1 and DIO3 can prevent T4 from being activated by converting it to the inactive reverse T3<sup>(68)</sup> (Fig. 2). DIO3 can also inactivate T3 by 5-deiodination to diiodothyronine. DIO2 is largely responsible for the local conversion of T4 to T3 in extrathyroidal target tissues<sup>(69)</sup>. A major role of DIO3 is to protect sensitive cells, such as fetal tissue, the placenta and central nervous system, from excessive concentrations of the active hormone, T3<sup>(69,70)</sup>.

The glutathione peroxidases. Extracellular glutathione peroxidase (GPX)3 is the only actively secreted GPX isozyme that is abundantly expressed in the thyroid gland<sup>(70)</sup>. It is secreted at the apical side of the thyrocyte membrane where it converts excess hydrogen peroxide that has not been used by TPO for the iodination of tyrosyl residues of Tg or for iodotyrosine coupling, into harmless water<sup>(71)</sup>.



Fig. 2. (Colour online) Action of the iodothyronine deiodinases, DIO1, DIO2 and DIO3, to produce the active and inactive forms of thyroid hormone.

Selenoprotein S. Selenoprotein S (SELENOS) is involved in the control of the inflammatory response in the endoplasmic reticulum by retrotranslocation of misfolded proteins from the endoplasmic reticulum lumen to the cytosol<sup>(72)</sup>. In a Portuguese study, the SELENOS – 105G/A promoter polymorphism (rs28665122) was strongly associated with circulating levels of cytokines such as IL-1β, IL-6, and TNF-α, known to be involved in the pathogenesis of HT<sup>(71,73)</sup>. Those with the SELENOS GA and AA genotypes were significantly more likely to have HT: OR (95 % CI) for HT was 2·22 (1·67, 2·95) and in male A-allele carriers, 3·94 (1·43, 10·84).

#### Effect of selenium status on thyroid disease

Selenium deficiency has been associated with a number of adverse thyroid conditions, including hypothyroidism, subclinical hypothyroidism, enlarged thyroid  $^{(74-77)}$ , thyroid cancer  $^{(67,77-79)}$  and AITD, including  $HT^{(75,77)}$  and  $GD^{(77,80)}$ .

A study of thyroid disease prevalence in more than 6000 people from two counties of Shaanxi Province, China, of very different selenium status, adequate and low, showed the protective effect of selenium adequacy<sup>(75)</sup>. Median (interquartile range) serum selenium concentration differed almost 2-fold (103-6 (79-7, 135-9) v. 57-4 (39-4, 82-1) µg/l; P = 0.001) between the two counties although iodine status was comparable<sup>(75)</sup>. After adjustment for potential confounders, the prevalence of pathological thyroid conditions was significantly lower in the adequate-selenium than in the lowselenium county (18·0 v. 30·5%; P < 0.001). Higher serum selenium was associated with significantly lower odds (OR (95 % CI)) of autoimmune thyroiditis (0.47 (0.35, 0.65)), hypothyroidism (0.75 (0.63, 0.90)), subclinical hypothyroidism (0.68 (0.58, 0.93)) and enlarged thyroid (0.75 (0.59, 0.97))<sup>(75)</sup>. The iodine intake was more-than-adequate<sup>(14,75,81)</sup> in both counties which may have accounted to some extent for the high prevalence of thyroid disease (15,82).

Selenium status has been found to be significantly lower in patients with GD than in normal controls in Danish<sup>(80)</sup> and Chinese studies<sup>(83)</sup>. In the latter, serum selenium was

negatively correlated with serum titre of TPO-antibodies (r = -0.161, P = 0.021), and Tg-antibodies  $(r = -0.237, P = 0.001)^{(83)}$ . In a prospective, case–control study in an Australian population, mean serum selenium decreased in parallel with increasing severity of Graves' orbitopathy: 94.0 (SD 15.8) µg/l in GD, 86.9 (SD 15.0) µg/l in moderate-to-severe Graves' orbitopathy and 86.1 (SD 13.4) µg/l in sight-threatening Graves' orbitopathy  $(P = 0.003)^{(84)}$ . However, these data may simply reflect the presence of inflammation in GD and more especially in Graves' orbitopathy; the expression of selenoproteins including plasma selenoprotein P is reduced by inflammatory cytokines resulting in a fall in plasma selenium (85,86).

## Randomised controlled trials of selenium in thyroid disease

Several trials of selenium supplementation have been carried out in AITD/HT and mild Graves' orbitopathy.

In a large, multicentre, RCT with selenium, patients with mild Graves' orbitopathy significantly improved on treatment with 100 µg selenium twice/d (as sodium selenite) for 6 months<sup>(7)</sup>. Patients on selenium treatment had improved quality of life (P < 0.001), less eye involvement (P = 0.01) and slower disease progression (P = 0.01). The benefit persisted at the 12-month follow-up. A protocol for an RCT of selenium in patients with Graves' hyperthyroidism (the GRASS trial) was published in 2013<sup>(87)</sup>. The primary outcome is the proportion of participants with anti-thyroid drug treatment failure at the end of the intervention period (24–30 months). Secondary outcomes include thyroid-specific quality of life and eye symptoms during the first year after randomisation<sup>(87)</sup>. The results of the trial have not yet been reported.

There have been a number of systematic reviews/ meta-analyses of controlled trials of selenium treatment in patients with AITD/HT<sup>(70,88–90)</sup>. The most recent is a 2016 meta-analysis of sixteen trials that found that selenium supplementation reduced serum TPO-antibodies levels after 3, 6 and 12 months in a population with chronic autoimmune thyroiditis treated with L-T4<sup>(88)</sup>. However, in an untreated autoimmune thyroiditis



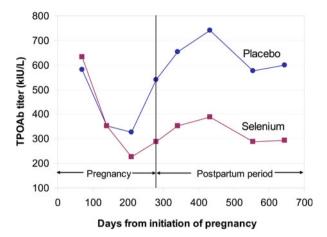
population, the effect was significant only after 3 months<sup>(88)</sup>. Some of these studies also saw a reduction in Tg-antibody titre at 12 months, an improvement in thyroid echogenicity and an increase in subjective wellbeing. Unfortunately, the methodology of many of the studies was flawed; underpowered, not double-blinded, not placebo-controlled and disparities in iodine intake were not considered (88,89,91). The beneficial effect in some studies and not in others cannot easily be explained on the basis of baseline selenium status, stage of disease, baseline TPO-antibody titres, form or dose of selenium used<sup>(4)</sup>. Later studies not included in these meta-analyses have been too small to contribute meaningful data<sup>(92,93)</sup>. Well designed, properly powered, RCT of selenium in the treatment of AITD/HT are therefore still needed before we can confidently recommend selenium supplementation in these patients. The protocol for a new, highquality, trial of selenium supplementation (Catalyst Trial) in patients with chronic autoimmune thyroiditis has been published<sup>(94)</sup>. We await the results with interest.

The presence of thyroid autoantibodies is relatively high in women of childbearing age<sup>(95)</sup>. One notable RCT has been carried out in pregnant women positive for TPO-antibodies. Up to 50 % of such women develop postpartum thyroiditis of whom 20-40 % subsequently become hypothyroid<sup>(96)</sup>. In an Italian study, 151 TPO-antibody positive women were randomly assigned supplementation with 200 µg selenium/d (as selenomethionine) or placebo during pregnancy and the postpartum period<sup>(97)</sup>. TPO-antibodies fell significantly during gestation in both groups but the reduction was significantly greater in the selenium-supplemented group (P = 0.01) and remained so in the postpartum period (P = 0.01) (see Fig. 3). Compared with women on placebo, those on selenium had a significantly lower incidence of post-partum thyroid disease (28.6 v. 48.6 %; P <0.01) and permanent hypothyroidism (11.7 v. 20.3 %; P < 0.01). In contrast to women on placebo, ultra-sound echogenicity did not fall in those supplemented with selenium. At the end of the postpartum period, grade 2-3 thyroiditis had developed in 44.3% of women on placebo but only in 27.3% of women on selenium  $(P < 0.01)^{(97)}$ .

The only other RCT that investigated the effect of selenium supplementation on AITD in pregnancy found no difference in the magnitude of TPO-antibody decrease between selenium and placebo groups (98). However the median baseline TPO-antibody concentrations in the women were much lower than in the earlier study, the selenium dose was less than one third as high (60  $\mu$ g/d) and the trial was not adequately powered (98). Clearly, there is a need for a further, high-quality, adequately powered RCT in the TPO-antibody-positive pregnant population to see if the results of the Italian study can be replicated (97).

Is selenium intake adequate to reduce the risk of thyroid disease?

Selenium intake differs vastly from one part of the world to another owing to differences in the selenium content of the soil on which crops and fodder are grown, selenium speciation, soil pH and organic-matter content (99). Intake



**Fig. 3.** (Colour online) Selenium protects against postpartum autoimmune thyroid disease (adapted from Negro *et al.*<sup>(97)</sup> with permission 0. TPOAb, thyroid peroxidase antibodies.

ranges from deficient  $(7 \mu g/d)$  to toxic  $(4990 \mu g/d)$  as shown in Fig. 4<sup>(100,101)</sup>. A vertical band in Fig. 4 shows the level of intake believed to be needed to optimise the activity of GPX3<sup>(101)</sup>, the main selenoenzyme that removes excess hydrogen peroxide from the thyroid. It is clear that the mean intake in many countries, notably those in Europe, does not achieve that level.

#### Recommendations for selenium intake

Although we lack evidence that selenium supplementation results in clinical improvement in autoimmune thyroiditis (other than in mild Graves' orbitopathy), it still makes sense to ensure that selenium intake is adequate, given the roles played by selenoproteins in human health (100) and particularly in the thyroid (70,71). Regions of deficient, more-than-adequate or high iodine intake may have more need for selenium owing to the capacity of selenoproteins to protect the thyroid from excessive hydrogen peroxide<sup>(71)</sup> and from inflammation<sup>(71)</sup> (see earlier). Hence, in such locations, clinicians need to ensure that selenium intake/ status is adequate. Women are at greater risk of thyroid disorders and may thus have a higher requirement for additional selenium, particularly in pregnancy. Geographical location will give a good indication of selenium adequacy or otherwise (see Fig. 4).

It is also important to enquire into the dietary habits of a given patient and see if he/she eats foods that supply selenium<sup>(100)</sup>. Although Brazil nuts are the richest selenium food source they cannot be recommended as a main source as the content is very variable, ranging from 0.03 to 512 mg/kg fresh weight, and they are high in barium, which can be toxic (102). Otherwise, organ meats and seafoods are the best sources, followed by muscle meats, cereals and grains, although the selenium content of the latter varies widely with location, being towards or at the bottom of the range in the UK and Europe but at the top in North America, most notably 5)(100,102) Fig. In Canada (see China. selenium-enriched tea is an option<sup>(75)</sup>. Given the sources described, vegans and vegetarians in the West are particularly at risk of inadequate selenium intake.



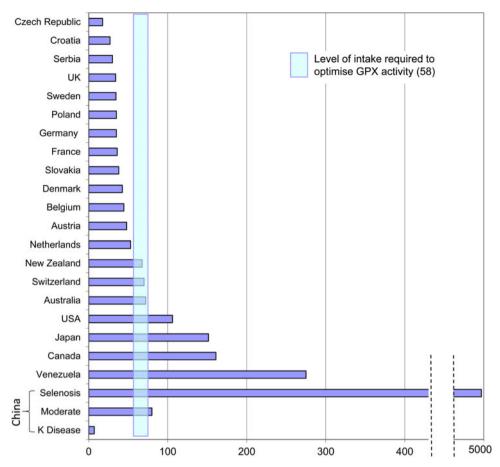


Fig. 4. (Colour online) Mean selenium intake levels (µg/d) in different countries and the range of selenium intake (55-75 µg/d) believed to be required for optimal activity of plasma glutathione peroxidase (GPX3) (adapted from Rayman<sup>(101)</sup>).

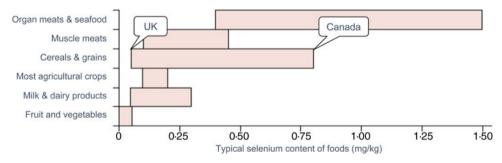


Fig. 5. (Colour online) Typical selenium content of food sources, adapted from WHO. Selenium. A report of the International Programme on Chemical Safety. Environmental Health Criteria number 58. Geneva: WHO, 1987 (reproduced from Rayman<sup>(100)</sup>).

If a patient's diet in a country of low-to-moderate selenium intake contains few, or no, selenium-rich sources, low-dose selenium supplementation can be advised although no more than 50-100 µg selenium/d is advisable. Multi-vitamin/mineral tablets may contain 50 µg selenium, a daily amount that will generally be adequate, particularly for women. A dose of 100 µg selenium/d (as selenium-yeast) given to someone in the UK will raise plasma selenium to about 140 µg/l which is more than enough to optimise the synthesis of all the selenoproteins<sup>(103)</sup>. Either selenium-yeast (which behaves in the body such as wheat-selenium) or sodium selenite which the body can readily use for selenoprotein synthesis without increasing the build-up of selenium, will  $do^{(102)}$ 

Clinicians should be aware that even if a hypothyroid patient is being treated with L-T4, a number of studies have found that giving selenium as well as L-T4 resulted in a greater reduction in TPO-antibodies, inflammatory cytokines and C-reactive protein<sup>(104,105)</sup>.



Although selenium is essential, excessive intake is toxic; supplements of selenium of 200 µg/d (as selenium yeast or selenomethionine), generally considered to be safe, have been associated with toxic effects (alopecia, dermatitis, squamous cell carcinoma, type-2 diabetes mellitus, high-grade prostate cancer) in North Americans (106-108). If selenium concentration reaches or exceeds 122 µg/l in plasma, as in the top tertile of the Nutritional Prevention of Cancer trial (109) or is already 137 µg/l in serum, as in SELECT<sup>(106)</sup>, supplementation should be avoided<sup>(106–108,110)</sup>. Furthermore, mortality was found to be increased in a European population of relatively low selenium status (plasma selenium 89 µg/l) on long-term supplementation with 300 µg/d (as selenium-yeast)(111). As for many nutrients, there is a U-shaped relationship between selenium status and risk of a number of adverse conditions (100). The aim should, therefore, be to have an intake sufficient to reduce the risk of thyroid disease without risking toxicity<sup>(100)</sup>.

#### Conclusion

As explained in detail earlier, the appropriate status of iodine, iron and selenium is crucial to thyroid health. Nutritional status of these micronutrients is frequently inadequate: iodine status is often low in countries without iodine fortification of the food supply; selenium status is generally fairly poor in Europe and many parts of China; iron status is frequently low in women of child-bearing age, particularly towards the end of pregnancy. Clinicians need to be aware of these dietary risk factors and to treat thyroid patients accordingly.

#### Acknowledgement

I thank Shiqian Hu, Department of Endocrinology, First Affiliated Hospital of Xi'an Jiaotong University, Xi'an, Shaanxi, China, who worked with me on investigating the effect of micronutrients on Hashimoto's thyroiditis when on a placement at the University of Surrrey.

#### **Financial Support**

None.

#### **Conflicts of Interest**

None.

#### **Authorship**

M. P. R. is the sole author of the review.

#### References

- 1. McLeod DS & Cooper DS (2012) The incidence and prevalence of thyroid autoimmunity. *Endocrine* **42**, 252–265.
- Okayasu I, Hara Y, Nakamura K et al. (1994) Racial and age-related differences in incidence and severity of focal autoimmune thyroiditis. Am J Clin Pathol 101, 698–702.
- 3. Hashimoto H (1912) Zur Kenntnis der lymphomatösen Veränderung der Schilddrüse (Struma lymphomatosa). *Archiv für Klinische Chirurgie (in German)* **97**, 219–248.
- 4. Effraimidis G & Wiersinga WM (2014) Mechanisms in endocrinology: autoimmune thyroid disease: old and new players. *Eur J Endocrinol* **170**, R241–R252.
- American Thyroid Association Hashimoto's thyroiditis. https://www.thyroid.org/hashimotos-thyroiditis/ (accessed June 2018).
- 6. Nacamulli D, Petricca D & Mian C (2013) Selenium and autoimmune thyroiditis. *J Endocrinol Invest* **36**, 8–14.
- Marcocci C, Kahaly GJ, Krassas GE et al. (2011) Selenium and the course of mild Graves' orbitopathy. N Engl J Med 364, 1920–1931.
- 8. Brent GA (2008) Clinical practice. Graves' disease. *N Engl J Med* **358**, 2594–2605.
- Hu S & Rayman MP (2017) Multiple nutritional factors and the risk of Hashimoto's thyroiditis. *Thyroid* 27, 597– 610
- Häggström M (2014) Synthesis of the thyroid hormones in the thyroid follicle, Medical gallery of Mikael Häggström 2014. Wiki Journal of Medicine 1.
- 11. Laurberg P, Cerqueira C, Ovesen L *et al.* (2010) Iodine intake as a determinant of thyroid disorders in populations. *Best Practice Res Clinical Endocrinol Metab* **24**, 13–27.
- 12. Bülow Pedersen ILP (2009) Antibodies to thyroid peroxidase and thyroglobulin in iodine deficiencies. In *Comprehensive Handbook of Iodine: Nutritional, Biochemical and Pathological Aspects*, pp. 575–585 [Preedy VR Burrow GN, Watson RR, editors]: Burlington, MA: Elsevier.
- 13. Pedersen IB, Knudsen N, Jorgensen T *et al.* (2003) Thyroid peroxidase and thyroglobulin autoantibodies in a large survey of populations with mild and moderate iodine deficiency. *Clin Endocrinol* (*Oxf*) **58**, 36–42.
- Teng W, Shan Z, Teng X et al. (2006) Effect of iodine intake on thyroid diseases in China. N Engl J Med 354, 2783–2793.
- 15. Teng X, Shan Z, Chen Y *et al.* (2011) More than adequate iodine intake may increase subclinical hypothyroidism and autoimmune thyroiditis: a cross-sectional study based on two Chinese communities with different iodine intake levels. *Eur J Endocrinol* **164**, 943–950.
- Peng NC, Shi LX, Zhang Q et al. (2013) An epidemiological survey of the prevalence of thyroid diseases in mild iodine deficiency city after salt iodization. Zhonghua Nei Ke Za Zhi 52, 16–20.
- 17. Zhang JY, Li SM, Leng JL *et al.* (2013) Changes of the spectrum on thyroid disease after the 10-year implementation of universal salt iodization in Guangxi Zhuang Autonomous Region. *Zhonghua liu xing bing xue za zhi* = *Zhonghua liuxingbingxue zazhi* 34, 970–974.
- 18. Pedersen IB, Knudsen N, Carle A *et al.* (2011) A cautious iodization programme bringing iodine intake to a low recommended level is associated with an increase in the prevalence of thyroid autoantibodies in the population. *Clin Endocrinol* (*Oxf*) **75**, 120–126.
- 19. Bjergved L, Jorgensen T, Perrild H *et al.* (2012) Predictors of change in serum TSH after iodine fortification: an 11-year follow-up to the DanThyr study. *J Clin Endocrinol Metab* **97**, 4022–4029.



- 20. Cerqueira C, Knudsen N, Ovesen L et al. (2011) Doubling in the use of thyroid hormone replacement therapy in Denmark: association to iodization of salt? Eur J Epidemiol 26, 629-635.
- 21. Camargo RY, Tomimori EK, Neves SC et al. (2008) Thyroid and the environment: exposure to excessive nutritional iodine increases the prevalence of thyroid disorders in Sao Paulo, Brazil. Eur J Endocrinol 159, 293-299.
- 22. Zaletel K, Gaberscek S & Pirnat E (2011) Ten-year follow-up of thyroid epidemiology in Slovenia after increase in salt iodization. Croat Med J 52, 615-621.
- 23. Fernando RF, Chandrasinghe PC & Pathmeswaran AA (2012) The prevalence of autoimmune thyroiditis after universal salt iodisation in Sri Lanka. Ceylon Med J 57, 116-119.
- 24. Aghini Lombardi F, Fiore E, Tonacchera M et al. (2013) The effect of voluntary iodine prophylaxis in a small rural community: the Pescopagano survey 15 years later. J Clin Endocrinol Metab 98, 1031–1039.
- 25. Zimmermann MB & Boelaert K (2015) Iodine deficiency and thyroid disorders. Lancet Diabetes Endocrinol 3, 286–295.
- 26. Burek CL & Talor MV (2009) Environmental triggers of autoimmune thyroiditis. J Autoimmun 33, 183–189.
- 27. Sharma R, Traore K, Trush MA et al. (2008) Intracellular adhesion molecule-1 up-regulation on thyrocytes by iodine of non-obese diabetic.H2(h4) mice is reactive oxygen species-dependent. Clin Exp Immunol 152, 13-20.
- 28. Duntas LH (2015) The role of iodine and selenium in autoimmune thyroiditis. Hormone and Metabolic Research = Hormon- und Stoffwechselforschung = Hormones et metabolisme 47, 721-726.
- 29. Luo Y, Kawashima A, Ishido Y et al. (2014) Iodine excess as an environmental risk factor for autoimmune thyroid disease. Int J Mol Sci 15, 12895–12912.
- 30. EFSA Panel on Dietetic Products NaAN (2014) Scientific Opinion on Panel on Micronutrients: Dietary Reference Values for iodine. EFSA Journal 12, 3660 [3657 pp]. https://www.efsa.europa.eu/en/efsajournal/pub/3660 (accessed May 2018).
- 31. Institute of Medicine (2001) Panel on Micronutrients: Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic, Boron, Chromium, Copper, Iodine, Iron, Manganese, Molybdenum, Nickel, Silicon, Vanadium, and Zinc. Washington, DC: National Academies Press (US).
- 32. WHO/UNICEF/ICCIDD (2007) Assessment of Iodine Deficiency Disorders and Monitoring their Elimination: A Guide for Programme Managers, 3rd ed. Geneva: World Health Organization.
- 33. Rasmussen LB, Carle A, Jorgensen T et al. (2008) Iodine intake before and after mandatory iodization in Denmark: results from the Danish Investigation of Iodine Intake and Thyroid Diseases (DanThyr) study. Br J Nutr 100, 166-173.
- 34. McCance and Widdowson's 'composition of foods integrated dataset (25 March 2015): Public Health England. https://www.gov.uk/government/publications/compositionof-foods-integrated-dataset-cofid (accessed May 2018).
- 35. Bath SC & Rayman MP (2016) Iodine Food Fact Sheet: British Dietetic Association (BDA). https://www.bda.uk. com/foodfacts/Iodine.pdf (accessed May 2018).
- 36. NIH Office of dietary supplements Iodine Fact Sheet for Health Professionals. https://ods.od.nih.gov/factsheets/ Iodine-HealthProfessional/#h3 (accessed May 2018).
- 37. Yeh T, Hung N & Lin T (2014) Analysis of iodine content in seaweed by GC-ECD and estimation of iodine intake. J Food Drug Anal 22, 189-196.
- 38. Dunn JT & Dunn AD (2001) Update on intrathyroidal iodine metabolism. Thyroid 11, 407-414.

- 39. Hess SY, Zimmermann MB, Arnold M et al. (2002) Iron deficiency anemia reduces thyroid peroxidase activity in rats. J Nutr 132, 1951-1955.
- 40. Favadat L. Niccoli-Sire P. Lanet J et al. (1999) Role of heme in intracellular trafficking of thyroperoxidase and involvement of H2O2 generated at the apical surface of thyroid cells in autocatalytic covalent heme binding. J Biol Chem 274, 10533-10538.
- 41. Sategna-Guidetti C, Bruno M, Mazza E et al. (1998) Autoimmune thyroid diseases and coeliac disease. Eur J Gastroenterol Hepatol 10, 927-931.
- 42. Fisher AH, Lomasky SJ, Fisher MJ et al. (2008) Celiac disease and the endocrinologist: a diagnostic opportunity. Endocr Pract 14, 381-388.
- 43. Pinto-Sanchez MI, Bercik P, Verdu EF et al. (2015) Extraintestinal manifestations of celiac disease. Dig Dis 33, 147-154.
- 44. Centanni M, Marignani M, Gargano L et al. (1999) Atrophic body gastritis in patients with autoimmune thyroid disease: an underdiagnosed association. Arch Intern Med 159, 1726-1730.
- 45. Checchi S, Montanaro A, Ciuoli C et al. (2010) Prevalence of parietal cell antibodies in a large cohort of patients with autoimmune thyroiditis. Thyroid: Official J Am Thyroid Assoc 20, 1385–1389.
- 46. Lahner E, Centanni M, Agnello G et al. (2008) Occurrence and risk factors for autoimmune thyroid disease in patients with atrophic body gastritis. Am J Med 121, 136–141.
- 47. Tozzoli R, Kodermaz G, Perosa AR et al. (2010) Autoantibodies to parietal cells as predictors of atrophic body gastritis: a 5-year prospective study in patients with autoimmune thyroid diseases. Autoimmun Rev 10, 80-83.
- 48. Erdal M, Sahin M, Hasimi A et al. (2008) Trace element levels in Hashimoto thyroiditis patients with subclinical hypothyroidism. Biol Trace Elem Res 123, 1–7.
- Nekrasova TA, Strongin LG & Ledentsova OV (2013) Hematological disturbances in subclinical hypothyroidism and their dynamics during substitution therapy. Klin Med (Mosk) 91, 29-33.
- 50. Szczepanek-Parulska E, Hernik A & Ruchala M (2017) Anemia in thyroid diseases. Polish Archives Internal Med **127**, 352–360.
- 51. Beard J, Tobin B & Green W (1989) Evidence for thyroid hormone deficiency in iron-deficient anemic rats. J Nutr 119, 772-778.
- 52. Beard JL, Brigham DE, Kelley SK et al. (1998) Plasma thyroid hormone kinetics are altered in iron-deficient rats. J Nutr 128, 1401–1408.
- 53. Beard J, Finch CA & Green WL (1982) Interactions of iron deficiency, anemia, and thyroid hormone levels in response of rats to cold exposure. Life Sci 30, 691-697.
- 54. Beard JL, Borel MJ & Derr J (1990) Impaired thermoregulation and thyroid function in iron-deficiency anemia. Am J Clin Nutr 52, 813-819.
- 55. Zimmermann MB, Burgi H & Hurrell RF (2007) Iron deficiency predicts poor maternal thyroid status during pregnancy. J Clin Endocrinol Metab 92, 3436-3440.
- 56. Yu X, Shan Z, Li C et al. (2015) Iron deficiency, an independent risk factor for isolated hypothyroxinemia in pregnant and nonpregnant women of childbearing age in China. J Clin Endocrinol Metab 100, 1594-1601.
- 57. Wiersinga WM, Duntas L, Fadeyev V et al. (2012) 2012 ETA guidelines: the use of L-T4 + L-T3 in the treatment of hypothyroidism. Eur Thyroid J 1, 55-71.
- 58. Soppi E (2015) Iron deficiency is the main cause of symptom persistence in patients treated for hypothyroidism. In 15th International Thyroid Congress, Thyroid 25, A-74.



- Donati RM, Fletcher JW, Warnecke MA et al. (1973) Erythropoiesis in hypothyroidism. Proc Soc Exp Biol Med 144, 78–82.
- Ravanbod M, Asadipooya K, Kalantarhormozi M et al. (2013) Treatment of iron-deficiency anemia in patients with subclinical hypothyroidism. Am J Med 126, 420–424.
- 61. Cinemre H, Bilir C, Gokosmanoglu F *et al.* (2009) Hematologic effects of levothyroxine in iron-deficient subclinical hypothyroid patients: a randomized, double-blind, controlled study. *J Clin Endocrinol Metab* **94**, 151–156.
- 62. Ajmera AV, Shastri GS, Gajera MJ *et al.* (2012) Suboptimal response to ferrous sulfate in iron-deficient patients taking omeprazole. *Am J Ther* **19**. 185–189.
- Martinez-Torres C, Cubeddu L, Dillmann E et al. (1984)
   Effect of exposure to low temperature on normal and irondeficient subjects. Am J Physiol 246, R380–R383.
- 64. EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA) (2015) Scientific opinion on dietary reference values for iron. *EFSA J* 13, 4254. https://doi.org/4210·2903/j.efsa.2015·4254 (accessed May 2018).
- 65. Hurrell R & Egli I (2010) Iron bioavailability and dietary reference values. *Am J Clin Nutr* **91**, 1461s-1467s.
- Kohrle J (2013) Selenium and the thyroid. Curr Opin Endocrinol Diabetes Obes 20, 441–448.
- 67. Schmutzler C, Mentrup B, Schomburg L *et al.* (2007) Selenoproteins of the thyroid gland: expression, localization and possible function of glutathione peroxidase 3. *Biol Chem* **388**, 1053–1059.
- Darras VM & Van Herck SL (2012) Iodothyronine deiodinase structure and function: from ascidians to humans. J Endocrinol 215, 189–206.
- Kohrle J, Jakob F, Contempre B et al. (2005) Selenium, the thyroid, and the endocrine system. Endocr Rev 26, 944–984.
- Schomburg L (2011) Selenium, selenoproteins and the thyroid gland: interactions in health and disease. *Nat Rev* Endocrinol 8, 160–171.
- Schomburg L & Kohrle J (2008) On the importance of selenium and iodine metabolism for thyroid hormone biosynthesis and human health. *Mol Nutr Food Res* 52, 1235–1246.
- Curran JE, Jowett JB, Elliott KS et al. (2005) Genetic variation in selenoprotein S influences inflammatory response. Nat Genet 37, 1234–1241.
- Santos LR, Duraes C, Mendes A et al. (2014) A polymorphism in the promoter region of the selenoprotein S gene (SEPS1) contributes to Hashimoto's thyroiditis susceptibility. J Clin Endocrinol Metab 99, E719–E723.
- Derumeaux H, Valeix P, Castetbon K et al. (2003)
   Association of selenium with thyroid volume and echostructure in 35- to 60-year-old French adults. Eur J Endocrinol 148, 309–315.
- Wu Q, Rayman MP, Lv H et al. (2015) Low population selenium status is associated with increased prevalence of thyroid disease. J Clin Endocrinol Metab 100, 4037–4047.
- Rasmussen LB, Schomburg L, Kohrle J et al. (2011) Selenium status, thyroid volume, and multiple nodule formation in an area with mild iodine deficiency. Eur J Endocrinol 164, 585–590.
- 77. Rayman MP & Duntas L (2018) Selenium deficiency and thyroid disease. In *The Thyroid and Its Diseases* [M Luster and L Duntas and L Wartofsky, editors]: New York: Springer.
- 78. Glattre E, Thomassen Y, Thoresen SO *et al.* (1989) Prediagnostic serum selenium in a case-control study of thyroid cancer. *Int J Epidemiol* **18**, 45–49.
- 79. Lin JC, Kuo WR, Chiang FY *et al.* (2009) Glutathione peroxidase 3 gene polymorphisms and risk of differentiated thyroid cancer. *Surgery* **145**, 508–513.

- 80. Bulow Pedersen I, Knudsen N, Carle A *et al.* (2013) Serum selenium is low in newly diagnosed Graves' disease: a population-based study. *Clin Endocrinol* (*Oxf*) **79**, 584–590.
- 81. WHO/UNICEF/ICCIDD (2007) Assessment of Iodine Deficiency Disorders and Monitoring their Elimination, 3rd ed. Geneva: WHO.
- 82. Teng X, Shi X, Shan Z *et al.* (2008) Safe range of iodine intake levels: a comparative study of thyroid diseases in three women population cohorts with slightly different iodine intake levels. *Biol Trace Elem Res* **121**, 23–30.
- 83. Liu Y, Liu S, Mao J *et al.* (2018) Serum trace elements profile in graves' disease patients with or without orbitopathy in Northeast China. *BioMed Res Int* **2018**, 3029379.
- 84. Khong JJ, Goldstein RF, Sanders KM *et al.* (2014) Serum selenium status in graves' disease with and without orbitopathy: a case-control study. *Clin Endocrinol* (*Oxf*) **80**, 905–910.
- 85. Hesse-Bahr K, Dreher I & Kohrle J (2000) The influence of the cytokines Il-1beta and INFgamma on the expression of selenoproteins in the human hepatocarcinoma cell line HepG2. *Biofactors* 11, 83–85.
- 86. Nichol C, Herdman J, Sattar N *et al.* (1998) Changes in the concentrations of plasma selenium and selenoproteins after minor elective surgery: further evidence for a negative acute phase response? *Clin Chem* **44**, 1764–1766.
- 87. Watt T, Cramon P, Bjorner JB et al. (2013) Selenium supplementation for patients with Graves' hyperthyroidism (the GRASS trial): study protocol for a randomized controlled trial. *Trials* 14, 119.
- 88. Wichman JWK, Bonnema SJ & Hegedus L (2016) Selenium supplementation significantly reduces thyroid autoantibody levels in patients with chronic autoimmune thyroiditis: a systematic review and meta-analysis. *Thyroid* **26**, 1081–1092.
- 89. van Zuuren EJ, Albusta AY, Fedorowicz Z *et al.* (2013) Selenium supplementation for Hashimoto's thyroiditis. *Cochrane Database Syst Rev* CD010223.
- 90. Fan Y, Xu S, Zhang H *et al.* (2014) Selenium supplementation for autoimmune thyroiditis: a systematic review and meta-analysis. *Int J Endocrinol* **2014**, 904573.
- 91. Duntas LH & Benvenga S (2015) Selenium: an element for life. *Endocrine* **48**, 756–775.
- 92. Pirola I, Gandossi E, Agosti B *et al.* (2016) Selenium supplementation could restore euthyroidism in subclinical hypothyroid patients with autoimmune thyroiditis. *Endokrynol Pol* **67**, 567–571.
- 93. de Farias CR, Cardoso BR, de Oliveira GM *et al.* (2015) A randomized-controlled, double-blind study of the impact of selenium supplementation on thyroid autoimmunity and inflammation with focus on the GPx1 genotypes. *J Endocrinol Invest* 38, 1065–1074.
- 94. Winther KH, Watt T, Bjorner JB *et al.* (2014) The chronic autoimmune thyroiditis quality of life selenium trial (CATALYST): study protocol for a randomized controlled trial. *Trials* **15**, 115.
- 95. Mehran L, Amouzegar A, Delshad H *et al.* (2013) Trimester-specific reference ranges for thyroid hormones in Iranian pregnant women. *J Thyroid Res* **2013**, 651517.
- 96. Stagnaro-Green A (2012) Approach to the patient with postpartum thyroiditis. *J Clin Endocrinol Metab* **97**, 334–342.
- 97. Negro R, Greco G, Mangieri T *et al.* (2007) The influence of selenium supplementation on postpartum thyroid status in pregnant women with thyroid peroxidase autoantibodies. *J Clin Endocrinol Metab* **92**, 1263–1268.
- 98. Mao J, Pop VJ, Bath SC *et al.* (2016) Effect of low-dose selenium on thyroid autoimmunity and thyroid function in



- UK pregnant women with mild-to-moderate iodine deficiency. Eur J Nutr 55, 55-61.
- 99. Johnson CC, Fordyce FM & Rayman MP (2010) Symposium on 'Geographical and geological influences on nutrition': factors controlling the distribution of selenium in the environment and their impact on health and nutrition. Proc Nutr Soc 69, 119-132.
- 100. Rayman MP (2012) Selenium and human health. Lancet (London, England) 379, 1256-1268.
- 101. Rayman MP (2005) Selenium in cancer prevention: a review of the evidence and mechanism of action. Proc Nutr Soc 64, 527-542.
- 102. Rayman MP (2008) Food-chain selenium and human health: emphasis on intake. Br J Nutr 100, 254-268.
- 103. Rayman MP, Thompson AJ, Bekaert B et al. (2008) Randomized controlled trial of the effect of selenium supplementation on thyroid function in the elderly in the United Kingdom. Am J Clin Nutr 87, 370-378.
- 104. Krysiak R & Okopien B (2011) The effect of levothyroxine and selenomethionine on lymphocyte and monocyte cytokine release in women with Hashimoto's thyroiditis. J Clin Endocrinol Metab 96, 2206-2215.
- 105. Duntas LH, Mantzou E & Koutras DA (2003) Effects of a 6 month treatment with selenomethionine in patients with autoimmune thyroiditis. Eur J Endocrinol 148, 389-393.

- 106. Lippman SM, Klein EA, Goodman PJ et al. (2009) Effect of selenium and vitamin E on risk of prostate cancer and other cancers: the Selenium and Vitamin E Cancer Prevention Trial (SELECT). JAMA 301, 39-51.
- 107. Duffield-Lillico AJ, Slate EH, Reid ME et al. (2003) Selenium supplementation and secondary prevention of nonmelanoma skin cancer in a randomized trial. J Natl Cancer Inst 95, 1477-1481.
- 108. Stranges S, Marshall JR, Natarajan R et al. (2007) Effects of long-term selenium supplementation on the incidence of type 2 diabetes: a randomized trial. Ann Intern Med **147**. 217–223.
- 109. Duffield-Lillico AJ. Reid ME. Turnbull BW et al. (2002) Baseline characteristics and the effect of selenium supplementation on cancer incidence in a randomized clinical trial: a summary report of the Nutritional Prevention of Cancer Trial. Cancer Epidemiol Biomarkers Prev 11, 630-639.
- 110. Kristal AR, Darke AK, Morris JS et al. (2014) Baseline selenium status and effects of selenium and vitamin e supplementation on prostate cancer risk. J Natl Cancer Inst 106, dit456.
- 111. Rayman MP, Winther KH, Pastor-Barriuso R et al. (2018) Effect of long-term selenium supplementation on mortality: results from a multiple-dose, randomised controlled trial. Free Radic Biol Med [Epublication ahead of print versionl.

